

High-throughput, High-resolution HLA Genotyping

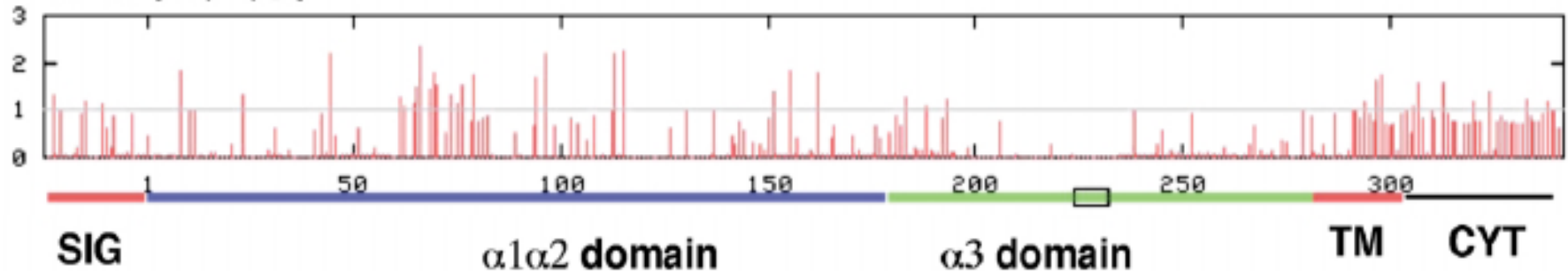
Chunlin Wang Ph.D.

Stanford Genome Technology Center

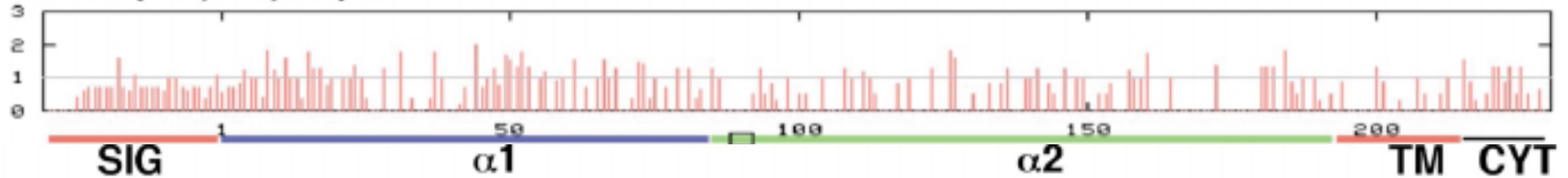
Stanford University

HLA Gene Structure and Diversity

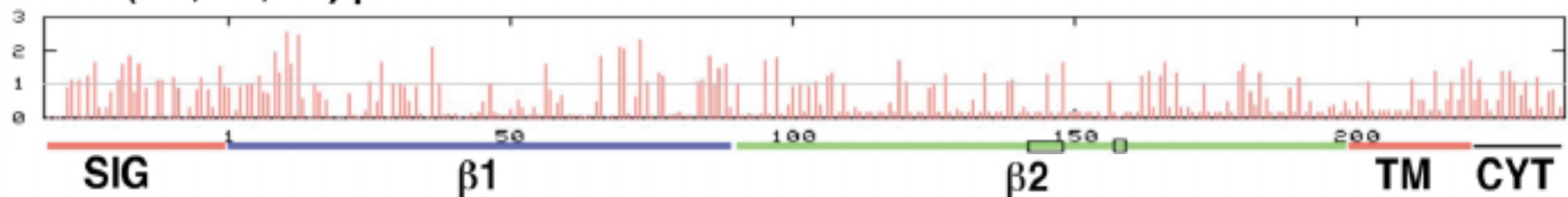
A HLA-(A,B,C)



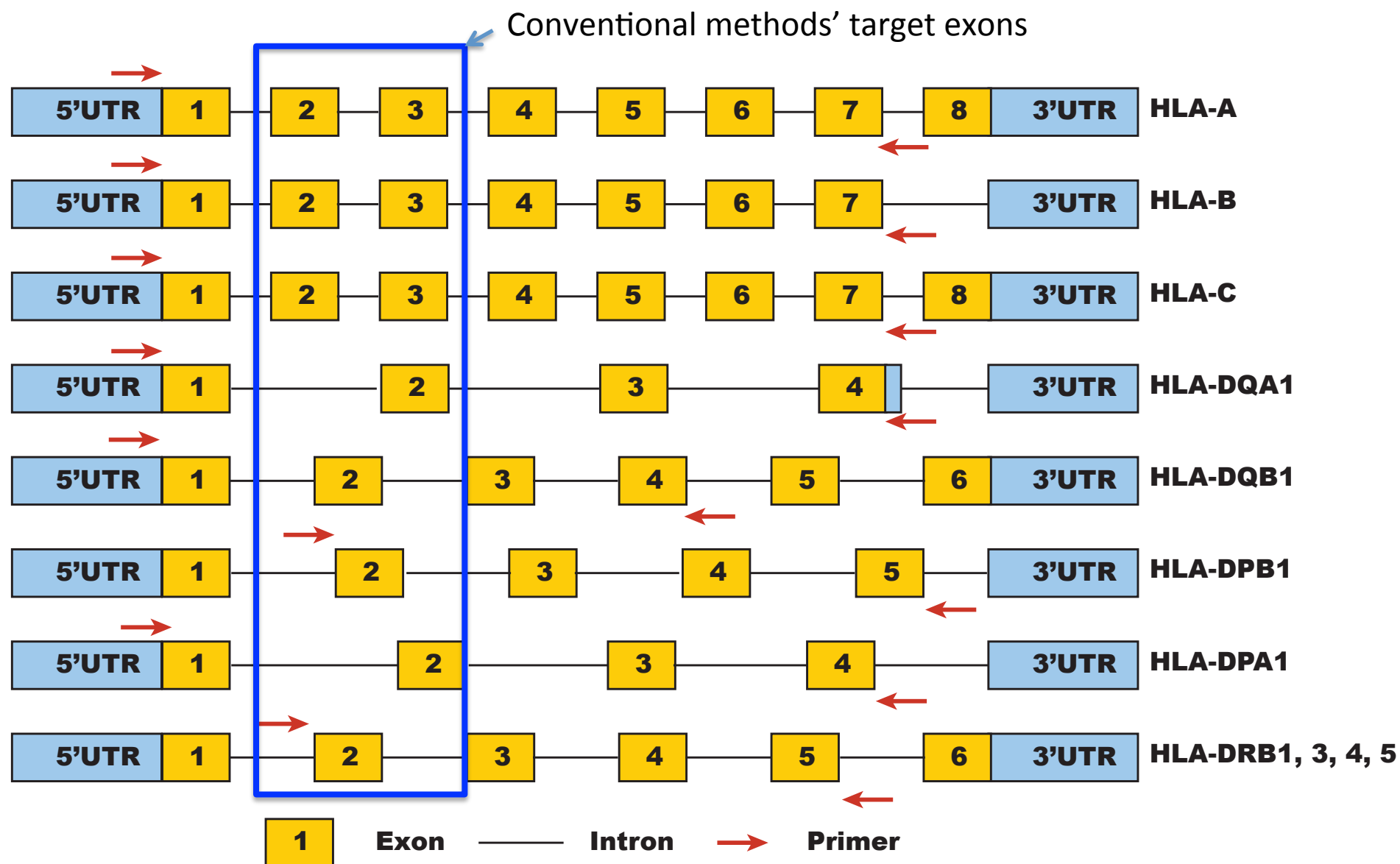
A HLA-(DP,DQ,DR) α



E HLA-(DP,DQ,DR) β

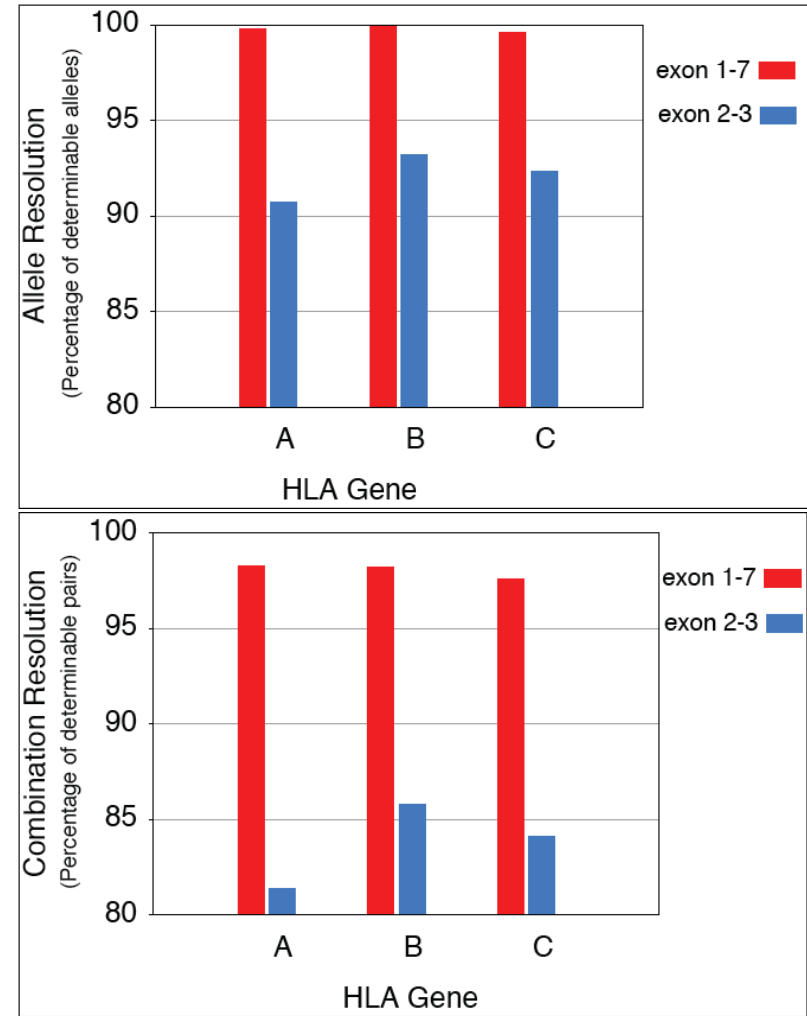


Our Strategy



Advantage of Our Strategy

- Sequencing more exons and introns significantly increases both allele resolution and combination resolution.
- With more high-quality refer sequences, the resolution of our method increases



Sample Preparation and Sequencing

Long Range PCR (LR-PCR)

Each locus is amplified by LR-PCR covering the major, most polymorphic coding region.



Random Fragmentation

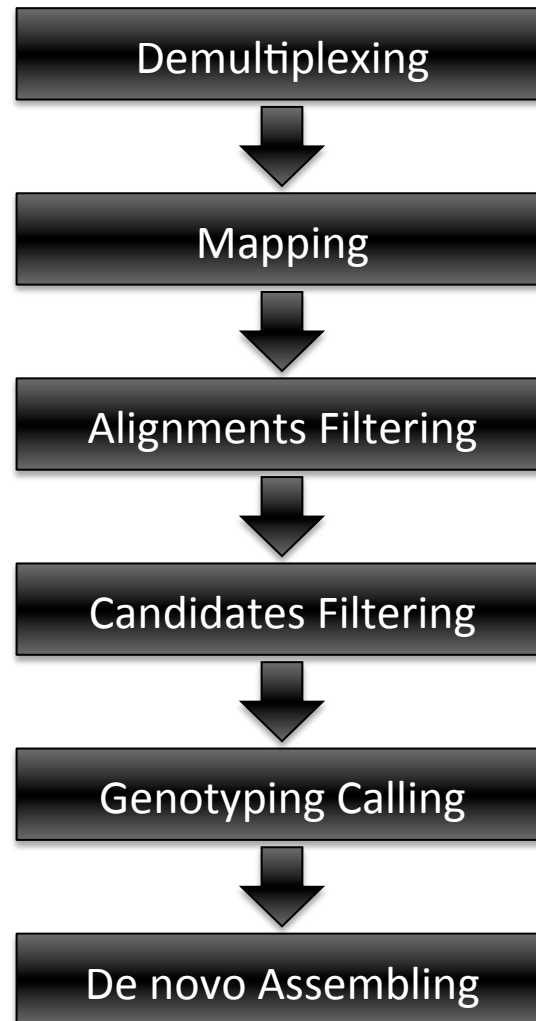
Amplification products of each sample are pooled, ligated together, fragmented through sonication.



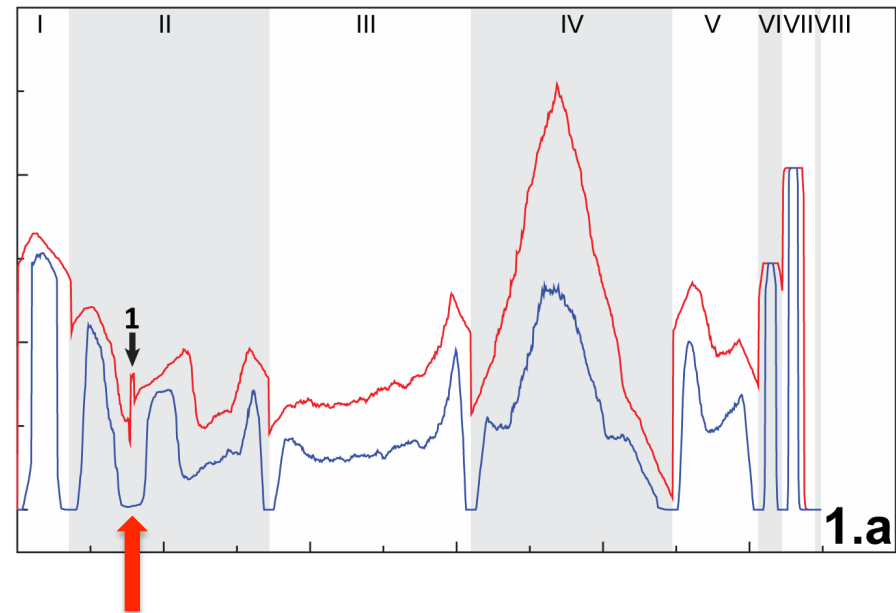
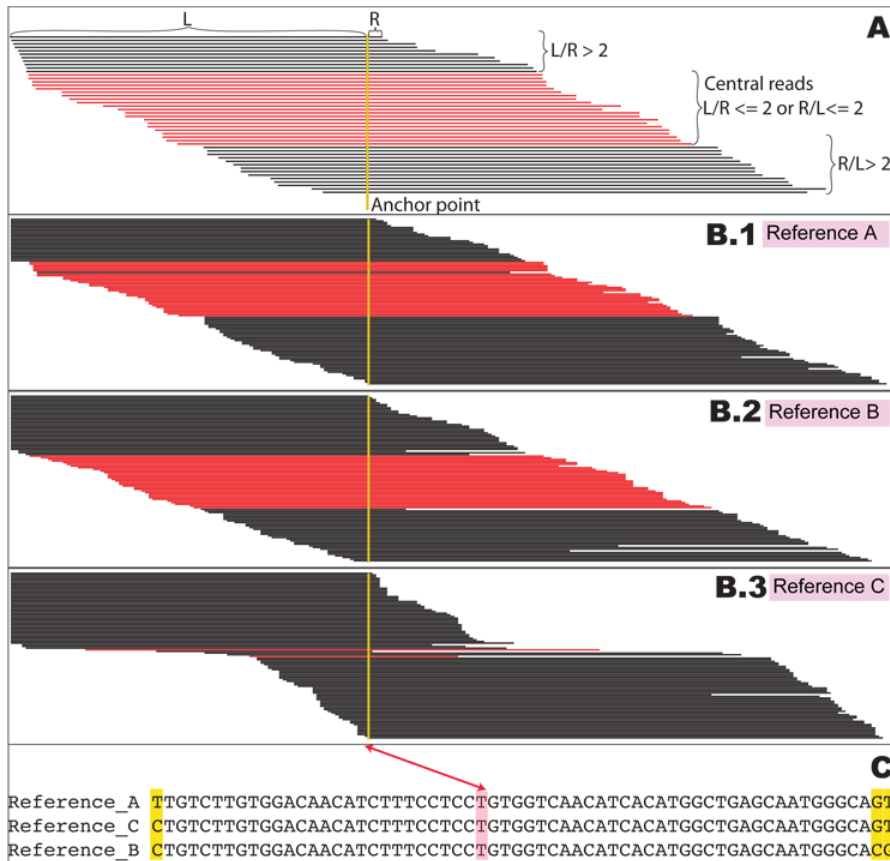
Multiplex Sequencing

Sonication products of each sample are ligated with sequencing adaptor of unique barcode. Barcoded fragments of several samples are pooled and size-selected. Isolation products are sequenced at both ends with Illumina sequencing platform.

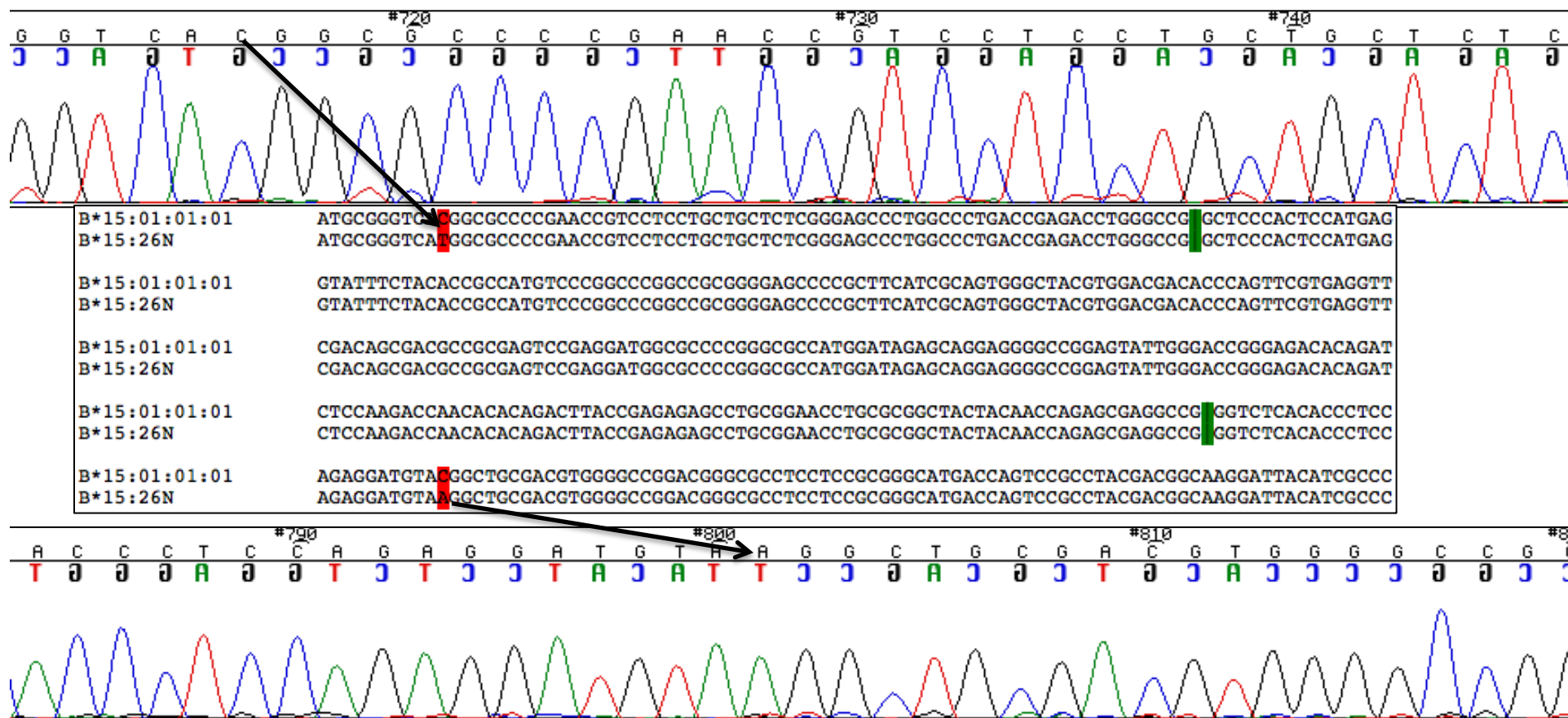
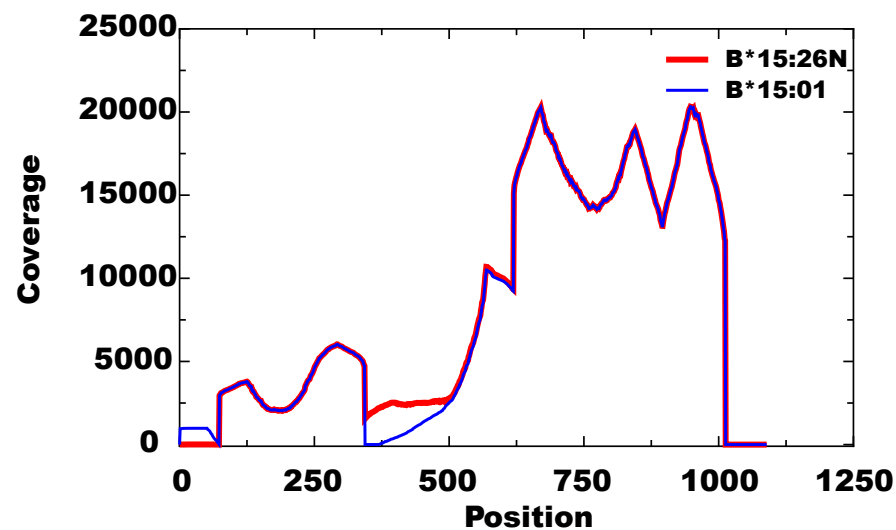
Data analysis pipeline



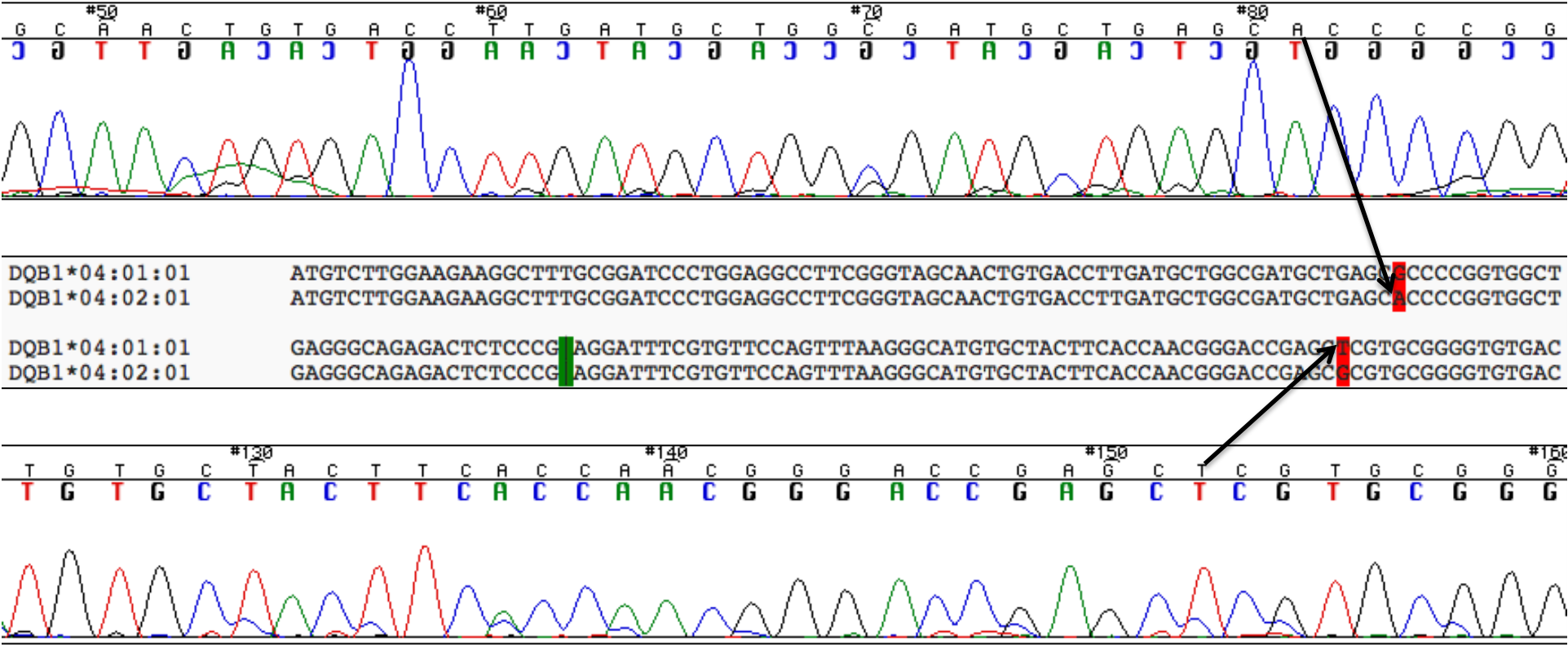
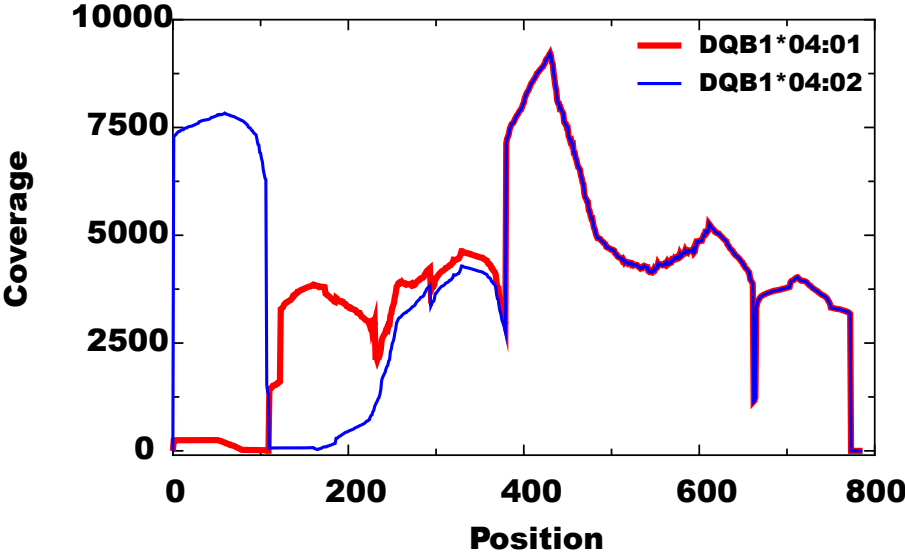
Central Reads Coverage



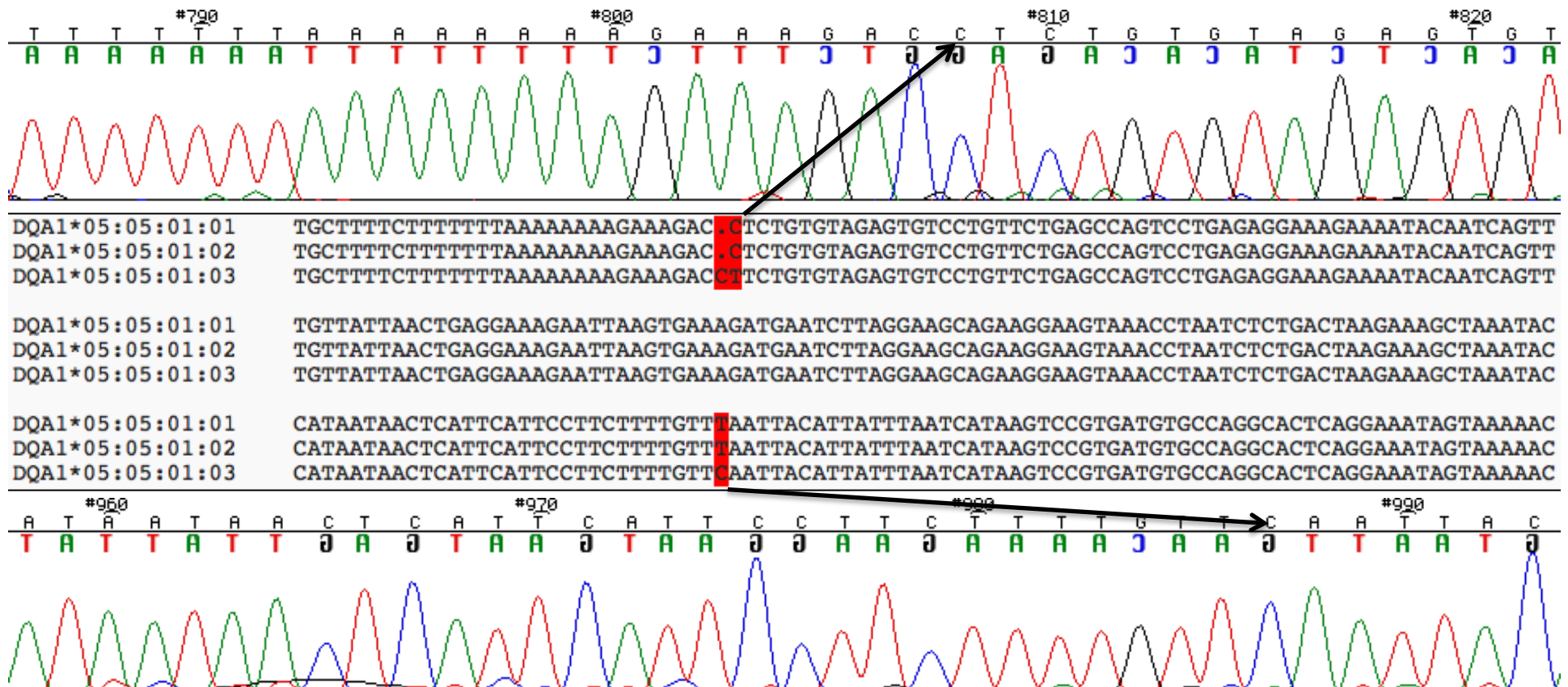
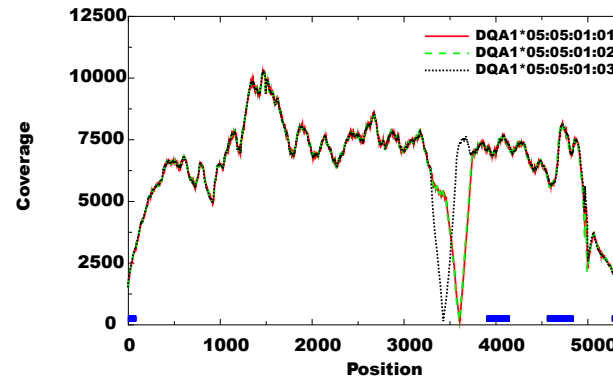
ISH3 novel



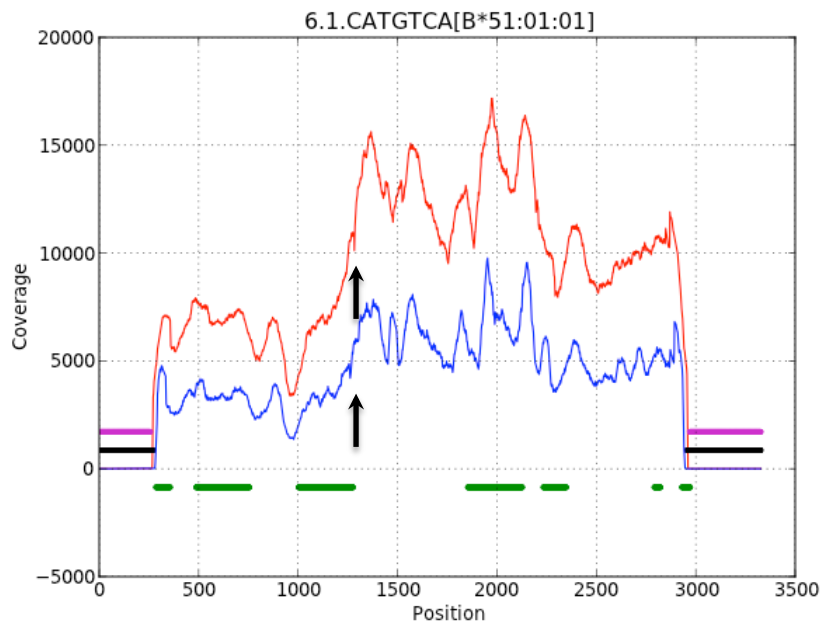
LKT3 novel



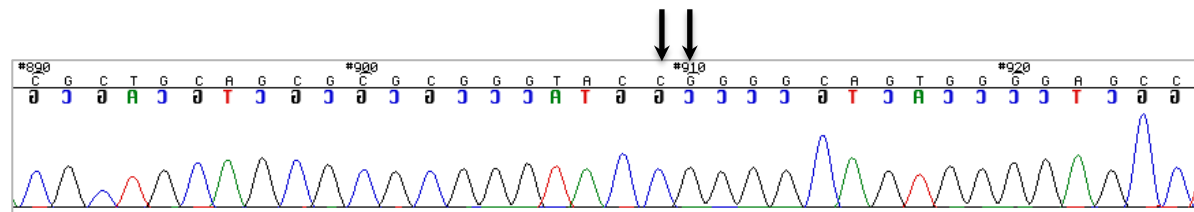
Detecting intron polymorphism



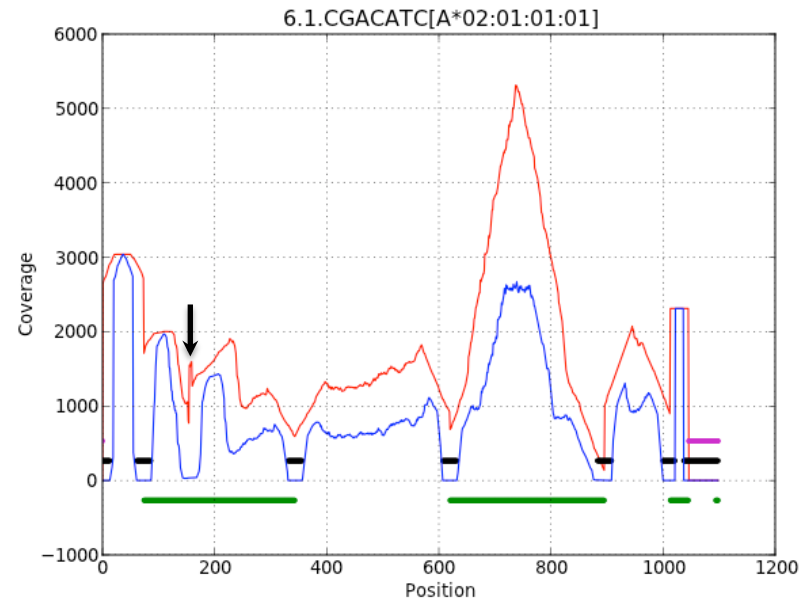
Detect a 1-bp deletion in an intron



```
>R GTGGAGTGGCTCCGCAGACACCTGGAGAACGGGAAGGAGACGCTGCAGCG  
>A GTGGAGTGGCTCCGCAGACACCTGGAGAACGGGAAGGAGACGCTGCAGCG  
  
>R CGCGGGTACCAGGGGCAGTGGGGAGCCTTCCCCATCTCCTATAGGTCGCC  
>A CGCGGGTACC-GGGGCAGTGGGGAGCCTTCCCCATCTCCTATAGGTCGCC
```



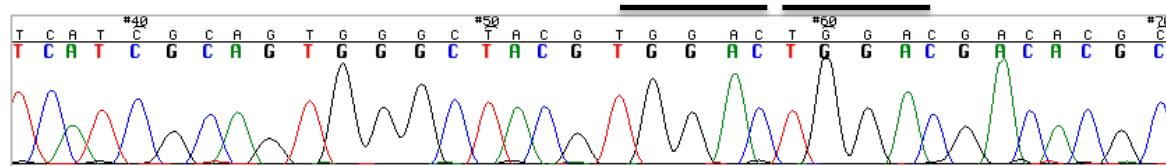
Detect 5-bp insertion in an exon



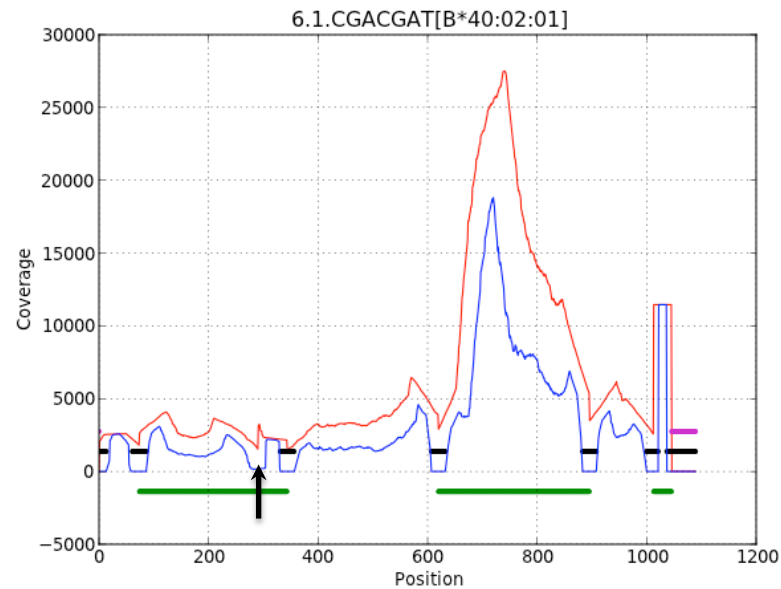
```

>R TCATCGCAGTGGGCTACG-----TGGACGACACGCAGTTCGTGCGGTTTCGACAGCGACGCCGCGAGCCAGAGGATGGAGCCGCGGGCGCCGTGGATAGAG
>A TCATCGCAGTGGGCTACGTGGACTGGACGACACGCAGTTCGTGCGGTTTCGACAGCGACGCCGCGAGCCAGAGGATGGAGCCGCGGGCGCCGTGGATAGAG

>R CAGGAGGGTCCGGAGTATTGGGACGGGGAGACACGGAAAGTGAAGGCCCACTCACAGACTCACCGAGTGGACCTGGGGACCCTGCGCGGCTACTACAACC
>A CAGGAGGGTCCGGAGTATTGGGACGGGGAGACACGGAAAGTGAAGGCCCACTCACAGACTCACCGAGTGGACCTGGGGACCCTGCGCGGCTACTACAACC
    
```

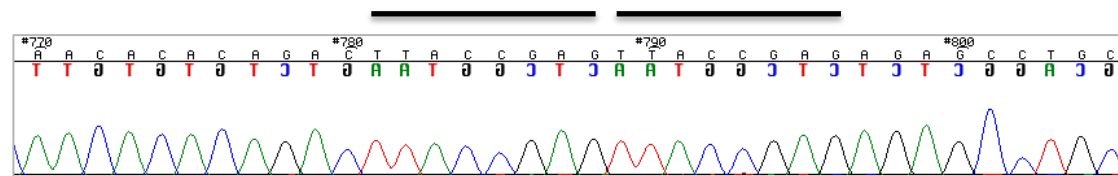


Detect 8-bp insertion in an exon



>R GCCACGAGTCCGAGGAAGGAGCCGCGGGCGCCATGGATAGAGCAGGAGGGGCCGGAGTATTGGGACCGGGAGACACAGATCTCCAAGACCAACACACAGA
>A GCCACGAGTCCGAGGAAGGAGCCGCGGGCGCCATGGATAGAGCAGGAGGGGCCGGAGTATTGGGACCGGGAGACACAGATCTCCAAGACCAACACACAGA

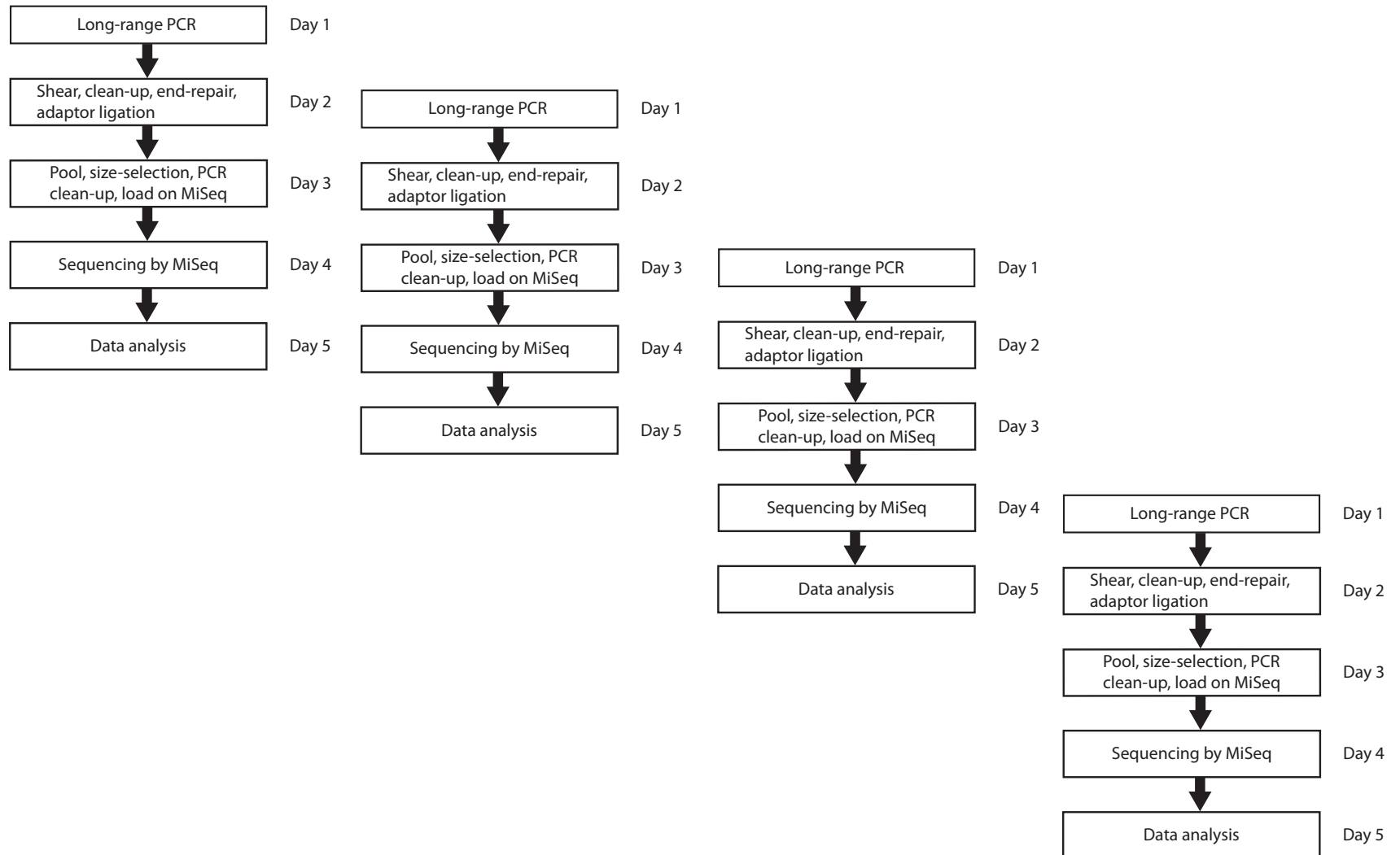
>R C-----TTACCGAGAGAGCCTGCGGAACCTGCGCGGCTACTACAACCAGAGCGAGGCCGGTGAGTGACCCCGGCCCGGGGCGCAGGTCACGACTCCC
>A CTTACCGAGTTACCGAGAGAGCCTGCGGAACCTGCGCGGCTACTACAACCAGAGCGAGGCCGGTGAGTGACCCCGGCCCGGGGCGCAGGTCACGACTCCC



Projected throughput with HiSeq2000

- Required minimum coverage = 20, average coverage = $20 \times 10 = 200$.
- For each sample, 8 (genes per sample) $\times 5000$ (average gene size) $\times 2$ (diploid) $\times 200$ (achieving $mc=20$) $\times 3$ (barcode variance) $\times 4$ (allele variance) = 192 million bp
- HiSeq2000 produces about 200 million reads or 40000 million bp per lane.
- Our experiences suggest that 80% of reads are mappable. Therefore, the multiplexing capacity per lane $40000 \text{ million bp} \times 0.8 / 192 \text{ million bp} =$ **166 samples per lane**. Or **2666 samples per instrumental run** (2 flow cells totaling 16 lanes) .

8-d turnaround with MiSeq for 20 samples



Further development

- Sequence regulatory regions such as 5'UTR and 3'UTR
- Build high-quality reference sequence database
- Target other polymorphic genes
- Test other sequencing platforms: Ion Torrent and Pacific Bioscience.

Acknowledgement

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 - Ron Davis
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