

Section 20. Ozone Bioindicator Plants

20.1 Overview	3
20.1.1 Scope and Application	3
20.1.2 Summary of Method	4
20.1.3 Summary of PDR Screens and Tally Procedures	4
20.1.4 Equipment And Supplies	4
20.1.5 Training and Quality Assurance	5
20.1.6 Voucher Specimens	5
20.1.7 Communications	5
20.2 Ozone Biomonitoring Procedures	7
20.2.1 Evaluation Window	7
20.2.2 Site Selection Procedures	7
20.2.3 Site Mapping	8
20.2.4 Split Plots	9
20.2.5 Species Selection	10
20.2.6 Plant Selection	12
20.2.7 Symptom Identification and Scoring	13
20.2.8 Collection of Leaf Samples and Voucher Data	15
20.2.9 Voucher Mailing Procedures	18
20.2.10 Crew Member Responsibilities	18
20.2.11 Field Procedures for Untrained Field Crews	18
20.3 Site Intensification	19
20.4 Plot Level Data	19
20.4.1 STATE	19
20.4.2 COUNTY	19
20.4.3 FIELD ID	20
20.4.4 SPLITPLOT ID	20
20.4.5 QA STATUS	20
20.4.6 CREW TYPE	20
20.4.7 OZONE SAMPLE KIND	21
20.4.8 CURRENT DATE	21
20.4.8.1 YEAR	21
20.4.8.2 MONTH	21
20.4.8.3 DAY	21
20.4.9 PLOT SIZE	22
20.4.10 ASPECT	22
20.4.11 TERRAIN POSITION	22
20.4.12 SOIL DEPTH	22
20.4.13 SOIL DRAINAGE (East only)	23
20.4.13 PLOT WETNESS (West only)	23
20.4.14 DISTURBANCE	23
20.4.15 INJURY CHECK	23
20.4.16 ELEVATION	24
20.4.17 Plot Notes	24
20.4.17.1 REMARK1 and REMARK2	24
20.5 GPS Coordinates	24
20.5.1 GPS Unit Settings, Datum, and COORDINATE SYSTEM	25
20.5.2 Collecting Readings	25
20.5.3 GPS UNIT	25
20.5.4 GPS SERIAL NUMBER	25
20.5.5 GPS ENTRY METHOD	26
20.5.6 GPS DATUM	26
20.5.7 Latitude	26
20.5.7.1 LATITUDE DEGREES	26

20.5.7.2	LATITUDE MINUTES	27
20.5.7.3	LATITUDE SECONDS.....	27
20.5.8	Longitude.....	27
20.5.8.1	LONGITUDE DEGREES	27
20.5.8.2	LONGITUDE MINUTES.....	27
20.5.8.3	LONGITUDE SECONDS	28
20.5.9	GPS ELEVATION	28
20.5.10	GPS ERROR.....	28
20.5.11	NUMBER OF GPS READINGS.....	28
20.5.12	GPS FILENAME (CORE OPTIONAL).....	28
20.6	Foliar Injury Data.....	29
20.6.1	SPECIES.....	29
20.6.2	AMOUNT.....	29
20.6.3	NUMBER OF PLANTS.....	29
20.6.4	SEVERITY	30
20.7	References	30
20.8	Acknowledgements.....	31
Appendix 20.A	Information and Data Sheets for the East.....	32
Appendix 20.B	Information and Data Sheets for the West.....	40
Appendix 20.C	Detailed Procedures for Handling Leaf Vouchers.....	46
Appendix 20.D	Site Ranking and Biosite Selection	48
Appendix 20.E	Supplemental Bioindicator Species	49

20.1 Overview

Air pollutants, such as ground-level ozone, are known to interact with forest ecosystems. Ozone is the only regional gaseous air pollutant that is frequently measured at known phytotoxic levels (Cleveland and Graedel 1979; Lefohn and Pinkerton 1988). Ozone pollution has been shown to have an adverse effect on tree growth and alter tree succession, species composition, and pest interactions (Forest Health and Ozone 1987; Miller and Millecan 1971; Smith 1974). In addition, we know that ozone causes direct foliar injury to many species (Skelly and others 1987; Treshow and Stewart 1973). We can use this visible injury response to detect and monitor ozone stress in the forest environment. This approach is known as biomonitoring and the plant species used are known as bioindicators (Manning and Feder 1980). Ozone bioindicator plants are used to monitor changes in air quality across a region, and to assess the relationship between ozone air quality and Phase 2 and Phase 3 indicators of forest condition (e.g., growth increment and dieback).

A useful bioindicator plant may be a tree, a woody shrub, or a nonwoody herb species. The essential characteristic is that the species respond to ambient levels of ozone pollution with distinct visible foliar symptoms that are easy to diagnose. Field studies and/or fumigation experiments have identified ozone sensitive species and characterized the ozone specific foliar response for both eastern (Davis and Umbach 1981; Duchelle and Skelly 1981; Krupa and Manning 1988) and western (Richards and others 1968; Mavity and others 1995; Brace 1996) bioindicators. Foliar injury symptoms include distinct patterns of coloration, often associated with accelerated senescence.

This section describes procedures to select field sites for ozone biomonitoring and to evaluate ozone injury on the foliage of sensitive plant species using the FIA ozone grid. Additional ozone sites, on an intensified ozone grid, may also be established by State and federal cooperators to improve the interpretive value of this indicator. This intensified sampling is done using the same methodology as the regular grid activities and is just as important.

20.1.1 Scope and Application

The scope of this indicator is national, but procedures are amended regionally as needed, particularly with regard to suitable sites and target species. Other variables, such as number of species, number of plants, and methods of scoring are standardized nationally. The procedures, reporting, and assessment goals were developed with the following considerations:

1. Ozone plot distribution across the landscape covers both the more remote and expansive forests away from population centers and the more fragmented forests located in close proximity to urban areas;
2. Ozone survey grid intensity reflects regional differences in air quality regimes and perceived risks to different forest types;
3. Sampling intensity in different regions is designed to allow links between ozone biomonitoring data and other FIA indicators;
4. Seasonal variability in ozone injury is addressed. We know that ozone injury must reach an undefined threshold within a leaf before the injury becomes visible to the human eye, and then tends to be cumulative over the growing season until fall senescence masks the symptoms.

There are certain regions of the country where ambient ozone concentrations, during the growing season, routinely exceed levels that are known to injure sensitive plants. Other regions have relatively clean air. In regions with poor air quality, the crew data underscore the extent and severity of ozone pollution in the nation's forests. In regions with better air quality, the emphasis must be on establishing a baseline for the ozone indicator. In this regard, field crews that do not find ozone injury (zero values for the ozone injury variables) are making a significant contribution to the national FIA database.

20.1.2 Summary of Method

Field procedures include the selection of a suitable site for symptom evaluation, identification of three or more known ozone-sensitive species at the site, and identification of ozone injury on the foliage of up to 30 plants of each species. Each plant is evaluated for the percentage of injured area and severity of injury on a five-point scale. Field crews record information on the location and size of the opening used for biomonitoring, and record injury amount and severity ratings for each plant.

In the East, to eliminate problems with seasonal variability in ozone response, all foliar evaluations are conducted during a four-week window towards the end of the growing season. In the West, due to differences in growing season, topography, target species, and other regional factors that influence plant response to ozone, the identification of an optimum evaluation window for this indicator is problematic. Nevertheless, to maintain national consistency and improve crew logistics, the western regions use a mid-season, five or six-week window for foliar injury evaluations.

In some States with a particular interest in air quality, foliar injury data are also collected from ozone sites on an intensified ozone grid. These supplementary ozone sites are standardized for certain site characteristics that influence ozone uptake by sensitive plants (Heck 1968; Krupa and Manning 1988), and are often co-located with physical air quality monitors. They are intended to improve the regional responsiveness of the ozone indicator.

Voucher specimens (pressed leaves with symptoms) are collected for each species for proper symptom identification. For each voucher, INJURY TYPE and INJURY LOCATION codes are recorded to fully describe the injury observed in the field. Additional quality control measures include field revisits and audits of 10 percent of the biomonitoring sites.

The implementation of an ozone survey grid independent of the traditional FIA plot system allows greater flexibility in plot location on the ground and greater sampling intensity in areas believed to be at high risk for ozone impact. In addition, plots are deliberately chosen for ease of access and for optimal size, species, and plant counts, thus maximizing data quality. Ozone is a regional pollutant, understood to have regional effects on vegetation. Therefore, data collected on the ozone grid will have direct application to the FIA P2 and P3 plots within the same region.

No specialized safety precautions are necessary to complete the fieldwork for the ozone indicator.

20.1.3 Summary of PDR Screens and Tally Procedures

Ozone indicator data are recorded on portable data recorders (PDRs). There are three basic data entry screens for ozone data: the Bioindicator Plot Identification Screen, the Bioindicator Plot Notes Screen, and the Bioindicator Species Screen. The Bioindicator Plot Identification Screen includes a record of site location and status as well as detail on site characteristics that influence ozone injury expression. The Plot Notes Screen prompts crews to record safety tips and additional information that will help analysts interpret the results or assist subsequent crews collecting data at the same location. The Bioindicator Species Screen prompts crews for injury AMOUNT and SEVERITY codes on a plant by plant basis. Help screens may be accessed for any variable from any of the three data entry screens.

For a written summary of the data entry procedures, definitions, and codes for the ozone measurement variables refer to section 20.4 through 20.6.

20.1.4 Equipment And Supplies

- A large diameter, 10X hand lens for close examination of plant leaves for ozone injury.
- Reference photographs and laminated leaf samples to aid in symptom identification.
- A forester-grade plant press with cardboard inserts to store leaf vouchers collected in the field.
- Envelopes ready for mailing the leaf vouchers to the National Ozone Field Advisor or the Western Regional Trainer.

- Stiff paper or cardboard for protecting the leaf vouchers in the mailing envelopes.
- Flagging: for temporary marking of sites or sample plants.
- Three field data sheets: (1) For documenting Foliar Injury Data in the event of a PDR failure; (2) For preparing the biosite location map; and (3) For recording Voucher Leaf Samples Data for QA. (see Appendix 20.A and 20.B).

20.1.5 Training and Quality Assurance

Each field crew member is trained and tested for familiarity with the site selection, species selection, and data collection procedures, and their ability to recognize ozone injury and discriminate against mimicking symptoms. Field crews are certified just prior to the beginning of the evaluation window for this indicator.

The National Ozone Field Advisor and one or more individuals in each region assume quality control responsibilities for the field season. Regional Advisors meet during a preseason session to refine methods and establish a unified approach to training, audits, and debriefing. Their responsibilities include: (1) training and certifying the field crews and QA staff, (2) documenting quality assurance procedures, and (3) conducting a debriefing session for the indicator.

A field audit crew revisits a subsample of the ozone ground plots in each region. In addition to hot audits of new crews, blind checks are performed on 10 plots per region. Auditing procedures cover species selection, symptom identification, and quantification of injury, as well as foliar sample collection, preservation and shipment.

Results of the field audit activities are used to determine if the measurement quality objectives are being met. Regional Advisors and Field Supervisors who are certified for the ozone indicator have the authority to implement whatever corrective action is needed in the field (e.g., retraining and retesting).

20.1.6 Voucher Specimens

Leaf samples are collected by field crews, cooperators, and all QA staff. They are to be placed in a small plant press immediately after removal from the selected plant. This is to preserve the integrity of the leaf sample and the injury symptoms until they can be validated by the National Indicator Field Advisor. A data sheet identifying the field crew and plot location is to be filled out and mailed with each sample.

Field crews, cooperators, and all QA staff collect leaf samples on the ozone biomonitoring sites according to procedures outlined in Subsection 20.2.8. These voucher specimens are pressed and mailed to the National Indicator Field Advisor for validation of the ozone symptom. If QA staff and regular field crews happen to be evaluating the same site at the same time, they collect and mail separate vouchers.

20.1.7 Communications

Any questions arising during the field season that cannot be answered by the Field Supervisor or State Coordinator, should be directed to the appropriate Regional Advisor or trainer for the ozone indicator. If any field crew or cooperator is uncertain about whom to call for information, or if a Regional Advisor is not indicated, they should contact the National Ozone Field Advisor or the National Ozone Advisor. Keep in mind that Advisors may be in the field and, therefore, unavailable for phone calls during normal workday hours. Messages left on answering machines should clearly identify who you are and when, where, and how to return your call. Field crews should be aware of differences in time zones and use email if possible.

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20.2 Ozone Biomonitoring Procedures

NOTE: In the following discussion the words site, biosite, and plot are used interchangeably to refer to the open area used for the ozone biomonitoring evaluations.

The primary objective of the field crew procedures for the ozone indicator is to establish an ozone biomonitoring site within each polygon on the FIA ozone grid using the site selection guidelines provided in the Decision Table – section 20.2.2. These sites are used to detect and monitor trends in ozone air pollution injury on sensitive species. Procedures include the selection of a suitable site for symptom evaluation, identification of three or more known ozone-sensitive species at the site, symptom identification and scoring on the foliage of up to 30 plants of each species, and the collection of voucher leaf samples. Each individual plant with ozone injury is scored for amount and severity of injury. Plants used for the selection of leaf vouchers are also evaluated for injury location and type. If a plant does not have ozone injury, it is still tallied with zeros for the amount and severity measurements. A hardcopy map, providing directions, plot coordinates, and key characteristics of the bioindicator site, is prepared for each plot.

All foliar evaluations are conducted during a defined ozone evaluation window. This is necessary to eliminate differences between plots that are caused by timing. During the evaluation window, all ozone sites on the ozone grid are evaluated for ozone injury. The same sites are evaluated every year.

20.2.1 Evaluation Window

Quantifying ozone injury on the FIA ozone plots is limited to an evaluation window starting in July and ending in mid-August. The evaluation window for crews in the Interior West begins the second week in July and extends through the third week in August. In the Pacific Northwest, the window is open from the third week in July through the third week in August. The evaluation window for crews in the North Region begins the last week of July and extends through the third week in August. In the Southern Region, the window is open from the third week in July through the third week in August.

All established biomonitoring sites are evaluated each year. The ozone injury evaluations are generally completed over a 5-to-20-day period during the window depending on the size of the State and the number of crews dedicated to the ozone survey. If possible, crews should adjust the timing of their evaluations so that the biomonitoring sites within each State are done at approximately the same time every year. Crews should adjust the timing of their evaluations for differences in elevation and latitude so that low elevation sites and/or more southern States use the earlier dates of the window while higher elevation sites and/or more northern States delay until the mid to later dates. Similarly, within each State, the low elevation, more southern biomonitoring sites should be evaluated first, the higher elevation, more northern sites last.

20.2.2 Site Selection Procedures

Site selection procedures begin with an in-office review of the ozone grid for each State. Candidate sites must be easily accessible open areas greater than one acre in size that are more than 100 feet (30 m) from a busy (paved) road. A site must contain at least 30 individuals of at least two bioindicator species to be evaluated for ozone injury. It is preferable that all sites have three or more species. Table 20-1 may be used as a decision guide for site selection.

Table 20-1. Decision guide for site selection

Decision Table	First Choice = Best Site	Second Choice
Access:	Easy	Easy
Location:	Single location is used.	One or two locations (split-plot).
Size of opening:	>3 acres (1.2h); wide open area; <50% crown closure.	Between 1-3 acres; long narrow or irregularly sized opening.
Species count:	More than three species.	Two or more species.
Plant count:	30 plants of 3 species; 10-30 plants of additional species.	30 plants of 2 species; 10-30 plants of additional species.
Soil conditions:	Low drought potential. Good fertility.	Moderate dry. Moderate fertility.
Site disturbance:	No recent (1-3 years) disturbance; No obvious soil compaction.	Little or no disturbance; No obvious soil compaction.

WEST NOTE: In many parts of the West, the forested landscape is characterized by large natural openings populated by a single overstory species. Large areas with a single bioindicator species (e.g., aspen or ponderosa pine) may be selected for biomonitoring, but every attempt should be made to combine this single species site with a nearby location that includes one or more of the understory bioindicator species. Ozone is a regional pollutant, affecting large geographic areas, and sites within 3 miles of each other generally have the same ozone exposure regime. Additional guidance on site ranking and biosite selection is provided in Appendix 20.D.

The best ozone sites are often associated with wildlife preserves on public land. Private landowners are often eager to participate in the ozone program. State and county parks and wildlife openings also provide good ozone sites. Other examples of suitable openings include old logging sites and abandoned pasture or farmland where you are reasonably certain that soil/site conditions are stable and free of chemical contaminants. Generally, if bedrock is exposed throughout an open area, then the soil conditions may be shallow, infertile, and often too dry to allow plants to respond to ozone stress. Sites that are routinely waterlogged are similarly unsuitable for biomonitoring. Avoid open areas where plants are obviously stressed by some other factor that could mimic or inhibit the ozone response. For example, the wooded edges of large parking lots in recreational areas are often highly compacted by car and foot traffic and should not be used. Do not select a site under a high-tension power line or on or near an active or reclaimed landfill. Do not select plants within 50 feet of the open edge around a cultivated field or tree plantation.

FIA crews and State Cooperators that have an established network of ozone sites may need to select and map replacement sites when previously mapped areas become overgrown or disturbed. Some sites may be split between two near-by locations to improve species and plant counts. In the case of split-plots, both have the same field identification number (i.e., FIELD ID) but different split-plot identification numbers (SPLITPLOT ID), so that each location can be uniquely identified and described.

No more than one half day should be spent locating a new or replacement bioindicator evaluation site. Crews must provide geographic coordinates (i.e., latitude and longitude) and the map datum (GPS_DATUM) for the geographic coordinates. If a site is split between two locations, the geographic coordinates for both locations are recorded.

20.2.3 Site Mapping

Once a bioindicator site is selected, the field crew records the estimated size of the site opening and other key site characteristics identified on the PDR or data sheet. The crew then maps the location of the site relative to some obvious and permanent marker such as a telephone pole, building, or property marker. Directions to the site, including road names and distances, are added to the map. Crews also mark the starting point for plant selection (see section 20.2.5) and approximate location of plant groupings used for evaluation (see section 20.2.6) on the site map. If available, a GPS unit is used to determine plot coordinates and elevation. Otherwise, this information is obtained from a USGS topographic map, generally the 7½ minute series quadrangle.

Ozone site maps are used by audit and regular crews in subsequent visits to the plot (see fig. 20-1) to ensure that the same site and the same population of plants are remeasured every year. This bioindicator site map must be kept with the appropriate State or federal cooperator so that it is readily available to whoever needs it.

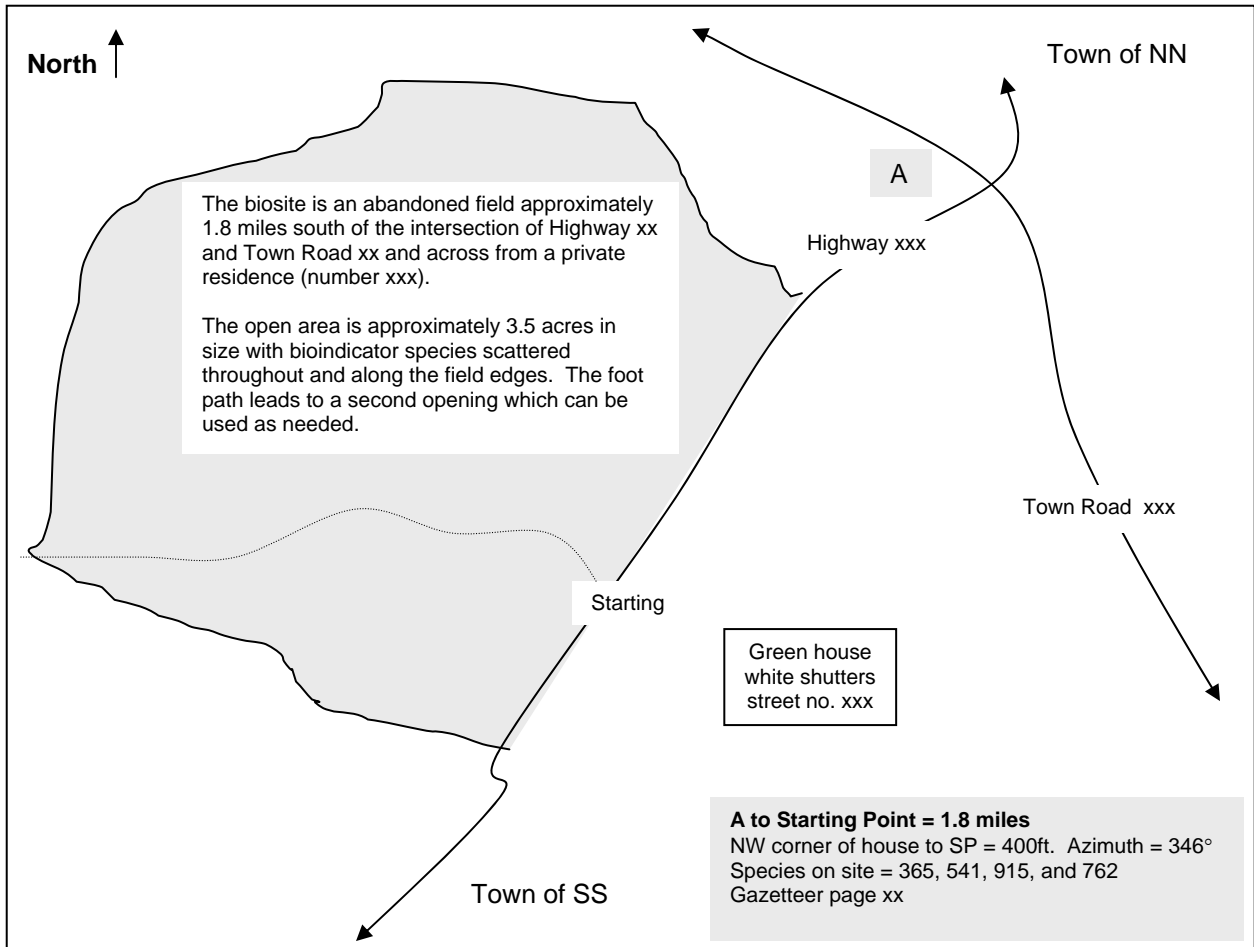


Figure 20-1. Example of a well-drawn map of an ozone biomonitoring site showing the location of the site relative to an obvious and permanent marker (road intersection and house number), road names and distances, North arrow, species codes and approximate location of plant groupings, location and distance to two major roads, distance and direction to two major towns, and Gazetteer reference page.

20.2.4 Split Plots

Maximizing the quality of each ozone plot with respect to the number of plants and species that are evaluated for ozone injury is a priority. As indicated in the site selection Decision Table in section 20.2.2, the best sites have more than 3 species; 30 plants of 3 species and between 10 and 30 plants of 1, 2, or 3 additional species. Finding high plant counts at a single wide-open location can be challenging. Split plots are intended to address this challenge. A split-plot consists of two different locations within 3 miles of each other, preferably with similar site characteristics (fig. 20-2). Species and plant counts from one location are combined with the species and plant counts from the second location to meet the species and plant count standards for site selection. On the PDR or data sheet, the same FIELD ID is assigned to each location. However, each location is assigned a different SPLITPLOT ID (1 or 2) so that each location can be uniquely identified and described. In the following example (fig. 20-2), the distance between the two open areas is less than 3 miles. The site selection criteria for a high quality ozone biosite are met as the total species and plant counts for FIELD ID XXXXXXXX are black cherry = 38, white ash = 30, milkweed = 30, and dogbane = 15.

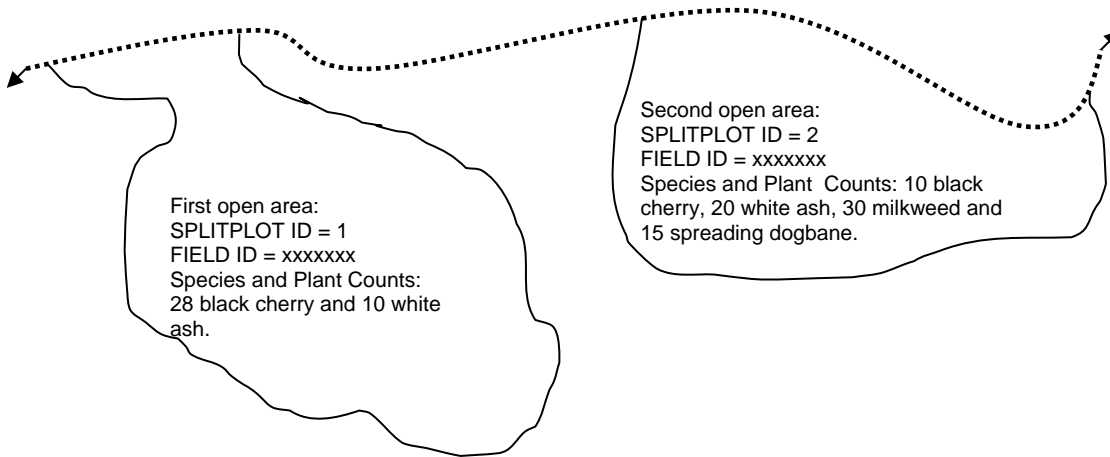


Figure 20-2. Split plot example: Distance between open areas along the access road is about 2 miles.

20.2.5 Species Selection

At the selected bioindicator site, the crew evaluates 30 individuals of three or more bioindicator species. If three species cannot be found at the site, then two species are still evaluated. Crews may combine species and plant counts from neighboring locations to obtain the required plant counts for each site. If 30 plants of two or more species cannot be found at the site, then a new site or additional location must be selected. A prioritized list of species is provided to the field crews for each region. The top three species in each list are the most common throughout the sampling region and should be selected for evaluation whenever possible. Species with 30 or more individual plants should be a first priority for choice of species, regardless of their position on the list. Key identifying characteristics of each species are provided in the Appendix 20.A (EAST) and Appendix 20.B (WEST). Photographs and species ID information can also be accessed from the ozone indicator web site. Go to the FIA home page (<http://fia.fs.fed.us>) and enter 'ozone' in the search window for access to the ozone indicator home page.

Field crews record the species code number for each selected species in the PDR or on the data sheet. The target species and codes are listed as follows: Table 20-2 – North and South Regions; Table 20-3 – Interior West Region; Table 20-4 – Pacific Northwest Region. Field crews in the Plains States including, but not limited to North Dakota, South Dakota, Nebraska, Kansas, Oklahoma, and Texas use a combination of eastern and western bioindicator species as shown in Table 20-5.

Table 20-2. Target species and codes for the North and South regions

Code	Definition – Bioindicator Species	Scientific Names
0915	Blackberry	<i>Rubus allegheniensis</i> (second year canes only)
0762	Black Cherry	<i>Prunus serotina</i>
0365	Common and Tall Milkweed	<i>Asclepias spp.</i>
0621	Yellow Poplar	<i>Liriodendron tulipifera</i>
0541	White Ash	<i>Fraxinus americana</i>
0931	Sassafras	<i>Sassafras albidum</i>
0366	Spreading Dogbane	<i>Apocynum androsaemifolium</i>
0364	Big Leaf Aster	<i>Aster macrophyllus: Eurybia macrophylla</i>
0611	Sweetgum	<i>Liquidambar styraciflua</i>
0761	Pin Cherry	<i>Prunus pensylvanica</i>

Table 20-3. Target species and codes for the Interior Region

Code	Definition	Scientific Names
0122	Ponderosa pine ¹	<i>Pinus ponderosa</i>
0116	Jeffrey pine ²	<i>Pinus jeffreyi</i>
0960	Blue elderberry	<i>Sambucus mexicana</i>
0746	Quaking aspen	<i>Populus tremuloides</i>
0924	Scouler's willow	<i>Salix scouleriana</i>
0351	Red alder ²	<i>Alnus rubra</i>
0909	Skunk bush	<i>Rhus trilobata</i>
0905	Ninebark	<i>Physocarpus malvaceus</i>
0969	Snowberry	<i>Symphoricarpos</i> spp ³
0907	Western wormwood	<i>Artemisia ludoviciana</i>
0961	Red elderberry	<i>Sambucus racemosa</i>
0965	Thin leaf huckleberry	<i>Vaccinium membranaceum</i>
0968	Evening primrose	<i>Oenothera elata</i>

¹ *Pinus ponderosa* var. *scopulorum* (WY, CO)

² *Pinus jeffreyi* (NV); *Alnus rubra* (ID)

³ *Symphoricarpos* spp. includes snowberry and coralberry species.

Table 20-4. Target species and codes for the Pacific Northwest Region

Code	Definition	Scientific Names
0122	Ponderosa pine ¹	<i>Pinus ponderosa</i>
0116	Jeffrey pine	<i>Pinus jeffreyi</i>
0960	Blue elderberry	<i>Sambucus mexicana</i> ²
0908	Mugwort	<i>Artemisia douglasiana</i>
0746	Quaking aspen	<i>Populus tremuloides</i>
0924	Scouler's willow	<i>Salix scouleriana</i>
0351	Red alder	<i>Alnus rubra</i>
0906	Pacific Ninebark (WC) ³	<i>Physocarpus capitatus</i>
0909	Skunk bush	<i>Rhus trilobata</i>
0905	Ninebark	<i>Physocarpus malvaceus</i>
0969	Snowberry	<i>Symphoricarpos</i> spp. ⁴
0907	Western wormwood	<i>Artemisia ludoviciana</i>
0961	Red elderberry	<i>Sambucus racemosa</i>
0965	Thin leaf huckleberry	<i>Vaccinium membranaceum</i>
0366	Spreading dogbane	<i>Apocynum androsaemifolium</i>
0968	Evening primrose	<i>Oenothera elata</i>

¹ *Pinus ponderosa* var. *ponderosa*

² Synonym for *S. mexicana* is *S. cerulea*.

³ WC = This species is only found west of the Cascades.

⁴ *Symphoricarpos* spp. includes coralberry and snowberry.

Table 20-5. Target species and codes for the Plains States

Code	Definition – Bioindicator Species	Scientific Names
0915	Blackberry	<i>Rubus allegheniensis</i> (second year canes only)
0762	Black Cherry	<i>Prunus serotina</i>
0365	Common and Tall Milkweed	<i>Asclepias</i> spp.
0621	Yellow Poplar	<i>Liriodendron tulipifera</i>
0541	White Ash	<i>Fraxinus americana</i>
0931	Sassafras	<i>Sassafras albidum</i>
0366	Spreading Dogbane	<i>Apocynum androsaemifolium</i>
0364	Big Leaf Aster	<i>Aster macrophyllus</i> : <i>Eurybia macrophylla</i>
0611	Sweetgum	<i>Liquidambar styraciflua</i>
0761	Pin Cherry	<i>Prunus pensylvanica</i>
0122	Ponderosa pine	<i>Pinus ponderosa</i>
0908	Mugwort	<i>Artemisia douglasiana</i>
0746	Quaking aspen	<i>Populus tremuloides</i>
0909	Skunk bush	<i>Rhus trilobata</i>
0905	Ninebark	<i>Physocarpus malvaceus</i>
0969	Snowberry and coralberry	<i>Symphoricarpos</i> spp.
0907	Western wormwood	<i>Artemisia ludoviciana</i>
0968	Evening primrose	<i>Oenothera elata</i>

Site selection requirements for species and plant counts (section 20.2.2, Decision Table for site selection) must be met using the species listed on the preceding tables (Tables 20-2, 20-3, 20-4, and 20-5). A list of supplemental bioindicator species (e.g., *Sambucus canadensis* American elder) that may be used as the 4th, 5th, and 6th, species at a selected biomonitoring site is provided in Appendix 20.E. This list may be updated annually as new information becomes available. Species on the supplemental list are for field trials only as they have not yet been adequately tested for ozone sensitivity under controlled conditions. Use the Plot Notes screen to maintain a record of when supplemental species have been used at a site.

NOTE: New data collection software may allow supplemental species (code 0998) to be tallied electronically using the same procedures as those used for the target species for each region.

20.2.6 Plant Selection

After site and species selection, the next task is contiguously to sample 30 individual plants of each species. Thirty plants of a target species must be sampled, if they are available on site. In fact, crews are strongly encouraged to evaluate 150 plants at each site (30 plants of five species), if possible. The value of the bioindicator data increases significantly with increased numbers of plants evaluated. This is true even if the crew records 30 consecutive zeros on three different species.

NOTE: The borders of some biomonitoring sites are difficult to determine and crews may be uncertain how much ground area to cover to complete the plant selection procedures. Specific guidelines are not set because the constraints on crew time and resources vary considerably from one State to the next. Time and safety concerns should take priority. Each crew must make every effort to maximize the number of plants and species evaluated for ozone injury at each plot location. Generally, ozone injury evaluations take 1 hour to complete and, assuming routine travel, crews in the East are expected to complete 3 ozone sites in a ten hour work day while crews in the West are expected to complete 2 ozone sites per work day.

The following procedures help crews to collect the bioindicator data in as systematic or unbiased a way as possible.

1. Identify a starting point at the edge of the opening. This point is mapped on the site data sheet so that audit and regular crews evaluate roughly the same population of plants in subsequent visits to the plot.

2. Move away from the starting point, towards the center of the opening.
3. Begin locating individuals in a sweeping pattern, selecting plants that are growing under the same or similar growing (microhabitat) conditions. Do not skip plants with little or no injury.
4. Select the more exposed plants (high sunlight exposure) and avoid suppressed and shaded individuals. Plants along the edge of an opening may be used if, in your judgment, they receive direct sunlight for three to four hours each day.
5. Avoid plants under 12 inches in height or so tall that you cannot see or touch at least half of the crown area.
6. Evaluate the foliage that you can see and touch on 30 plants of each species in the opening.
7. Record the amount and severity of injury for each plant evaluated (with or without symptoms) on the PDR, personal data assistant, or data sheet.

NOTE: A pop-up menu keeps track of the plant counts by species. For any one species, stop when the pop-up display indicates you have tabulated 30 plants, or when no additional plants of that species can be found on site. You can tabulate 30 plants of 5 species or any combination of species and plants that adds up to 150 data line entries.

Several bioindicator species (e.g., milkweed and blackberry) can spread vegetatively. This means that neighboring plants are often genetically identical. To avoid repeat sampling of clonal material, take several steps between each plant selected for evaluation. Use a systematic approach to select individual plants. For example, select the plant closest to your left side then take several steps and select the plant closest to your right side and repeat. A comparable approach should be used for all species to minimize bias in the plant selection process. If it is difficult to distinguish individual plants or stems for species like blackberry that grow in clumps, use an approximate 2-foot square area to represent a single plant.

NOTE: When a crew tabulates more than 30 plants of any one species from the list of official bioindicator species, the extra plants are referred to as **non-tallied plants**. Non-tallied plants may be used to indicate the presence or absence of ozone injury at a biosite. The most common use of non-tallied plants occurs when a crew samples 30 plants of a bioindicator species without finding injury and then encounters an additional plant or group of plants of this same species with obvious injury (see 20.4.15 INJURY CHECK).

NOTE: Crews are encouraged to evaluate species from the **supplemental species** list (Appendix 20.E), especially if plant counts of official bioindicator species are less than 30 per species at a given biosite. Supplemental species may eventually be added to the target bioindicator tables.

20.2.7 Symptom Identification and Scoring

The bioindicator species selected for each region are those that have been determined through field and laboratory studies to be highly sensitive to ozone air pollution. However, within a species, differences in genetics between individuals result in differential sensitivities to ozone. This means that you often find an individual of a species with severe air pollution injury growing immediately adjacent to another individual of the same species with few or no symptoms.

In addition to genetics, the age of the leaves (position on the stem, branch, or rosette) affects a plant's susceptibility to ozone air pollution. In general, leaves at 75 percent full expansion are the most sensitive and tend to show symptoms most definitively toward the center of the leaf. Older leaves show symptoms more widespread over the leaf surface, while younger leaves show symptoms more commonly near the leaf tip. If leaves on one branch are affected, then leaves at a similar leaf position on another branch should be affected, especially for branches on the same side of the plant under similar environmental conditions (sun or shade leaves).

The most common leaf injury symptom on broadleaf bioindicator species is upper-leaf-surface ozone stipple. On ponderosa and Jeffery pine, the most common needle injury symptom is ozone chlorotic mottle.

When scoring foliar symptoms on broadleaf bioindicator plants, check for the following characteristics of ozone injury .

- Symptoms are more severe on mid-aged and older leaves. New leaves will have no or very little injury.
- Symptoms are most likely confined to the upper leaf surface, and are typically visible as tiny purple-red to black spots (stippling).
- Check leaves covering each other. Overlapped leaves will have no injury on the bottom leaf.
- There will be some uniformity to size and shape of the lesions (stippling) on a leaf.
- Later in the growing season, stippling may be associated with leaf yellowing or premature senescence. Check the ground for fallen leaves.

When scoring foliar symptoms on pines, check for the following characteristics of ozone injury:

- Symptoms are visible as diffuse yellow areas (chlorotic mottle) without sharp borders between green and yellow zones, on older needles. Not all needles in a fascicle will be uniformly affected.
- Chlorotic mottle is rarely seen on current-year needles except in high-ozone areas. On young needles it may appear more olive than yellow.
- Older needles that are directly exposed to sunlight may show the most severe chlorotic mottle. However, almost all exposed branches on a plant will be affected to some degree.
- Premature needle drop frequently occurs on ozone-injured pines, even on trees that do not show other ozone injury symptoms. Check for missing older annual whorls and for large numbers of needles on the ground. Live crowns may appear small and thin.

WEST NOTE: Missing whorls on ponderosa pine should not be recorded as ozone injury without reliable evidence of other foliar injury symptoms, such as chlorotic mottle.

Each plant (broadleaf and conifer) with ozone injury is evaluated for the percent of the plant that is injured and the average severity of injury. For each plant located, the percentage of injured area and the severity of injury are both rated on a scale of 0 to 5 (see below). Both injury AMOUNT and injury SEVERITY estimates are confined to the exposed portion of the plant. If a plant does not have injury, it is still tallied with zeros for these measurements. For broad-leaved species, the AMOUNT and SEVERITY estimates are based on injury to the upper surface area of the leaves. For the pine species, examine all needle surfaces including the under sides, particularly if the needles have large amounts of winter fleck (NOT an ozone injury symptom) on the upper surfaces.

Percent Scale for injury AMOUNT: Estimate and record the percentage of leaves (or needles) on the plant with ozone injury symptoms relative to the total number of leaves (or needles) on the plant.

CODE	DEFINITION
0	No injury; the plant does not have any leaves/needles with ozone symptoms.
1	1 to 6 percent of the leaves/needles have ozone symptoms.
2	7 to 25 percent of the leaves/needles are injured.
3	26 to 50 percent of the leaves/needles are injured.
4	51 to 75 percent of the leaves/needles are injured.
5	>75 percent of the leaves/needles have ozone symptoms.

Percent Scale for SEVERITY of injury: Estimate and record the mean severity of symptoms on injured foliage.

CODE	DEFINITION
0	No injury; the plant does not have any leaves/needles with ozone symptoms.
1	On average, 1 to 6 percent of the leaf area of injured leaves/needles have ozone symptoms.
2	On average, 7 to 25 percent of the leaf area of injured leaves/needles have ozone symptoms.
3	On average, 26 to 50 percent of the leaf area of injured leaves/needles have ozone symptoms.
4	On average, 51 to 75 percent of the leaf area of injured leaves/needles have ozone symptoms.
5	On average, >75 percent of the leaf area of injured leaves/needles have ozone symptoms.

EAST NOTE: Blackberry and white ash have compound leaves. Use the whole leaf, not each leaflet, to estimate injury AMOUNT and injury SEVERITY. A typical clump of blackberry plants will have both current year (vegetative) and second year (flower and fruit bearing) canes available for evaluation. The injury AMOUNT and injury SEVERITY measurements are confined to the foliage on the second year canes. The foliage on the current year canes is naturally resistant to ozone injury. Do not use blackberry if you can find only current year canes.

WEST NOTE: Red and blue elderberry have compound leaves. Use the whole leaf, not each leaflet, to estimate injury AMOUNT and injury SEVERITY.

The percent scale for ozone injury evaluations has a long history of application in plant disease research. The scale utilizes break points that correspond to the ability of the human eye to distinguish gradations of healthy and unhealthy leaf tissue (see Horsfall and Cowling 1978). However, the recognition of ozone injury symptoms in the field is not an exact science, and mimicking symptoms can make field diagnosis difficult. Crews are expected to record AMOUNT and SEVERITY estimates for injury that they are unsure of as well as the more obvious or classic injury symptoms.

Proceed as follows:

1. Record the injury AMOUNT and the injury SEVERITY ratings for each plant on the PDR or data sheet.
2. Use the notes section on the PDR or data sheet to add other information that will help interpret the results (e.g., below average rainfall for the area).
3. Collect a voucher leaf sample (three leaves of each injured species evaluated at each location) and mail them to the National Field Advisor or Western Regional Trainer using the guidelines presented in Subsection 20.2.8 and 20.2.9.

NOTE: Do not take measurements in steady rain. Foliar symptoms are easiest to see under overcast skies. Bright sun will make it difficult to see the ozone stipple. Stand so that you reduce the glare on the leaf surface. Long periods without rain will inhibit symptom development even on the most sensitive plants. If you are experiencing below average rainfall for your area, please note this in the PDR or on the data sheet.

20.2.8 Collection of Leaf Samples and Voucher Data

The voucher leaf samples (leaves and/or needles) are a critical aspect of the data collection procedures as they provide the necessary validation of the ozone injury symptom observed in the field by the field crews. A plant press is essential to the collection of useable leaf samples and must be taken into the field by the field crews. Crew data that do not include a useable voucher leaf sample with a completed voucher data sheet are removed from the FIA database.

During the evaluation window, a voucher leaf sample must be collected for each injured species evaluated on the bioindicator site. For each injured broadleaf species, the voucher consists of three leaves that clearly show the ozone injury symptom. For pine species with ozone injury, the voucher consists of two small branches (small terminal or lateral branch containing the full complement of needles) with obvious chlorotic mottle. If a field crew in the East records ozone injury on blackberry, black cherry, and milkweed then a minimum of one voucher (3 LEAVES) from each of the three species (9 LEAVES IN ALL) is collected and mailed, with the corresponding voucher data sheet(s), to the National Field Indicator Advisor. If a field crew in the West records ozone injury on red alder, Scouler's willow, and ninebark then a minimum of one voucher (3 leaves) from each of the three species (9 leaves in all) is collected and mailed to the Western Regional Trainer.

The most useful voucher leaf samples show obvious foliar injury symptoms. If injury symptoms are not obvious and severe, send whatever leaf sample is available even if it is only one leaf with faint symptoms. Cut the leaf at the petiole, shake off any excess moisture, and place the leaf on blotter paper in the plant press. Each leaf is placed in the press so that it does not overlap another leaf. Include a label with each leaf

sample placed into the plant press that identifies which biosite the sample came from (i.e., FIELD ID) and the date. Petiole labels are provided for this purpose. Record the information on the labels with indelible ink and then wrap them around the petiole of at least one leaf per sample.

EAST NOTE: Blackberry and white ash have compound leaves. Select the whole leaf (not individual leaflets) when preparing a voucher sample.

WEST NOTE: Blue and red elderberry have compound leaves. Select the whole leaf (not individual leaflets) when preparing a voucher sample.

NOTE: If QA staff and regular field crews happen to be evaluating the same site at the same time, they collect and mail separate vouchers.

NOTE: The recognition of ozone injury symptoms in the field is not an exact science, and many other foliar injury symptoms can be mistaken for ozone injury. Crews are encouraged to collect and mail in voucher specimens of both known and suspected ozone injury for verification by the National Field Advisor (East) or Western Regional Trainer (West).

NOTE: At every biosite, crews evaluate up to 30 plants of a given species. Occasionally, crews identify ozone injury on a bioindicator species after entering 30 zero values for the first 30 plants of a given species. These extra plants are referred to as **non-tallied plants**. Crews that observe injury on non-tallied plants collect 3 leaves from the symptomatic plants and include them on the voucher mail-in sheet for the associated biosite indicating that the extra leaves are from non-tallied plants. If injury to non-tallied plants is validated, the biosite can be identified as plus ozone for certain maps and tables (see 20.4.15 INJURY CHECK).

NOTE: Crews are encouraged to evaluate species from the **supplemental species** list (Appendix 20.E), especially if plant counts of official bioindicator species are low for a given biosite. Supplemental species have passed certain field and fumigation trials that indicate they may be good bioindicators. Crews enter injury data for supplemental species on the plot notes screen unless new software is available to enter injury data using the same procedures that are used for target species. Crews that observe injury on supplemental species collect leaf vouchers to mail in with the voucher mail-in sheet using standard procedures.

The voucher data sheet must be completed for plot identification codes (e.g., STATE, COUNTY, FIELD ID, and SPLITPLOT ID), CURRENT DATE, CREW ID, QA STATUS, and SPECIES code(s). This sheet is filled out at the bioindicator site on the same day the sample is collected. In addition, the plants from which the leaf vouchers are selected must be evaluated by the field crews for INJURY LOCATION and INJURY TYPE (defined below), and for the amount of injury present on the leaf that is not ozone stipple. This information, together with the visible injury symptoms on the leaf samples, is used to validate the ozone injury data observed and recorded in the field by the field crews. For each species, the INJURY LOCATION and INJURY TYPE codes are intended to represent what the crew observed on the majority of the injured plants in the sample population. In contrast, the recorded estimates of percent injury caused by some stress other than ozone are based on what the crew observed on the injured leaf samples mailed in with the voucher data sheet.

The INJURY LOCATION and INJURY TYPE codes are recorded on the upper half of the voucher data sheet as follows:

INJURY LOCATION: Specify the leaf age or position of the leaves with ozone injury (EAST).

CODE	DEFINTION
1	>50% of the injured leaves are younger leaves. Younger leaves are usually located towards the branch tip (e.g., blackberry, black cherry, yellow poplar, white ash, sassafras, sweetgum, pin cherry, and spreading dogbane) or top of the plant (e.g., milkweed and big-leaf aster).
2	>50% of the injured leaves are mid-aged or older leaves. Mid-aged and older leaves are located halfway along the branch (e.g., blackberry, black cherry, yellow poplar, white ash, sassafras, sweetgum, pin cherry, and spreading dogbane), or main stem of the plant (e.g., milkweed and big-leaf aster), or more towards the base of the branch or stem.
3	Injured leaves are not concentrated in any one location, leaf age or position. Injury may be spread more or less evenly over the plant or is, otherwise, difficult to describe.

INJURY LOCATION: Specify the leaf age or position of the leaves with ozone injury (West – broadleaf species).

Code	Definition
1	>50% of the injured leaves are younger leaves. Younger leaves are usually located towards the branch tip (e.g., aspen, willow, oak, ninebark, and huckleberry), or top of the plant (e.g., elderberry, wormwood and snowberry).
2	>50% of the injured leaves are mid-aged or older leaves. Mid-aged and older leaves are located halfway along the branch (e.g., aspen, willow, oak, ninebark, and huckleberry) or main stem of the plant (e.g., elderberry, wormwood, and snowberry), or more towards the base of the branch or stem.
3	Injured leaves are not concentrated in any one location, leaf age or position. Injury may be spread more or less evenly over the plant or is, otherwise, difficult to describe.

INJURY LOCATION: Specify the leaf age or whorl with ozone injury (WEST – pine species).

Code	Definition
1	>50% of the injured needles are on the current whorl.
2	>50% of the injured needles are on whorls 1 year old and older.
3	Injury is not concentrated on any one needle whorl but is spread more or less evenly along the branch or is, otherwise, difficult to describe.

INJURY TYPE: Specify the visible injury symptom (EAST and WEST broadleaf species).

CODE	DEFINITION
1	The injury on >50% of the injured leaves is best described as upper-leaf-surface stipple, i.e., tiny purple-red to black spots occurring between the veins. Stippling may be associated with leaf yellowing and leaf drop late in the evaluation window; When injury is severe, stipples may coalesce and appear as uniform discoloration of the leaf surface.
2	The injury on >50% of the injured leaves is something other than upper-leaf-surface stipple. For example, small white to tan flecks occurring between the veins, or injury that is clearly visible on both leaf surfaces, or a general discoloration of the leaf that resembles early fall coloration.
3	The visible injury is varied or, otherwise, difficult to describe.

INJURY TYPE: Specify the visible injury symptom (WEST pine species).

Code	Definition
1	The injury on >50% of the injured needles is best described as chlorotic mottle i.e., small patches of yellow tissue with diffuse borders and surrounded by apparently healthy (green) tissue. Chlorotic mottle may be associated with premature needle drop.
2	The injury on >50% of the injured needles is something other than chlorotic mottle. For example, winter fleck on the upper surface of the needles, or tipburn (i.e., reddish brown discoloration of the needle tips).
3	The visible injury is varied or, otherwise, difficult to describe.

NOTE: Not all location and type codes are indicative of ozone injury. Certain combinations of location and type codes, considered with a questionable leaf voucher, may invalidate the injury data. Other combinations provide quality assurance for the injury assessment. Crews should describe any unusual or questionable symptoms on the upper half of the voucher data sheet.

20.2.9 Voucher Mailing Procedures

Vouchers are mailed in bulk at the end of the evaluation window, or earlier, depending on the crew's work schedule. It is very important to mail only dry, pressed leaf samples. Before mailing, make sure the upper half of the voucher data sheet is filled out. This sheet is filled out on the same day the sample is collected even if the sample is not mailed on that day. The STATE, COUNTY, FIELD ID, SPLITPLOT ID, DATE, QA STATUS, and crew identification codes must be entered correctly on the voucher data sheet before mailing. Please comment on the weather or general plot conditions that might help interpret the injury data. For example, *"It's been 14 days now without rain," "Every plant showed the same response and it was very obvious,"* or *"This was a highly disturbed site."* Avoid noting whether the crew thinks the leaf sample shows ozone injury or a mimicking symptom, and referring to the amount and severity ratings so as not to influence the validation process. Additional guidance on voucher preparation and mailing procedures is provided in Appendix 20.C.

WEST NOTE: Western crews are encouraged to add information on the biosite location to the voucher data sheet such as the uncoded name of the county or closest town. This helps the Western Regional Trainer map the initial findings from the leaf vouchers and alert FIA staff to high ozone areas.

The lower half of the voucher data sheet is filled out by the National Field Ozone Advisor (East) or Western Regional Trainer (West) to whom the sample is being sent. Place the voucher data sheet and the leaf sample between two pieces of stiff paper or cardboard before placing into a mailing envelope addressed to the National Field Advisor (East) or Western Regional Trainer (West). Manila folders and newspaper may also be used for voucher mailings. Do not tape the leaves or needles to the folders, paper or cardboard. Taped samples often break apart when they are handled, making evaluation difficult. Include as many samples as fit easily into each mailing envelope. There must be a unique voucher data sheet for each sample or species, unless the form is being used for multi-species. Keep leaf samples and the corresponding leaf voucher data sheets together. Leaf samples that are separated from the corresponding leaf voucher data sheets may be mislaid, especially if the petiole leaf labels are missing or incomplete.

WEST NOTE: The Western Regional Trainer will make every effort to provide immediate feedback on the leaf vouchers. To facilitate this, crews must fill in the contact information on the voucher data sheet.

20.2.10 Crew Member Responsibilities

1. Although one or two crew partners may be trained for this indicator, one person typically takes the lead responsibility for site selection, plant selection, and ozone injury evaluations. All procedures can be successfully completed by one person. Two person crews are recommended for safety reasons.
2. All members of the field crew may assist each other in the site selection process. Once a site is selected, one crew member is responsible for mapping the site and the location of bioindicator species on the field data sheet.
3. Only the crew member trained and certified in ozone injury evaluations may collect the amount and severity data and the leaf voucher. Other crew members may assist by recording the injury scores on the PDR or data sheet and by getting the plant press supplies ready.
4. The crew member that evaluates the plants for injury is responsible for collecting and mailing the voucher sample with air pollution symptoms.

20.2.11 Field Procedures for Untrained Field Crews

There are certain procedures for the ozone indicator that may be performed by individuals that have not attended the ozone training and been certified to collect ozone data. These procedures still require some explanation and oversight by the certified crew member. Untrained personnel may assist in the selection and mapping of the ozone biomonitoring site and in the location and identification of bioindicator species on the

selected site. They may not rate plant injury. It may also be helpful for the untrained crew person to act as the data recorder for the certified crew member, thus speeding up the data collection process.

20.3 Site Intensification

In addition to the unique ozone plots that are identified by the base ozone grid, some Cooperators have established additional biomonitoring sites to represent the local plant populations and environmental conditions. This is not an auxiliary effort, but an integral part of the monitoring activities for this indicator. In some States, additional biomonitoring sites are limited in number and are deliberately located close to weather and air quality monitoring stations. In other States, the ozone grid is intensified to allow for an unbiased allocation of additional biomonitoring sites. It is recommended that additional sites, whether few or many in number, be located on public land to facilitate the annual measurement activities.

Ozone biomonitoring sites added to the base grid typically possess attributes of an ideal site for evaluating ozone injury on sensitive species. They are larger than three acres, contain the maximum number of indicator species, and have soil/site conditions with low drought potential and adequate fertility. They are evaluated for ozone injury using the same methods and during the same time frame as described above in section 20.2.

20.4 Plot Level Data

All plot-level measurement codes for the ozone indicator are defined below. The words 'site' and 'biosite' are used interchangeably with the word 'plot' to refer to the ground location where ozone injury data are collected.

Ozone biosites vary in size and do not have set boundaries. Most biosites consist of a single ground location or plot, but two locations may be used to increase species and plant counts for an ozone biosite. If two locations are used, plot-level measurement codes are described separately for each location. When describing plot-level characteristics, use the predominant characteristics where most of the plant species are located. If conditions vary markedly across the site, or by species, then describe this in the plot notes or on the site map. Specify the aspect, terrain position, soil depth, soil drainage, disturbance, and elevation for the highest priority species (Subsection 20.4) found on the site. The soil depth, soil drainage, and disturbance variables are intended to describe general conditions on the plot and are not based on actual measurements. For a complete explanation of the procedures associated with these measurement codes, refer to section 20.2.

20.4.1 STATE

Record the unique FIPS (Federal Information Processing Standard) code identifying the State where the plot center is located.

When collected: All plots

Field width: 2 digits

Tolerance: No errors

MQO: At least 99% of the time

Values: See Appendix 1

20.4.2 COUNTY

Record the unique FIPS (Federal Information Processing Standard) code identifying the county, parish, Borough (or unit in AK) where the plot center is located.

When collected: All plots

Field width: 3 digits

Tolerance: No errors

MQO: At least 99% of the time

Values: See Appendix 1

20.4.3 FIELD ID

Record the unique code assigned to each ozone polygon. In some cases this will be a former FHM or P3 polygon number or special use plot number.

When collected: All plots
Field width: 7 digits
Tolerance: No errors
MQO: At least 99% of the time
Values: 0000001 - 9999999

20.4.4 SPLITPLOT ID

Record the SPLITPLOT ID number that describes whether an ozone plot consists of one or two locations. If two locations are selected, they must be within 3 miles of each other. Two locations are selected as needed to obtain optimal species and plant counts for each ozone biosite. The FIELD ID is the same for both locations.

When collected: All plots
Field width: 1 digit
Tolerance: No errors
MQO: At least 99% of the time
Values: 1 to 2

- 1 The ozone biosite consists of a single location or this is the first location of a biosite split between two locations.
- 2 The ozone biosite is split between two locations. This code identifies the second location added by the field crew to increase species and plant counts for a single polygon number.

20.4.5 QA STATUS

Record the code to indicate the type of plot data collected.

When collected: All plots
Field width: 1 digit
Tolerance: No errors
MQO: At least 99% of the time
Values: 1 to 2 and 4 to 7

- 1 Standard ozone plot
- 2 Cold check
- 4 Training/practice plot (off grid)
- 5 Botched plot file
- 6 Blind check
- 7 Hot check (production plot)

20.4.6 CREW TYPE

Record the code to specify what type of crew is measuring the plot. Select standard field crew when QA status has a value of 1; Select QA crew when QA STATUS has a value of 6.

When collected: All plots
Field width: 1 digit
Tolerance: No errors
MQO: At least 99% of the time
Values: 1 to 2

- 1 Standard field crew
- 2 QA crew (any crew member present collecting quality assurance data)

20.4.7 OZONE SAMPLE KIND

Record the code that describes the kind of plot being visited.

When collected: All plots
Field width: 1 digit
Tolerance: No errors
MQO: At least 99% of the time
Values: 1 to 3

- 1 Initial plot establishment on the base grid or on a newly intensified grid.
- 2 Remeasurement of a previously established plot, or replacement biosite within 3 miles of the previously established plot.
- 3 Replacement when the replacement biosite is more than 3 miles from the previously established plot.

20.4.8 CURRENT DATE

Record the year, month, and day that the current plot visit was completed as follows:

20.4.8.1 YEAR

Record the year that the plot was completed.

When collected: All plots
Field width: 4 digits
Tolerance: No errors
MQO: At least 99% of the time
Values: Beginning with 1998, constant for a given year

20.4.8.2 MONTH

Record the month that the plot was completed.

When collected: All plots
Field width: 2 digits
Tolerance: No errors
MQO: At least 99% of the time
Values: 01 to 12

January	01	May	05	September	09
February	02	June	06	October	10
March	03	July	07	November	11
April	04	August	08	December	12

20.4.8.3 DAY

Record the day of the month that the plot was completed.

When collected: All plots
Field width: 2 digits
Tolerance: No errors
MQO: At least 99% of the time
Values: 01 to 31

20.4.9 PLOT SIZE

Record the code that indicates the size of the opening used for biomonitoring.

When collected: All plots

Field width: 1 digit

Tolerance: No errors

MQO: At least 99% of the time

Values: 1 to 2

- 1 Greater than or equal to three acres.
- 2 Greater than one acre, but less than three acres.

20.4.10 ASPECT

Record the code that identifies the direction of slope for land surfaces with at least 5 percent slope as measured with a hand compass to the nearest degree.

When collected: All plots

Field width: 3 digits

Tolerance: +/- 30°

MQO: At least 99% of the time

Values: 000 - 360

- | | |
|-----|------------------------------|
| 000 | No aspect, slope < 5 percent |
| 001 | 1 degree |
| 002 | 2 degrees |
| . | . |
| . | . |
| 360 | 360 degrees, due north |

20.4.11 TERRAIN POSITION

Record the code that identifies the position of the plot in relation to the surrounding topography.

When collected: All plots

Field width: 1 digit

Tolerance: No error

MQO: At least 99% of the time

Values: 1 to 5

- 1 Ridge top or upper slope
- 2 Bench or level area along a slope
- 3 Lower slope
- 4 Flat land unrelated to slope
- 5 Bottom land with occasional flooding

20.4.12 SOIL DEPTH

Record the code that indicates the general depth of the soil where most of the bioindicator species are growing.

When collected: All plots

Field width: 1 digit

Tolerance: No error

MQO: At least 99% of the time

Values: 1 to 2

- 1 Bedrock is not exposed.
- 2 Bedrock is exposed; Soil is generally shallow.

20.4.13 SOIL DRAINAGE (East only)

Record the code that identifies the general soil drainage conditions where most of the bioindicator species are growing.

When collected: All plots

Field width: 1 digit

Tolerance: No error

MQO: At least 99% of the time

Values: 1 to 3

- 1 Soil is well drained
- 2 Soil is generally wet
- 3 Soil is excessively dry

20.4.13 PLOT WETNESS (West only)

Record the code that identifies the degree of wetness where most of the bioindicator plants are growing.

When collected: All plots

Field width: 1 digit

Tolerance: No error

MQO: At least 99% of the time

Values: 1 to 3

- 1 This is a wet plot; Riparian zone or bottomland.
- 2 This plot is moderately dry; Meadow or Northeast facing slope.
- 3 This plot is very dry; Exposed ledge, desert or alpine area.

20.4.14 DISTURBANCE

Record the code that identifies the presence and kind of disturbance where most of the bioindicator plants are growing. The area affected by any human caused or natural disturbance must be clearly visible and recent enough to influence plant health and condition. Disturbance that results in significant soil compaction is especially significant.

When collected: All plots

Field width: 1 digit

Tolerance: No error

MQO: At least 99% of the time

Values: 0 to 2

- 0 No recent or significant disturbance.
- 1 Evidence of overuse; Human activity causing obvious soil compaction or erosion.
- 2 Evidence of natural disturbance including fire, wind, flooding, grazing, pests, etc.

20.4.15 INJURY CHECK

Record the code that indicates whether ozone injury was observed on non-tallied plants or species. This variable allows a plot to be identified as impacted by ozone even though there is no quantitative data on injury severity for trend analyses. A leaf voucher must be collected to validate the injury.

When collected: All plots

Field width: 1 digit

Tolerance: No error

MQO: At least 99% of the time

Values: 0 to 1

- 0 No injury was observed on non-tallied plants or species.
- 1 Ozone injury was observed on non-tallied plants or species and a leaf voucher collected.

20.4.16 ELEVATION

Obtain elevation data from USGS topographic maps, generally the 7½ minute series quadrangle. Locate the area where most of the bioindicator species are growing and record elevation to the nearest foot.

When collected: When GPS UNIT = 0

Field width: 6 digits

Tolerance: +/-200 feet

MQO: At least 99% of the time

Values: -00100 to +20000

20.4.17 Plot Notes

Use these fields to record notes pertaining to the entire plot. If the notes apply to a specific aspect of the plot, then make that clear in the notes. Record the location where GPS coordinates were collected and the Datum used. If no GPS Unit was available, record the geographic coordinates (i.e., latitude and longitude) of the plot center in Degrees, Minutes, and Seconds using USGS topographic maps, generally the 7½ minute series quadrangle.

20.4.17.1 REMARK1 and REMARK2

Record any information on site characteristics, use of supplemental species, safety, plant location, injury patterns, or recent rainfall amounts that will assist subsequent crews visiting the site or help interpret the results.

When collected: All plots

Field width: Unlimited alphanumeric character field

Tolerance: N/A

MQO: N/A

Values: English language words, phrases and numbers

20.5 GPS Coordinates

Use a global positioning system (GPS) unit to determine the plot coordinates and elevation of all field visited ozone plot locations, even if GPS has been used to locate the plot in the past. GPS readings are collected according to procedures outlined in the FIA National Core Field Guide for Phase 2 & 3 Plots, Version 5.0. The ozone data entry applications accept GPS readings obtained using a geographic coordinate system (not UTM). If you are using UTM, record readings on the field data sheet for mapping and on the PDR Plot Notes screen. If GPS coordinates cannot be collected, elevation and plot coordinates are obtained from USGS topographic maps, generally the 7½ minute series quadrangle. Record ELEVATION on the Plot ID screen and approximate latitude and longitude on the Plot Notes screen.

NOTE: For several of the following GPS variables, the term plot center is used. There may be no obvious center to the ozone plots. Coordinates are collected as close as possible to a central location or marker that clearly locates the plot for returning crews. Explanatory notes are added to the plot map and Plot Notes screen as needed.

20.5.1 GPS Unit Settings, Datum, and COORDINATE SYSTEM

Consult the GPS unit operating manual or other regional instructions to ensure that the GPS unit internal settings, including Datum and Coordinate system, are correctly configured. Each FIA unit will use the NAD83 Datum to collect coordinates.

Each FIA unit will also determine which coordinate system to use. Regions using a Geographic system will collect coordinates in Degrees, Minutes, and Seconds of Latitude and Longitude; the regions using the UTM coordinate system will collect UTM Easting, Northing, and Zone.

20.5.2 Collecting Readings

Collect at least 180 GPS readings at the plot center (see Note above). These may be collected in a file for post-processing or may be averaged by the GPS unit. Each individual position should have an error of less than 70 feet if possible (the error of all the averaged readings is far less).

Soon after arriving at plot center, use the GPS unit to attempt to collect coordinates. If suitable positions (180 readings at error less than or equal to 70 feet) cannot be obtained, try again before leaving the plot center.

If it is still not possible to get suitable coordinates from plot center, attempt to obtain them from a location within 200 feet of plot center. Obtain the azimuth and horizontal distance from the "offset" location to plot center. Record the azimuth and horizontal distance as described in Sections 1.19.14 and 1.19.15.

Coordinates may be collected further away than 200 feet from the plot center if a laser measuring device is used to determine the horizontal distance from the "offset" location to plot center. Record the azimuth and horizontal distance as described in Sections 1.19.14 and 1.19.15.

In all cases try to obtain at least 180 positions before recording the coordinates. Coordinates not collected by automatic means shall be manually double-entered into the data recorder, both forward and backward.

20.5.3 GPS UNIT

Record the kind of GPS unit used to collect coordinates. If suitable coordinates cannot be obtained, record 0.

When collected: All field visited plots
Field width: 1 digit
Tolerance: No errors
MQO: At least 99% of the time
Values: 0 to 4

- 0 GPS coordinates not collected
- 1 Rockwell Precision Lightweight GPS Receiver (PLGR)
- 2 Other brand capable of field-averaging
- 3 Other brands capable of producing files that can be post-processed
- 4 Other brands not capable of field-averaging or post processing

20.5.4 GPS SERIAL NUMBER

Record the last six digits of the serial number on the GPS unit used.

When collected: When GPS UNIT >0
Field width: 6 digits
Tolerance: No errors
MQO: At least 99% of the time
Values: 000001 to 999999

20.5.5 GPS ENTRY METHOD

Identify the method used to record GPS data. If GPS data are manually entered, record 0. If GPS data are transferred electronically from the GPS receiver to the data recorder, record 1.

Upon entering a 1 the following variables are automatically populated in accordance with the GPS receiver setup in 1.19.1 (coordinates LATITUDE, LONGITUDE or UTM, GPS ELEVATION, GPS ERROR, and NUMBER OF READINGS). All other GPS variables must be populated via manual key-entry.

When Collected: GPS UNIT > 0

Field width: 1 digit

Tolerance: No errors

MQO: at least 99% of the time

Values: 0 to 1

- 0 GPS data manually entered
- 1 GPS data electronically transferred

20.5.6 GPS DATUM

Record the acronym indicating the map datum that the GPS coordinates are collected in (i.e., the map datum selected on the GPS unit to display the coordinates).

When collected: When GPS UNIT > 0

Field width: 5 characters (cccn)

Tolerance: No errors

MQO: At least 99% of the time

Values:

NAD83 North American Datum of 1983

20.5.7 Latitude

Record the latitude of the plot center to the nearest hundredth second, as determined by GPS.

NOTE: The following can be customized at the region level (e.g., decimal minutes to the nearest thousandth) as long as the final results recorded are within the specified tolerance to the nearest hundredth of a second or +/- 1.01 ft.

20.5.7.1 LATITUDE DEGREES

Record the latitude degrees of the plot center as determined by GPS.

When collected: When GPS UNIT = 1, 2, 3 or 4

Field width: 2 digits

Tolerance: When GPS ENTRY METHOD = 0, No errors in data entry

When GPS ENTRY METHOD = 1, not applicable

MQO: When GPS ENTRY METHOD = 0, at least 99% of the time

When GPS ENTRY METHOD = 1, not applicable

Values: 00-90

20.5.7.2 LATITUDE MINUTES

Record the latitude minutes of the plot center as determined by GPS.

When collected: When GPS UNIT = 1, 2, 3 or 4

Field width: 2 digits

Tolerance: When GPS ENTRY METHOD = 0, No errors in data entry
When GPS ENTRY METHOD = 1, not applicable

MQO: When GPS ENTRY METHOD = 0, at least 99% of the time
When GPS ENTRY METHOD = 1, not applicable

Values: 01 – 59

20.5.7.3 LATITUDE SECONDS

Record the latitude decimal seconds of the plot center to the nearest hundredth place as determined by GPS.

When collected: When GPS UNIT = 1, 2, 3 or 4

Field width: 4 digits

Tolerance: When GPS ENTRY METHOD = 0, No errors in data entry
When GPS ENTRY METHOD = 1, not applicable

MQO: When GPS ENTRY METHOD = 0, At least 99% of the time
When GPS ENTRY METHOD = 1, not applicable

Values: 0.00 – 59.99

20.5.8 Longitude

Record the longitude of the plot center to the nearest hundredth second, as determined by GPS.

NOTE: The following can be customized at the region level (e.g., decimal minutes to the nearest thousandth) as long as the final results recorded are within the specified tolerance to the nearest hundredth of a second or +/- 1.01 ft.

20.5.8.1 LONGITUDE DEGREES

Record the longitude degrees of the plot center as determined by GPS

When collected: When GPS UNIT = 1, 2, 3 or 4

Field width: 3 digits

Tolerance: When GPS ENTRY METHOD = 0, No errors in data entry
When GPS ENTRY METHOD = 1, not applicable

MQO: When GPS ENTRY METHOD = 0, At least 99% of the time
When GPS ENTRY METHOD = 1, not applicable

Values: 001-180

20.5.8.2 LONGITUDE MINUTES

Record the longitude minutes of the plot center as determined by GPS.

When collected: When GPS UNIT = 1, 2, 3 or 4

Field width: 2 digits

Tolerance: When GPS ENTRY METHOD = 0, No errors in data entry
When GPS ENTRY METHOD = 1, not applicable

MQO: When GPS ENTRY METHOD = 0, At least 99% of the time
When GPS ENTRY METHOD = 1, not applicable

Values: 01 – 59

20.5.8.3 LONGITUDE SECONDS

Record the longitude decimal seconds of the plot center to the nearest hundredth place as determined by GPS.

When collected: When GPS UNIT = 1, 2, 3 or 4

Field width: 4 digits

Tolerance: When GPS ENTRY METHOD = 0, No errors in data entry

When GPS ENTRY METHOD = 1, not applicable

MQO: When GPS ENTRY METHOD = 0, At least 99% of the time

When GPS ENTRY METHOD = 1, not applicable

Values: 0.00 – 59.99

20.5.9 GPS ELEVATION

Record the elevation above mean sea level of the plot center, in feet, as determined by GPS. If no GPS Unit is available, record elevation from the appropriate USGS topographic map.

When collected: When GPS UNIT = 1, 2 or 4

Field width: 6 digits

Tolerance: No errors

MQO: At least 99% of the time

Values: -00100 to 20000

20.5.10 GPS ERROR

Record the error as shown on the GPS unit to the nearest foot.

When collected: When GPS UNIT = 1 or 2

Field width: 3 digits

Tolerance: No errors

MQO: At least 99% of the time

Values: 000 to 070 if possible

071 to 999 if an error of less than 70 cannot be obtained

20.5.11 NUMBER OF GPS READINGS

Record a 3-digit code indicating how many readings were averaged by the GPS unit to calculate the plot coordinates. Collect at least 180 readings if possible.

When collected: When GPS UNIT = 1 or 2

Field width: 3 digits

Tolerance: No errors

MQO: At least 99% of the time

Values: 001 to 999

20.5.12 GPS FILENAME (CORE OPTIONAL)

Record the filename containing the GPS positions collected on the plot.

When collected: When GPS UNIT = 3

Field width: 8 characters.3 characters e.g. R0171519.ssf

Tolerance: No errors

MQO: At least 99% of the time

Values: Letters and numbers

20.6 Foliar Injury Data

All measurement codes for the foliar injury data are defined below. Plants selected for ozone injury evaluations are rated for the percent of injured area and the severity of injury on a scale of 0 to 5 (see section 20.2.7). If a plant does not have injury, it is tallied with zeros for these measurements. A pop-up menu keeps track of plant counts by species. The plot is complete only when 30 plants of 3 or more species have been tallied, or when no additional plants can be found on the plot. If three species cannot be found at the site, then two species are still evaluated. Ozone plots vary in size and do not have set boundaries. Time and safety concerns should dictate how much ground area to cover to complete the foliar injury evaluation procedures.

20.6.1 SPECIES

Record the three-digit code that identifies each species on the plot. Species codes may be entered in the order they are encountered as you walk through the plot evaluating plants. A pop-up menu keeps a running total of numbers of plants and species evaluated.

When collected: All plots
Field width: 4 digits
Tolerance: No error
MQO: At least 90% of the time
Values: See 20.2.5

20.6.2 AMOUNT

Record the code that identifies the percentage of leaves on the plant with ozone injury symptoms relative to the total number of leaves on the plant. The percent scale code and definitions are fully described in Subsection 20.2.7.

When collected: All plots
Field width: 1 digit
Tolerance: +/- 1 class
MQO: At least 90% of the time
Values: 0 to 5

- 0 No injury; The evaluated plant does not have any leaves or needles with ozone symptoms.
- 1 1 to 6 percent of the leaves/needles have ozone symptoms
- 2 7 to 25 percent of the leaves/needles are injured.
- 3 26 to 50 percent of the leaves/needles are injured.
- 4 51 to 75 percent of the leaves/needles are injured.
- 5 Greater than 75 percent of the leaves/needles have ozone symptoms.

20.6.3 NUMBER OF PLANTS

Record the number of plants tallied so far with no injury. When 0 is entered for AMOUNT, the PDR prompts for the NUMBER OF PLANTS with no injury. When a number greater than zero is entered for AMOUNT, the PDR prompts for the associated SEVERITY value. Zero and non-zero values for any species can be entered as they are encountered on the plot. The pop-up menu keeps track of plant counts by species.

When collected: When AMOUNT = 0
Field width: 2 digits
Tolerance: No error
MQO: At least 90% of the time
Values: 01 to 30

20.6.4 SEVERITY

Record the code that identifies the mean severity of symptoms on injured foliage. The percent scale code and definitions are fully described in Subsection 20.2.7.

When collected: When AMOUNT > 0

Field width: 1 digit

Tolerance: +/- 1 class

MQO: At least 90% of the time

Values: 0 to 5

- 0 No injury. The evaluated plant does not have any leaves or needles with ozone symptoms.
- 1 On average, 1 to 6% of the leaf area of injured leaves/needles has ozone symptoms.
- 2 On average, 7 to 25% of the leaf area of injured leaves/needles has ozone symptoms.
- 3 On average, 26 to 50% of the leaf area of injured leaves/needles has ozone symptoms.
- 4 On average, 51 to 75% of the leaf area of injured leaves/needles has ozone symptoms.
- 5 On average, greater than 75% of the leaf area of injured leaves/needles has ozone symptoms.

20.7 References

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20.8 Acknowledgements

The National Ozone Advisors wish to thank the individuals within FHM and FIA, as well as those outside the Forest Service, who have helped to review and improve this guide. Special thanks to Sally Campbell, Pat Temple, Jay Lackey, Teague Prichard, Ed Jepsen, William Manning, Art Chappelka, and John Skelly. Additional thanks to Dan Duriscoe, John Pronos, David Karnosky, Robert Kohut, Jim Renfro, and Dave Peterson who provided slides demonstrating ozone injury on the target bioindicator species. John Coulston, National Ozone Advisor, is the primary contact for questions about data access and analysis. He can be reached at the USDA Forest Service, Southern Research Station, 4700 Old Kingston Pike, Knoxville, TN 37919 or via email at jcoulston@fs.fed.us. For questions about the field program, contact Gretchen Smith at the Department of Natural Resources Conservation, 160 Holdsworth Way University of Massachusetts, Amherst, MA 01003-4210 or via email at gcsmith@nrc.umass.edu.

**Appendix 20.A Information and Data Sheets for the East
North and South regions, and Plains States**

Includes: (1) species key for eastern bioindicators, (2) species key for Plains States bioindicators, (3) data sheet for biosite characteristics, (4) data sheet for mapping biosite location, (5) foliar injury data sheet for eastern bioindicators, (6) voucher leaf sample data sheet for eastern bioindicators, and (7) voucher leaf sample data sheet for Plains States bioindicators.

Species Key for eastern bioindicators:

1. **Blackberry** is an upright or arching shrub; greenish to greenish-red stems are ridged with stout prickles. Alternate leaves have 3-7, mostly 5, leaflets, sparingly pubescent above, velvety beneath, green on both sides. Flowers white, May-July. Fruits black, July-September. Dewberry is very similar to common blackberry, but it is a vine with prickly stems trailing over the ground. Raspberry has smaller leaves and rounded stems covered with a whitish bloom. Blackberry is found in dry fields, clearings, and sunny thickets.

2. **Black Cherry** is a small to large tree. Twigs have a bitter-almond smell and taste. The alternate leaves are narrow, shiny, 2-6 inches long, and blunt-toothed, with the midrib prominently fringed beneath with white to brown hair. Leaves of choke cherry, a similar species, have a hairless midrib beneath and are sharp toothed. Leaves of pin cherry are longer and narrower with finely serrated margins. Black cherry is found on a variety of forest soils, deep and moist to dry and gravelly, and along the edges of disturbed areas.

3. **Common Milkweed** is recognized by a solitary, simple stem 1-6 feet tall that may or may not be covered with hair. The opposite or whorled leaves are twice as long (2 to 12 inches) as they are wide, have smooth margins, and stems with milky juice. The surface of the leaf is hairy below and smooth above. The petioles are short and thick. Flowers are borne in large clusters on stalks in the upper nodes. They appear rose or greenish-white, from June to August. You may see developmental stages of the Monarch butterfly or feeding injury on the plants. Milkweed is common along roadsides, in fields and meadows.

4. **Yellow Poplar** is a tall, straight, forest tree found on good sites with many hardwoods and loblolly pine in the South. Leaves are 4 to 6 inches in diameter, squarish at base, mostly 4-lobed, with smooth margins. Twigs stout, bitter to taste, with diaphragmed pith. Bud shaped like a duck's bill.

5. **White ash** is characterized by opposite, compound leaves; leaflets 5-9, stalked, green above and white or pale beneath, usually with smooth margins, slightly toothed near the leaf tips. Buds are inset in the leaf scar. Twigs are round, shiny, and mostly hairless. White ash is difficult to distinguish from green ash; Green ash leaves tend to be narrower, with more teeth, and hairy beneath; buds are set above the leaf scar and branch stems are usually hairy. Ash is sometimes confused with hickory, but can be readily distinguished by its opposite leaves and buds.

6. **Sassafras** has a characteristic odor and taste, spicy. Leaves are simple, narrowly lobed (mitten shaped) or entire. Twigs are green. Found from southwestern Maine, south to Florida, north to central Michigan, and west.

7. **Sweetgum** has star shaped leaves, deeply 5-7 lobed, margin finely serrate, bright green above, hairy in the axils of the leaf veins below. Twigs shiny and green to yellowish brown, somewhat fragrant when crushed. Fruit a spiny ball, often hanging. Common on bottomland soils and old fields from southern Connecticut, south to Florida and west.

8. **Pin Cherry** is a small, shrubby tree often found on cut over, burned, or abandoned sites. Leaves are long, narrow, finely serrate, and yellow-green; less shiny than those of black cherry. Pin cherry leaves may look like black cherry leaves, but they have no hair beneath. Maine to northern Georgia and west.

9. **Spreading Dogbane** is a perennial herb characterized by its opposite leaves with smooth margins and red stems with milky juice. The simple leaves are oblong or egg-shaped, dark green above and pale beneath; 2-3 inches long. The plant grows 1-4 feet high and has wide-spreading branches that give the plant an awkward appearance. It flowers throughout the summer; pinkish with a pink stripe in the center. Pods are long and narrow, in pairs. Young milkweed may be confused with dogbane, but differs in having larger, thicker leaves, hairy on the under surface. If evident, milkweed flowers are showy and the pods are large. Dogbane prefers the edges of dry woods from Canada to Mexico, but is also found in dry fields and thickets.

10. **Bigleaf Aster** is a perennial wild flower commonly found as an understory plant in dry woods. The leaves of this aster are heart shaped, 3 or more inches wide, with unevenly toothed margins, and have a stem nearly as long as the length of the leaf. Near the flat-topped flower cluster, the leaves become smaller and the stems are margined by a wavy leaf portion called a wing. Flowers may be violet, lavender, or light blue; evident in August and September. The plant grows 1-4 feet high and is native over eastern U.S. and south to North Carolina, west to Illinois.

Species Key for bioindicators used in the Plains States (e.g., North Dakota, South Dakota, Nebraska, Kansas, Oklahoma, and Texas). This key combines the 10 eastern bioindicator species with 8 western bioindicator species.

1. **Blackberry** is an upright or arching shrub; greenish to greenish-red stems are ridged with stout prickles. Alternate leaves have 3-7, mostly 5, leaflets, sparingly pubescent above, velvety beneath, green on both sides. Flowers white, May-July. Fruits black, July-September. Dewberry is very similar to common blackberry, but it is a vine with prickly stems trailing over the ground. Raspberry has smaller leaves and rounded stems covered with a whitish bloom. Blackberry is found in dry fields, clearings, and sunny thickets.
2. **Black Cherry** is a small to large tree. Twigs have a bitter-almond smell and taste. The alternate leaves are narrow, shiny, 2-6 in long, and blunt-toothed, with the midrib prominently fringed beneath with white to brown hair. Leaves of choke cherry, a similar species, have a hairless midrib beneath and are sharp toothed. Leaves of pin cherry are longer and narrower with finely serrated margins. Black cherry is found on a variety of forest soils, deep and moist to dry and gravelly, and along the edges of disturbed areas.
3. **Common Milkweed** is recognized by a solitary, simple stem 1-6 feet tall that may or may not be covered with hair. The opposite or whorled leaves are twice as long (2 to 12 in) as they are wide, have smooth margins, and stems with milky juice. The surface of the leaf is hairy below and smooth above. The petioles are short and thick. Flowers are borne in large clusters on stalks in the upper nodes. They appear rose or greenish-white, from June to August. You may see developmental stages of the Monarch butterfly or feeding injury on the plants. Milkweed is common along roadsides, in fields and meadows.
4. **Yellow Poplar** is a tall, straight, forest tree found on good sites with many hardwoods and loblolly pine in the South. Leaves are 4 to 6 in in diameter, squarish at base, mostly 4-lobed, with smooth margins. Twigs stout, bitter to taste, with diaphragmed pith. Bud shaped like a duck's bill.
5. **White ash** is characterized by opposite, compound leaves; leaflets 5-9, stalked, green above and white or pale beneath, usually with smooth margins, slightly toothed near the leaf tips. Buds are inset in the leaf scar. Twigs are round, shiny, and mostly hairless. White ash is difficult to distinguish from green ash; Green ash leaves tend to be narrower, with more teeth, and hairy beneath; buds are set above the leaf scar and branch stems are usually hairy. Ash is sometimes confused with hickory, but can be readily distinguished by its opposite leaves and buds.
6. **Sassafras** has a characteristic odor and taste, spicy. Leaves are simple, narrowly lobed (mitten shaped) or entire. Twigs are green. Found from southwestern Maine, south to Florida, north to central Michigan, and west.
7. **Sweetgum** has star shaped leaves, deeply 5-7 lobed, margin finely serrate, bright green above, hairy in the axils of the leaf veins below. Twigs shiny and green to yellowish brown, somewhat fragrant when crushed. Fruit a spiny ball, often hanging. Common on bottomland soils and old fields from southern Connecticut, south to Florida and west.
8. **Pin Cherry** is a small, shrubby tree often found on cut over, burned, or abandoned sites. Leaves are long, narrow, finely serrate, and yellow-green; less shiny than those of black cherry. Pin cherry leaves may look like black cherry leaves, but they have no hair beneath. Maine to northern Georgia and west.
9. **Spreading Dogbane** is a perennial herb characterized by its opposite leaves with smooth margins and red stems with milky juice. The simple leaves are oblong or egg-shaped, dark green above and pale beneath; 2-3 in long. The plant grows 1-4 feet high and has wide-spreading branches that give the plant an awkward appearance. It flowers throughout the summer; pinkish with a pink stripe in the center. Pods are long and narrow, in pairs. Young milkweed may be confused with dogbane, but differs in having larger, thicker leaves, hairy on the under surface. If evident, milkweed flowers are showy and the pods are large. Dogbane prefers the edges of dry woods from Canada to Mexico, but is also found in dry fields and thickets.
10. **Bigleaf Aster** is a perennial wild flower commonly found as an understory plant in dry woods. The leaves of this aster are heart shaped, 3 or more in wide, with unevenly toothed margins, and have a stem nearly as long as the length of the leaf. Near the flat-topped flower cluster, the leaves become smaller and the stems are margined by a wavy leaf portion called a wing. Flowers may be violet, lavender, or light blue; evident in August and September. The plant grows 1-4 feet high and is native over eastern U.S. and south to North Carolina, west to Illinois.
11. **Ponderosa Pine** is a large tree, up to 70 meters in height. Young tree bark is often thin and dark brown to black. Older tree bark is thick becoming yellow-red to cinnamon red and forming plates which slough off freely. Needles in bundles of three, 12-26 cm in length, not glaucous and yellow-green in color. Buds are resinous with red-brown scales

and dark-hairy. Cones with a prickle at the tip of each scale. May be confused with Jeffrey pine which differs by having non-resinous, light-brown buds, and grayish blue-green glaucous needles.

12. **Quaking Aspen** is a medium sized tree up to 36 meters in height. Bark is smooth, greenish-white. Buds shiny but not resinous. Leaf petiole is strongly flattened. The leaf blade is broadly ovate (almost round) with a tapering tip and finely toothed margins, upper surface smooth, lower surface covered with a bloom. Aspen could be confused with black cottonwood which differs in its resinous buds, rough bark and round leaf petioles.

13. **Ninebark** is an erect, loosely branched shrub with maple-like leaves and shreddy bark. May be up to 2 meters in height. Leaves and flowers similar to Pacific ninebark except the ovaries are densely hairy. May be confused with Douglas maple which has opposite leaves, or sticky currant, which has leaves that are sticky to the touch. Often associated with ponderosa pine and Douglas-fir forests at low to mid-elevation.

14. **Western Wormwood** is an aromatic perennial herb, 0.3 to 1.0 meter in height. Leaves mostly 3-11 cm long, variable in shape but most often with 3-5 narrow lobes, white hairy beneath, sometimes above as well. Flowers small and arranged in a loose, narrow flower cluster, 5-30 cm long. May be confused with Douglas' wormwood which has wider leaves and is usually found in moister habitats. Also similar to Riverbank wormwood which occurs only near streams and outwash areas.

15. **Mugwort** is a large perennial herb 0.5 to 1.5 meters tall, usually found in large colonies in wet areas, ditches, or drainages. Leaves are evenly-spaced, 1 to 10 cm long, the upper leaves are narrowly elliptical, the lower widely oblanceolate, often coarsely 3 to 5 lobed near the leaf tip, 2 to 3 cm wide, green above, covered with dense white hair beneath. Differs from western wormwood in having wider lower leaves and in its generally damp habitat.

16. **Evening Primrose** is a large biennial with elliptical leaves up to 25 cm long in a dense rosette the first year. The large (>1m) flowering stalk with long red-tinged elliptical leaves and large bright yellow four-petaled flowers forms in the second year. Both the leaves and stem are densely hairy, and the hairs often have red, blister-like bases. Usually found in moist, sunny habitats, like seeps or meadows.

17. **Snowberry** is a shrub, 0.5 to 1.5 meters in height with a solid brown pith. Bark: shreddy, brownish. Young twigs: hairy. Leaves opposite, elliptical, 1.0 to 3.5 cm long and half as wide. Flowers (May-June), the petals white with a pink tube. Fruit a white berry. Common snowberry differs by having a hollow pith. Trailing snowberry is a trailing shrub; and Utah honeysuckle has larger leaves and a solid white pith. Coralberry is similar to common snowberry except the fruit is a red berry. All are members of the honeysuckle family.

18. **Skunkbush** is a small, diffusively-branched shrub, 0.5 to 1 meter tall. The tips of the branches often droop down almost to ground level. The leaves are alternate, compound, with three leaflets, each of which is 3-lobed. The leaves resemble those of poison oak, but the leaflets of skunkbush are smaller, more hairy, and much more deeply-lobed. The leaves of skunkbush also emit a strong, ill-scented odor when crushed. However, if unsure, DO NOT crush the leaves of a shrub with three leaflets to determine the odor. Skunkbush is usually found on dry, open, brushy hillsides, while poison oak prefers damp or shaded forested areas and riparian habitats. Skunkbush is found throughout the southwest, from California and Arizona north to Colorado and Idaho.

OZONE BIOINDICATOR PLANTS - BIOSITE CHARACTERISTICS

Crew Reminder: Take this sheet to the Biosite! Complete it in the field!

This sheet must be completed only if you have *not* entered this same information on the Bioindicator Plot ID screen.

To be filled out by the FIELD CREW or Cooperator: Refer to Ozone Field Guide for code definitions.

STATE	COUNTY	FIELD ID	SPLITPLOT ID ¹	MONTH	DAY	YEAR	QA STATUS
							Standard O3 site QA check

¹SPLITPLOT ID refers to the number of locations (1 or 2) used for each FIELD ID. A separate sheet should be used for each location.

√ Please put a check mark beside the correct information. Please complete all data fields.

OZONE SAMPLE KIND:	
<input type="checkbox"/>	Initial biosite establishment on the FIA ozone grid. (Data collection in a previously empty polygon)
<input type="checkbox"/>	Remeasurement of a previously established biosite, or a replacement site within 3 miles of the previously established site.
<input type="checkbox"/>	Remeasurement when the replacement biosite is more than 3 miles from the previously established biosite.

Biosite size (PLOT SIZE):		TERRAIN POSITION:	
<input type="checkbox"/>	≥ 3.0 acres (1.2 hectares)	<input type="checkbox"/>	Ridge top or upper slope
<input type="checkbox"/>	1 to 3 acres (0.4 – 1.2 hectares)	<input type="checkbox"/>	Bench or level area along a slope
<input type="checkbox"/>	Other: please describe	<input type="checkbox"/>	Lower slope
<input type="checkbox"/>		<input type="checkbox"/>	Flat land unrelated to slope
<input type="checkbox"/>		<input type="checkbox"/>	Bottom land with occasional flooding

ASPECT: 000° = no aspect; 360° = N aspect	ELEVATION: record estimate in feet or meters	
Record to nearest degree =	Feet =	Meters =

SOIL DRAINAGE:		SOIL DEPTH:	
<input type="checkbox"/>	Well-drained	<input type="checkbox"/>	Bedrock not exposed
<input type="checkbox"/>	Wet	<input type="checkbox"/>	Bedrock exposed
<input type="checkbox"/>	Excessively dry	<input type="checkbox"/>	

DISTURBANCE: Disturbance on the site or in localized areas where the bioindicator plants are growing.	
<input type="checkbox"/>	No recent or significant disturbance; Do not count disturbance >3 years old.
<input type="checkbox"/>	Evidence of overuse; Human activity causing obvious soil compaction or erosion.
<input type="checkbox"/>	Evidence of natural disturbance including fire, wind, flooding, grazing, pests, etc.

Fill in below all that apply. Check here if geographic coordinates were obtained from a topographic map:

GPS UNIT:	GPS DATUM:	GPS SERIAL NUMBER:
Latitude =		GPS ERROR =
Longitude =		NUMBER OF READINGS =
GPS ELEVATION =		GPS FILE NAME =

¹If no GPS Unit is available, please use a map and record estimated latitude, longitude, and elevation for each biosite location.

Comments: Include information on additional species in the area, safety, directions, or additional site characteristics that may be useful.

File this completed data sheet with the sheet used for mapping the Bioindicator Site Location and then store it in the appropriate Ozone Plot Folder for your State or Region.

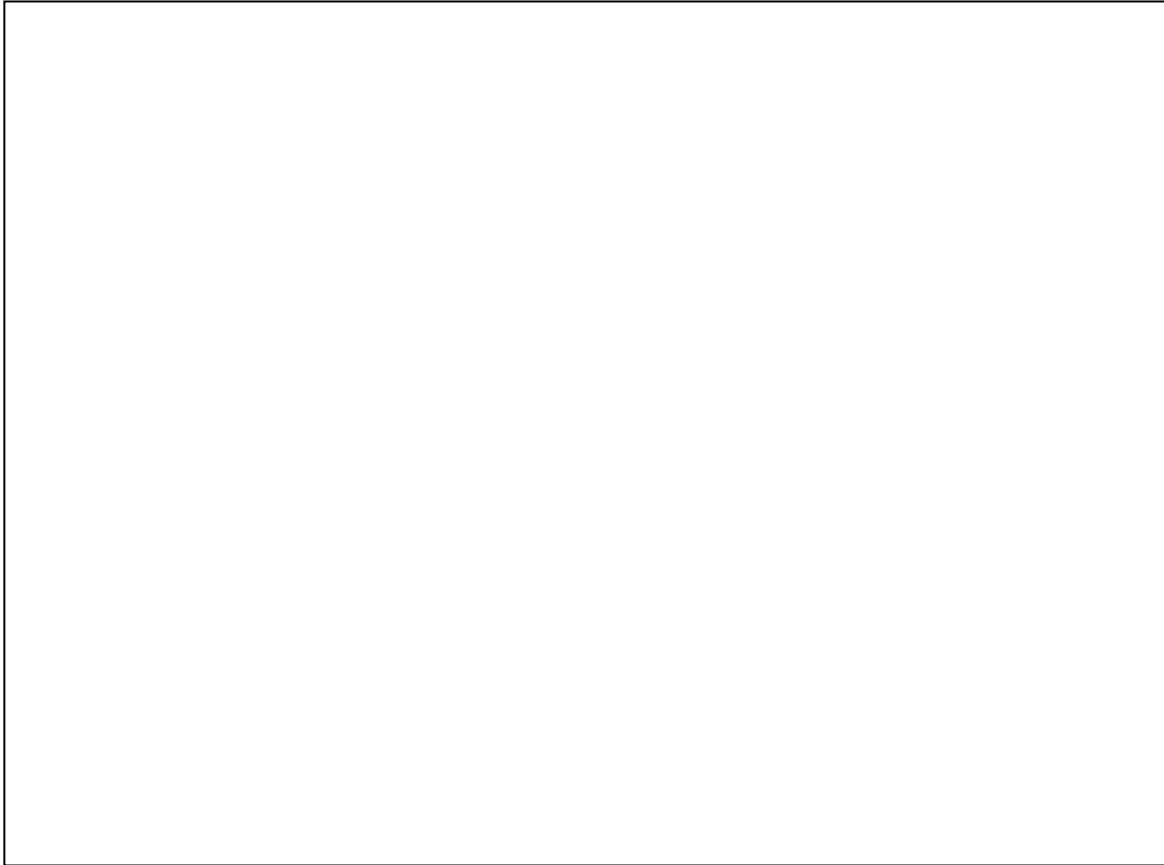
**Phase 3 Field Guide – Ozone Bioindicator Plants (combined), Version 5.1
October, 2011**

OZONE BIOINDICATOR PLANTS
Data Sheet for Mapping the Bioindicator Site Location
Add mileage information on reverse side.

To be filled out by the FIELD CREW or Cooperator: Refer to the Field Guide for code definitions.

STATE	COUNTY	FIELD ID	SPLITPLOT ID ¹	MONTH	DAY	YEAR	CREW ID

¹SPLITPLOT ID refers to the number of locations (1 or 2) used for each FIELD ID. A separate sheet should be used for each location.



Include the following information on the map. Use a legend that indicates distances between obvious points on the map.

- (1) Location of the site relative to some obvious and permanent marker.
- (2) Road names and distances as needed.
- (3) North arrow.
- (4) Species codes and approximate location of plant groupings used for the ozone injury evaluations.
- (5) Starting point
- (6) Location of and distance to two major roads
- (7) Distance and direction to two major towns
- (8) Gazetteer reference page if available.

Return the original of this map to the corresponding Plot Folder so it can be used by audit and regular crews in subsequent visits to the biosite. Mail a copy to the National Indicator Field Advisor the year that the site is established.

Geographic coordinates	GPS DATUM =
GPS LATITUDE =	GPS LONGITUDE =
Latitude estimated from a topographic map =	
Longitude estimated from a topographic map =	

REDRAW THE MAP AND ADD NEW INFORMATION EACH YEAR AS NEEDED!

OZONE BIOINDICATOR FOLIAR INJURY DATA SHEET

STATE	COUNTY	FIELD ID	SPLITPLOT ID	MONTH	DAY	YEAR	QA STATUS
							___ STANDARD ___ QA CHECK

Code	Species
915	Blackberry
762	Black Cherry
365	Milkweed
621	Yellow Poplar
541	White Ash
931	Sassafras
366	Spreading Dogbane
364	Big Leaf Aster
611	Sweetgum
761	Pin Cherry

Amount of Injury – % of leaves injured relative to the total leaf number
Severity of Injury – Average severity of symptoms on the injured leaves

Code	Scale
0	No Injury
1	1-6%
2	7-25%
3	26-50%
4	51-75%
5	>75%

Example 1

Amount: 8 inj out of 8 = 100%, Code 5
Severity: mean of 8 inj lvs = Code 4

Example 2

Amount: 4 inj out of 8 = 50%, Code 3
Severity: mean of 4 inj lvs = Code 2

Plant	Species Code		Species Code		Species Code		Species Code		Species Code	
	Amount	Severity	Amount	Severity	Amount	Severity	Amount	Severity	Amount	Severity
1										
2										
3										
4										
5										
6										
7										
8										
9										
10										
11										
12										
13										
14										
15										
16										
17										
18										
19										
20										
21										
22										
23										
24										
25										
26										
27										
28										
29										
30										

Did you collect 3 leaves that clearly show ozone stipple, for each injured species? Enter injury location and type codes for the collected leaf vouchers.

	Location = Type =	Location = Type =	Location = Type =	Location = Type =	Location = Type =
--	----------------------	----------------------	----------------------	----------------------	----------------------

Refer to the Ozone Bioindicator Plants section of the FIA Field Guide for codes and definitions.

Notes:

**OZONE BIOINDICATOR PLANTS
Data Sheet for the Voucher Leaf Samples**

To be filled out by the FIELD CREW or Cooperator: Refer to the Ozone Field Guide for code definitions.

STATE	COUNTY	FIELD ID	SPLITPLOT ID ¹	MONTH	DAY	YEAR	CREW ID	QA STATUS
								Standard QA Check

¹SPLITPLOT ID refers to the number of locations (1 or 2) used for each FIELD ID. Separate sheets should be used for each location.

Fill in the required codes. ONE SPECIES PER LINE. Code definitions are in the Field Guide.

Bioindicator Species Code or Common Name	Injury Location	Injury Type	Is the leaf sample injury close to 100% ozone stipple (✓), or is some other upper-leaf-surface injury also present (e.g., insect injury or fungal lesions)?
1 st			Close to 100% _____ Estimated percent other _____
2 nd			Close to 100% _____ Estimated percent other _____
3 rd			Close to 100% _____ Estimated percent other _____
4 th			Close to 100% _____ Estimated percent other _____

Species codes:

- 915 Blackberry
- 762 Black cherry
- 365 Milkweed
- 621 Yellow poplar
- 541 White ash
- 931 Sassafras
- 611 Sweetgum
- 761 Pin cherry
- 366 Spreading dogbane
- 364 Bigleaf aster.
- 999 Unknown
- 998 Supplemental
(write out common name)

Injury Location codes:

- 1 = greater than 50% of the injured leaves are younger leaves;
- 2 = greater than 50% of the injured leaves are mid-aged or older;
- 3 = injured leaves are all ages.

Injury type codes:

- 1 = greater than 50% of the injury is upper-leaf-surface stipple;
- 2 = greater than 50% is not stipple (tan flecks, bifacial or general discoloration);
- 3 = injury is varied or difficult to describe.

Biosite Notes:

CHECK ✓ all that apply:

Voucher leaves are from 1 plant:	Weather has been very dry:
Voucher leaves are from multiple plants:	Weather has been very wet:
Voucher leaves are undersized:	Biosite growth conditions are poor:
Normal sized leaves were uninjured or unavailable:	Biosite conditions are unsafe:
Voucher leaves are from NON-TALLIED plants:	Comment on back:

Mail this sheet with the leaf samples to:

Gretchen Smith
Department of Natural Resources Conservation
160 Holdsworth Way
University of Massachusetts
Amherst, MA 01003

QA/QC PERSON: To be filled out by the National Ozone Field Advisor or Regional Expert. ✓

Date checked	Date rechecked	Sample condition			Plot Status	
		GOOD easy to read - ID obvious	FAIR	POOR samples unreadable or not labeled correctly	(+ozone)	(- ozone)

Bioindicator Species	Positive for ozone	Negative for ozone	Explanation

OZONE BIOINDICATOR PLANTS
Data Sheet for the Voucher Leaf Samples – PLAINS STATES

To be filled out by the FIELD CREW or Cooperator: Refer to the Ozone Field Guide for code definitions.

STATE	COUNTY	FIELD ID	SPLITPLOT ID ¹	MONTH	DAY	YEAR	CREW ID	QA STATUS
								Standard QA Check

¹SPLITPLOT ID refers to the number of locations (1 or 2) used for each FIELD ID. Separate sheets should be used for each location.

Fill in the required codes. ONE SPECIES PER LINE. Code definitions are in the Field Guide.

Bioindicator Species Code or Common Name	Injury Location	Injury Type	Is the leaf sample injury close to 100% ozone stipple (✓), or is some other upper-leaf-surface injury also present (e.g., insect injury or fungal lesions)?
1 st			Close to 100% _____ Estimated percent other _____
2 nd			Close to 100% _____ Estimated percent other _____
3 rd			Close to 100% _____ Estimated percent other _____
4 th			Close to 100% _____ Estimated percent other _____

Species codes:

- 0915 Blackberry
- 0762 Black cherry
- 0365 Milkweed
- 0621 Yellow poplar
- 0541 White ash
- 0931 Sassafras
- 0611 Sweetgum
- 0761 Pin cherry
- 0366 Spreading dogbane
- 0364 Bigleaf aster.
-
- 0122 Ponderosa pine
- 0746 Quaking aspen
- 0905 Ninebark
- 0907 Wormwood
- 0908 Mugwort
- 0909 Skunkbush
- 0968 Evening primrose
- 0969 Snowberry
- 0998 Supplemental (e.g. Rudbeckia sp.)
please write out common name
- 0999 Unknown

Injury Location codes:

- 1 = greater than 50% of the injured leaves are younger leaves (or current whorl);
- 2 = greater than 50% of the injured leaves are mid-aged or older (older whorls);
- 3 = injured leaves are all ages.

Injury type codes:

- 1 = greater than 50% of the injury is upper-leaf-surface stipple (or chlorotic mottle)
- 2 = greater than 50% is not stipple (tan flecks, bifacial or general discoloration);
- 3 = injury is varied or difficult to describe.

Biosite Notes:

CHECK ✓ all that apply:

- | | |
|--|-------------------------------------|
| Voucher leaves are from 1 plant: | Weather has been very dry: |
| Voucher leaves are from multiple plants: | Weather has been very wet: |
| Voucher leaves are undersized: | Biosite growth conditions are poor: |
| Normal sized leaves were uninjured or unavailable: | Biosite conditions are unsafe: |
| Voucher leaves are from NON-TALLIED plants: | Comment on back: |

Mail This Sheet With Leaf Samples To:

[EASTERN SPECIES]

Gretchen Smith
Department of Natural Resources Conservation
160 Holdsworth Way
University of Massachusetts
Amherst, MA 01003

[WESTERN SPECIES]

Pat Temple
USDA FS, PSW Experiment Station
4955 Canyon Crest Drive
Riverside, CA 92507

QA/QC PERSON: To be filled out by the National Ozone Field Advisor or Regional Expert. ✓

Date checked	Date rechecked	Sample condition			Plot Status	
		GOOD easy to read - ID obvious	FAIR	POOR samples unreadable or not labeled correctly	(+ozone)	(- ozone)
Bioindicator Species	Positive for O3	Negative for O3	Explanation			

Appendix 20.B Information and Data Sheets for the West

Includes: (1) species key for western bioindicators, (2) data sheet for biosite characteristics, (3) data sheet for mapping biosite location, (4) foliar injury data sheet for western bioindicators, and (5) voucher leaf sample data sheet for western bioindicators.

Key Identifying Characteristics of the Western Ozone Bioindicator Species**

1. **Ponderosa Pine** is a large tree, up to 230 feet in height. Young tree bark is often thin and dark brown to black. Older tree bark is thick becoming yellow-red to cinnamon red and forming plates which slough off freely. Needles in bundles of three, 5-10 inches in length, not glaucous and yellow-green in color. Buds are resinous with red-brown scales and dark-hairy. Cones with a prickle at the tip of each scale. May be confused with Jeffrey pine which differs by having non-resinous, light-brown buds, and grayish blue-green glaucous needles.
2. **Jeffrey Pine** is a smaller tree than ponderosa pine, with darker cinnamon-red bark that may be tinged with lavender on old trunks. Needles in bundles of three, 5-10 inches in length, blue-green, and somewhat twisted. Crushed needles and twigs have a violet-like or pineapple odor. Buds are *never* covered with resin droplets. Cones with a prickle at the tip of each scale. May be confused with ponderosa pine.
3. **Quaking Aspen** is a medium sized tree up to 118 feet in height. Bark is smooth, greenish-white. Buds shiny but not resinous. Leaf petiole is strongly flattened. The leaf blade is broadly ovate (almost round) with a tapering tip and finely toothed margins, upper surface smooth, lower surface covered with a bloom. Aspen could be confused with black cottonwood which differs in its resinous buds, rough bark and round leaf petioles.
4. **Scouler's Willow** is a small tree or shrub up to 32 feet in height. Leaf blade is 1-4 inches in length, narrowly elliptic with the widest portion toward the tip, entire to irregularly toothed margins, lower surface smooth, upper surface shiny. This willow is NOT restricted to riparian zones. It can be easily confused with a number of other willow species. The combination of leaves widest toward the tip (mostly rounded ends and narrowly tapered bases) and the tolerance for upland (drier) habitats makes this willow relatively easy to identify.
5. **Pacific Ninebark** is a deciduous shrub 6-12 feet in height. Leaves alternate, 3 or 5 lobed (maple-like), 2-3 inches long, serrate, dark green and smooth above, paler and hairy below. Twigs red to grayish brown, splits longitudinally into long strips. Flowers small, white, borne in a cluster, stems hairy. Very similar to ninebark (see below) which is generally smaller, in drier habitats, and with densely hairy ovaries.
6. **Ninebark** is an erect, loosely branched shrub with maple-like leaves and shreddy bark. May be up to 6 feet in height. Leaves and flowers similar to Pacific ninebark except the ovaries are densely hairy. May be confused with Douglas maple which has opposite leaves, or sticky currant, which has leaves that are sticky to the touch. Often associated with ponderosa pine and Douglas-fir forests at low to mid-elevation.
7. **Thin leaf huckleberry** is an erect shrub 3 to 5 feet high. Leaves 1 to 2 inches long, half as wide, thin and pale green on both surfaces, smooth or occasionally minutely hairy, margins toothed, apex and base both acute. Fruit deep purple to black round berry around 6 mm diameter. Twigs slender, green and ridged. Found on dry to moist sites, sun or shade. Similar, and often found with oval-leaved huckleberry which has entire (smooth) rather than toothed leaves.
8. **Blue Elderberry** is a tall deciduous shrub, sometimes tree-like, up to 20 feet in height. Twigs with a soft pith inside. Leaves opposite, pinnately compound, the 5-9 leaflets sharply serrate and strongly uneven at the base. Flowers small, white, flat-topped cluster. Fruit a blue-black berry covered with a white powdery bloom. This species could be confused with red elderberry which differs by having flowers in a spike and red-purple fruit. Found mostly on moist, well-drained sites in the sun; sea level to 9,000 ft.
9. **Red Elderberry** is a tall deciduous shrub, sometimes tree-like, up to 20 feet in height. Twigs with a soft pith inside. Leaves opposite, pinnately compound, the 5-7 leaflets sharply toothed and often uneven at the base. Flowers small, white, and clustered into a long spike. Fruit is a berry, most often red in color but sometimes purplish-black or yellow. Similar to blue elderberry which has a flat-topped flower cluster and a blue-black berry.
10. **Western Wormwood** is an aromatic perennial herb, 1 to 3 feet in height. Leaves mostly 3-11 1-4 in cm long, variable in shape but most often with 3-5 narrow lobes, white hairy beneath, sometimes above as well. Flowers small and arranged in a loose, narrow flower cluster, 2-12 inches long. May be confused with Douglas' wormwood which has wider leaves and is usually found in moister habitats. Also similar to Riverbank wormwood which occurs only near streams and outwash areas.

11. **Mugwort** is a large perennial herb 2 to 5 feet tall, usually found in large colonies in wet areas, ditches, or drainages. Leaves are evenly-spaced, 0.4 to 4.0 inches long, the upper leaves are narrowly elliptical, the lower widely oblanceolate, often coarsely 3 to 5 lobed near the leaf tip, 0.8 to 1.0 inches wide, green above, covered with dense white hair beneath. Differs from western wormwood in having wider lower leaves and in its generally damp habitat.
12. **Evening Primrose** is a large biennial with elliptical leaves up to 10 inches long in a dense rosette the first year. The large (>3ft) flowering stalk with long red-tinged elliptical leaves and large bright yellow four-petaled flowers forms in the second year. Both the leaves and stem are densely hairy, and the hairs often have red, blister-like bases. Usually found in moist, sunny habitats, like seeps or meadows.
13. **Snowberry** is a shrub, 1.5 to 5 feet in height with a solid brown pith. Bark: shreddy, brownish. Young twigs: hairy. Leaves opposite, elliptical, 0.4 to 1.4 inches long and half as wide. Flowers (May-June), the petals white with a pink tube. Fruit a white berry. Common snowberry differs by having a hollow pith. Trailing snowberry is a trailing shrub; and Utah honeysuckle has larger leaves and a solid white pith. Coralberry is similar to common snowberry except the fruit is a red berry. All are members of the honeysuckle family.
14. **Red Alder** is a deciduous tree up to 65 feet tall with dark green leaves 2.4 to 4.7 inches long. The leaves are coarsely toothed, with smaller teeth on the leaf margins, and the leaf veins are also tightly inrolled. Red alder is a common tree in damp situations and is a frequent colonizer of clearings, especially following clearcuts in coniferous forests.
15. **Skunkbush** is a small, diffusively-branched shrub, 1.6 to 3.3 feet tall. The tips of the branches often droop down almost to ground level. The leaves are alternate, compound, with three leaflets, each of which is 3-lobed. The leaves resemble those of poison oak, but the leaflets of skunkbush are smaller, more hairy, and much more deeply-lobed. The leaves of skunkbush also emit a strong, ill-scented odor when crushed. However, if unsure, DO NOT crush the leaves of a shrub with three leaflets to determine the odor. Skunkbush is usually found on dry, open, brushy hillsides, while poison oak prefers damp or shaded forested areas and riparian habitats. Skunkbush is found throughout the southwest, from California and Arizona north to Colorado and Idaho.
16. **Spreading dogbane** is a perennial herb characterized by its opposite leaves with smooth margins and red stems with milky juice. The simple leaves are oblong or egg-shaped, dark green above and pale beneath; 2-3 in long. The plant grows 1-4 feet high and has wide-spreading branches that give the plant an awkward appearance. It flowers throughout the summer; pinkish with a pink stripe in the center. Pods are long and narrow, in pairs. Young milkweed may be confused with dogbane, but differs in having larger, thicker leaves, hairy on the under surface. If evident, milkweed flowers are showy and the pods are large. Dogbane prefers the edges of dry woods from Canada to Mexico, but is also found in dry fields and thickets.

**Phase 3 Field Guide – Ozone Bioindicator Plants (combined), Version 5.1
October, 2011**

OZONE BIOINDICATOR PLANTS
BioSite Characteristics – West

This sheet must be completed only if you have *not* entered this same information on the Bioindicator Plot ID screen.

To be filled out by the FIELD CREW or Cooperator: Refer to Field Guide for code definitions.

STATE	COUNTY	FIELD ID	SPLITPLOT ID ¹	MONTH	DAY	YEAR	CREW ID	QA STATUS
								Standard QA Check

¹SPLITPLOT ID refers to the number of locations (1 or 2) used for each FIELD ID. A separate sheet should be used for each location.

OZONE SAMPLE KIND:	
	Initial biosite establishment on the FIA ozone grid. (Data collection in a previously empty polygon)
	Remeasurement of a previously established biosite, or replacement site within 3 miles of the previously established site
	Remeasurement when the replacement site is more than 3 miles from the previously established biosite.

√ Please put a check mark beside the correct information. Please complete all data fields.

BioSite size: (PLOT SIZE)	TERRAIN POSITION:
<input type="checkbox"/> ≥ 3.0 acres (1.2 hectares)	<input type="checkbox"/> Ridge top or upper slope
<input type="checkbox"/> 1 to 3 acres (0.4 – 1.2 hectares)	<input type="checkbox"/> Bench or level area along a slope
<input type="checkbox"/> Other: please describe	<input type="checkbox"/> Lower slope
	<input type="checkbox"/> Flat land unrelated to slope
	<input type="checkbox"/> Bottom land with occasional flooding

ASPECT: 000° = no aspect; 360° = N aspect	ELEVATION: record estimate in feet or meters	
Record to nearest degree =	Feet =	Meters =

BioSite Wetness (PLOTWET):	SOIL DRAINAGE:	SOIL DEPTH:
<input type="checkbox"/> Wet site Ex: riparian zones, bottomland.	<input type="checkbox"/> Well-drained	<input type="checkbox"/> Bedrock not exposed
<input type="checkbox"/> Moderately dry Ex: meadow, NE slopes.	<input type="checkbox"/> Wet	<input type="checkbox"/> Bedrock exposed
<input type="checkbox"/> Very dry Ex: exposed ledge, desert, alpine.	<input type="checkbox"/> Excessively dry	

DISTURBANCE: Disturbance on the site or in localized areas where the bioindicator plants are growing.
<input type="checkbox"/> No recent or significant disturbance; Do not count disturbance >3 years old.
<input type="checkbox"/> Evidence of overuse; Human activity causing obvious soil compaction or erosion.
<input type="checkbox"/> Evidence of natural disturbance including fire, wind, flooding, grazing, pests, etc.

Fill in below all that apply. Check here if geographic coordinates were obtained from a topographic map:

GPS UNIT:	GPS DATUM:	GPS SERIAL NUMBER:
Latitude =		GPS ERROR =
Longitude =		NUMBER OF READINGS =
GPS ELEVATION =		GPS FILE NAME =
EASTING:	NORTHING:	+/-Error(ft):
		Grid Zone:

Comments: Include information on additional species in the area, safety, directions, or additional site characteristics that may be useful.

File this completed data sheet with the bioindicator site map and driving directions in the ozone plot files for your State or Region.

OZONE BIOINDICATOR PLANTS
Data Sheet for Directions and Mapping for the Bioindicator Site Location

Use the table on the back of this sheet to document directions (mileage and key landmarks) to the ozone biosite.

To be filled out by the FIELD CREW or Cooperator: Refer to Field Guide for code definitions.

STATE	COUNTY	FIELD ID	SPLITPLOT ID ¹	MONTH	DAY	YEAR	CREW ID

¹SPLITPLOT ID refers to the number of locations (1 or 2) used for each FIELD ID. A separate sheet should be used for each location.

Include the following information on the map:

1. Location of the site relative to some obvious and permanent marker.
2. Road names and distances as needed.
3. North arrow.
4. Species codes and approximate location of plant groupings used for the ozone injury evaluations.
5. Location and distance to two major roads; distance and direction to two major towns.
6. Gazetteer reference page if available.

GPS UNIT:	GPS DATUM =	GPS SERIAL NUMBER:
Latitude =	GPS ERROR =	
Longitude =	NUMBER OF READINGS =	
Elevation =	GPS FILE NAME =	
EASTING:	NORTHING:	+/-Error(ft): Grid Zone:

Return the original of this map to the corresponding Biosite Folder so that it can be used by audit and regular crews in subsequent visits to the plot. Mail a copy to the National Indicator Field Advisor the year that the site is established

OZONE BIOINDICATOR FOLIAR INJURY DATA SHEET
Pacific Northwest and Interior West and Plains States

STATE	COUNTY	FIELD ID	SPLITPLOT ID	MONTH	DAY	YEAR	QA STATUS
							___ Standard ___ QA Check

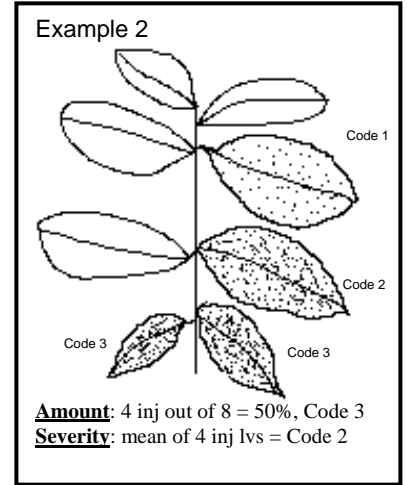
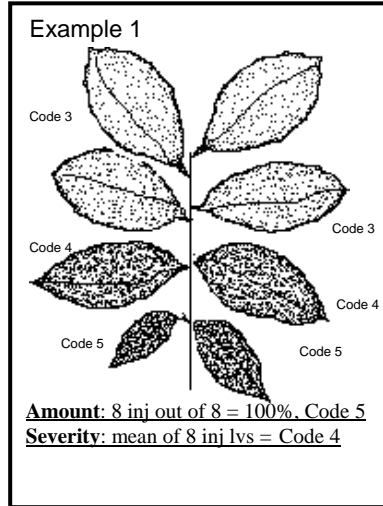
Code	Species
122	Ponderosa pine
746	Quaking aspen
924	Scouler's willow
116	Jeffrey pine
351	Red alder
969	Snowberry spp.
905	Ninebark
906	Pacific Ninebark
965	Huckleberry
960	Blue elderberry
961	Red elderberry
907	Western wormwood
908	Mugwort
909	Skunk bush
366	Spreading dogbane
968	Evening primrose

Amount of Injury – % of leaves injured relative to the total leaf number

Severity of Injury – Average severity of symptoms on the injured leaves

Code	Scale
0	No Injury
1	1-6%
2	7-25%
3	26-50%
4	51-75%
5	>75%

Add Injury Location and Injury Type Codes to the bottom of this sheet under each Species Column. Refer to the FIA Field Guide for all codes and definitions.



Plant	Species Code		Amount	Severity	Species Code	Amount	Severity	Species Code	Amount	Severity	Species Code	Amount	Severity
	Amount	Severity											
1													
2													
3													
4													
5													
6													
7													
8													
9													
10													
11													
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Did you collect 3 leaves that clearly show ozone stipple, for each injured species?

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Notes:

OZONE BIOINDICATOR PLANTS
Data Sheet for the Voucher Leaf Samples

To be filled out by the FIELD CREW or Cooperator: Refer to the Field Guide for code definitions.

STATE	COUNTY	FIELD ID	SPLITPLOT ID ¹	MONTH	DAY	YEAR	CREW ID	QA STATUS
								Standard QA Check

¹SPLITPLOT ID refers to the number of locations (1 or 2) used for each FIELD ID. Separate sheets should be used for each location.

Crew Name(s)	e-mail address	Phone number

Fill in the required codes. Code definitions are in the Field Guide. For quick reference, see below.

Bioindicator Species Code or Common Name	Injury Location	Injury Type	Is the leaf sample injury close to 100% ozone stipple (√), or is some other upper-leaf-surface injury also present (e.g., insect or fungal lesion)?
1 st			Close to 100% _____ Estimated percent other _____
2 nd			Close to 100% _____ Estimated percent other _____
3 rd			Close to 100% _____ Estimated percent other _____
4 th			Close to 100% _____ Estimated percent other _____

List species evaluated at this location that did not have ozone symptoms: →

Notes: Add notes on the leaf samples, plot conditions, safety, and weather as needed.

0122 Ponderosa pine	0116 Jeffrey pine	0746 Quaking aspen	0924 Scouler's willow	0351 Red alder
0960 Blue elderberry	0961 Red elderberry	0965 Huckleberry	0905 Ninebark	0906 Pacific ninebark
0907 Western wormwood	0908 Mugwort	0909 Skunk bush	0968 Evening primrose	0969 Snowberry
		0999 Unknown	0998 Supplemental species: write out common name	

Injury Location Codes:
 1= greater than 50% of the injured leaves are younger leaves (broadleaf) or current whorl (pine)
 2= greater than 50% of the injured leaves are mid-aged or older (broadleaf) or on whorls 1 year and older (pine)
 3= injured leaves are all ages.

Injury Type Codes:
 1= greater than 50% of the injury is upper-leaf-surface stipple (broadleaf) or chlorotic mottle (pine).
 2= greater than 50% is not stipple (tan flecks, bifacial or general discoloration), or something other than chlorotic mottle (pine)
 3= injury is varied or difficult to describe.

Mail this sheet with the leaf samples to:

[Do not write below this line]

Pat Temple
 USDA FS, PSW Experiment Station
 4955 canyon Crest Drive
 Riverside, CA 92507

QA/QC PERSON: To be filled out by the National Ozone Field Advisor or Regional Expert. ✓

Date checked	Date rechecked	Sample condition			Plot Status	
		GOOD Easy to read – ID obvious	FAIR	POOR Samples unreadable or not labeled correctly	(+ozone)	(-ozone)

Bioindicator Species	Positive for ozone	Negative for ozone	Explanation

Additional questions for the data collector:

Appendix 20.C Detailed Procedures for Handling Leaf Vouchers

Detailed Procedures For Handling Leaf Vouchers For Broadleaf Species

Leaf Collection in the Field

1. Collect 3 leaves from each species showing ozone injury symptoms
 - These 3 leaves should be from different plants, if possible
 - These 3 leaves should show obvious injury rather than the range of different symptoms you may have observed
2. Once the leaf vouchers are cut from a plant they should be placed immediately into a plant press
 - Each leaf should have its own space on the blotter paper – do not overlap leaves
 - Each leaf should be marked with the date and the FIELD ID in case they get shuffled
 - Leaves that are not put into a plant press immediately will wrinkle and break easily when handled
 - Leaves that are laid on top of each other will “bleed” such that all overlapped leaves become murky and the ozone injury symptom is no longer visible
3. Before you leave the plot where you have collected voucher leaves, fill out the leaf voucher data sheet and complete the Bioindicator Plot Identification Screen on the PDR. There is important information on the voucher sheet and on the PDR screen that you need to record while you are standing on the biosite. You will not remember these details later on, so take the time to get it right while you are on the plot.
4. Pressed leaves can be removed from the plant press after 36 hours. Once they are flat and dry they can be kept in mailing envelopes, folders, or newspaper until you have time to mail them in to be validated.

Leaf Preparation for Mailing

5. Each mailing envelope may contain leaves from ONE or SEVERAL biosites. Use common sense to decide how many leaves will fit comfortably into each envelope. Mark the outside back of each envelope with the list of hex numbers that have been included. Labels are provided for this, or you can write the numbers on with pen.
6. Each pile of leaves from each plot should be placed on top of the corresponding voucher data sheet. It is very helpful to include an additional blank piece of recycled paper or newspaper to help keep larger piles of leaves separated from each other.
7. At least 1 of the 3 leaves (and preferably all 3 leaves) you have selected for each species should have a petiole label with the FIELD ID written on it. This will prevent data loss if the leaf pile is dropped and separated from its corresponding voucher data sheet when the leaves are removed from the envelope.
 - Do not put large piles of leaves into a single mailing envelope. Help minimize human error by mailing the leaves and data sheets 1 plot per envelope, or no more than 5 plots per envelope, depending on how many species were injured and how many leaves and data sheets must be mailed in.
 - Use the 10”x12” size mailing envelopes that are provided at training. If you make a substitution, make sure it is approximately the same size or larger.
 - If you choose to mail larger piles in a single large mailing container, please use newspaper or manila folders to separate vouchers and their corresponding data sheets by FIELD ID.
 - The only way it is safe and acceptable to mail unmarked leaves (no petiole label), is if each group of leaves from each biosite is contained in a separate mailing envelope (10”x12”) that contains the corresponding voucher data sheet and is clearly marked with the appropriate FIELD ID number.
8. If you have the time and the resources, supply a cover sheet that lists all of the FIELD ID numbers you have included in your mailing(s). It is also extremely helpful if the leaves and vouchers are organized by FIELD ID number in increasing or decreasing order, e.g., XXXX328, XXXX422, XXX521, etc.
9. Feel free to ask for the return of your leaves and a copy of the voucher data sheets. This will only be done on request. Remember that the validation process begins in mid-September and may take until December to complete. If you have a time constraint and need a quick response, please note this on the OUTSIDE of the mailing envelope so that it will be noticed upon arrival.

10. If you have mailed extra leaves (>3 per species) for any purpose, please attach a handwritten note explaining what you have done. Clearly mark which 3 leaves should be used to validate the ozone injury at each site. Explain clearly what additional review of leaf samples is of interest to you and include a separate voucher data sheet for this purpose.
11. If you have mailed in samples of supplementary species that are not on the official bioindicator list, please keep these separated by FIELD ID number or off grid location and provide a separate data sheet for these extra species, 1 sheet per species. If possible, provide GPS coordinates of any off-grid sampling locations. Species found within approximately 3 miles of the established biosite are still on the grid and do not require additional GPS data.

Proper Handling of Leaf Vouchers for Pine Species

1. Collect 3 leaves or 2 small branch samples from each species showing ozone injury symptoms.
 - The 3 leaves or 2 branch samples should be from different plants, if possible
 - The 3 leaves or 2 branch samples should show obvious injury rather than a range of different injury symptoms
2. Place the leaf or branch samples immediately into a plant press
 - Do not overlap leaves or needles on the blotter paper
 - Mark each sample with the date and FIELD_ID
 - Before leaving the site, fill out the top half of the voucher data sheet
3. Pressed leaves or branch samples can be removed from the plant press after 36 hours. Once they are flat and dry they can be stored in mailing envelopes or newspaper until they are mailed.
4. Branch samples that are not pressed immediately **MUST BE KEPT COOL** and **MUST BE MAILED** promptly so that they remain in good condition for the QA check on injury symptoms.
 - Keep samples out of the sun and other hot situations
 - Do not leave unprotected samples overnight in vehicles
 - Keeping samples in a cooler with ice provides adequate protection for up to 1 week
 - Refrigerate any samples that cannot be mailed promptly
5. Label each mailing envelope with the FIELD ID's of the samples that are placed in each envelope.
 - Double-check the mailing address before mailing – use the mailing labels provided at Training.
 - Double-check that all samples are labeled with their corresponding FIELD ID number and that the voucher data sheet is mailed with the corresponding leaf or branch samples
 - If you want the QA findings returned to you, please note this on the voucher data sheet and provide appropriate contact information
6. Make every effort to mail samples within 1 week of collection. If this is not possible, keep pressed leaves in a cool and dry location. Keep pine branch samples clean and dry in a plastic container in the refrigerator.

Appendix 20.D Site Ranking and Biosite Selection

BEST SITE

Crews are encouraged to use good judgment when selecting the best possible biosite for ozone injury evaluations. In all regions, the BEST biosite is a 1-3 acre opening with 3 or more bioindicator species and 30 plants of each species. Each biosite is identified by a unique, 7-digit field identification number (FIELD ID). Each biosite may consist of one or two locations on the ground. If two locations are selected, they must be within 3 miles of each other. The two locations have the same FIELD ID and are identified as SPLITPLOT ID = 1 and SPLITPLOT ID = 2. Crews must evaluate 3 species and up to 30 plants per species using whatever plants are be found at 1 or 2 locations. If more than 3 bioindicator species are found on a site, crews are strongly encouraged to evaluate at least 10 plants of these additional species.

ACCEPTABLE SITE

An ACCEPTABLE biosite is defined as one with 30 plants of only two species or fewer than 30 plants of 3 or more species. When crews are forced to settle for an acceptable biosite, they should take the time to look for and establish a second location in the current or subsequent year to increase species and plant counts.

MARGINAL SITE

Biosites of MARGINAL quality lack an acceptable number of species, or plants per species as described in the following table. Data are collected from marginal biosites only after the crew has invested ½ day without finding a biosite that meets BEST or ACCEPTABLE site rankings. Marginal sites must be replaced the following year.

SITE RANKING ↓	Number Of Species At Each Site	Preferred Number of Plants of Each Species	Acceptable Number of Plants of Each Species	Minimum Number of Plants of Each Species
BEST: one location	Species1	30	25-30	10
Use two locations to obtain the best species and plant counts, as needed.	Species2	30	10-30	10
	Species3	30	10-30	10
	SpeciesN	10-30	10-30	10
ACCEPTABLE	Species1	30	25-30	20
One or two locations	Species2	30	10-30	10
MARGINAL	Species1	30	25-30	25

Decision Table for Site Selection		Crew Action
First Choice		Maintain this biosite for as many years as possible
Second Choice		Maintain biosite. Look for additional species, plants, or a second location each year
Third Choice		Replace biosite or look for additional species and plants or second location
Fourth Choice (use site for only 1 year)		Biosite must be replaced by one with acceptable species and plant counts asap

Biosites ranked BEST are relatively easy to find in eastern forest types and relatively difficult to find in some of the drier, western forest types. In western regions, SPLITPLOT ID 1 may be all one species (e.g., *Populus tremuloides* or *Pinus ponderosa*) necessitating a search for a second location, i.e., SPLITPLOT ID 2. The second location may be some distance from the first and may have very different bioindicator species that are growing under very different soil/site conditions. Each location is described separately in a separate plot file even though both SPLITPLOT ID 1 and SPLITPLOT ID 2 are assigned the same 7-digit FIELD ID.

Average number of species per biosite for each FIA Region as of 2002			
Northeast	3.5	Interior West	2.5
North Central	3.5	Pacific Northwest	2.5
Southern	3.0		

Appendix 20.E Supplemental Bioindicator Species

The species listed in the following table may be used as the 4th, 5th, or 6th species at a biosite. With the exception of green ash (541), erect dogbane (366), thornless blackberry (915), and tall milkweed (365), these species may not be used to meet the requirement for 3 species per biosite as described in Section 20.2.5 in the Field Guide. If a location has only supplemental species, the crew must find a new site that meets the biosite selection guidelines.

	Scientific name	Common name	Region	Species code ¹	Bio Rank ²
Tree spp.	<i>Acer saccharinum</i>	Silver maple	East	998	3
	<i>Alnus rugosa</i>	Speckled alder	East	998	1
	<i>Cercis canadensis</i>	Red bud	East	998	1
	<i>Fraxinus pennsylvanica</i>	Green ash	East	541	2
	<i>Oxydendron arboreum</i>	Sourwood	East	998	3
	<i>Prunus americana</i>	American plum	East and West	998	3
	<i>Tilia americana</i>	Basswood	East	998	3
Woody	<i>Apocynum cannabinum</i>	Erect dogbane (Indianhemp)	East and West	366	2
	<i>Campsis radicans</i>	Trumpet creeper	East and West	998	3
	<i>Cephalanthus occidentalis</i>	Common buttonbush	East and West	998	3
	<i>Lindera benzoin</i>	Spicebush	East	998	3
	<i>Rhus copallinum</i>	Winged sumac	East	998	2
	<i>Rubus occidentalis</i>	Black Raspberry	East	998	3
	<i>Rubus canadensis</i>	Thornless (smooth) blackberry	East	915	1
	<i>Sambucus canadensis</i>	American (common) elderberry	East and West	998	1
	<i>Viburnum lentago</i>	Nannyberry	East	998	3
	<i>Vitis labrusca</i>	Northern Fox grape	East	998	1
Non-woody	<i>Apios americana</i>	Common ground nut	East	998	1
	<i>Asclepias exaltata</i>	Tall milkweed (poke milkweed)	East	365	1
	<i>Asclepias incarnata</i>	Swamp milkweed	East and Plains	365	2
	<i>Asclepias speciosa</i>	Showy milkweed	West and Plains	998	3
	<i>Eupatorium rugosum</i>	White snake root	East	998	1
	<i>Helianthus divaricatus</i>	Woodland sunflower	East	998	3
	<i>Rudbeckia laciniata</i>	Cutleaf coneflower	East and West	998	1
	<i>Silphium perfoliatum</i>	Cup plant	East	998	3
	<i>Verbesina occidentalis</i>	Crownbeard	East and Plains	998	1
	<i>Vernonia noveboracensis</i>	Iron weed	East	998	3

¹New ozone software may accept the 998 species code for supplemental species and allow crews to enter injury amount and severity data for trend analysis. Record the species common name in the plot notes and on the voucher data sheet. *Fraxinus* spp. (541) includes both white and green ash; *Apocynum* spp. (366) includes both spreading and erect dogbane. *Rubus* spp. (915) includes both common and thornless blackberry; *Asclepias* spp. (365) includes both common and tall milkweed.

²Rank 1 = species ozone sensitivity has been confirmed in field and exposure chamber; Rank2 = species sensitivity has been confirmed in field or exposure chamber; Rank3 = conflicting evidence of species ozone sensitivity.