

Adaptation and colonization history affect the evolution of clines in two introduced species

Stephen R. Keller¹, Dexter R. Sowell¹, Maurine Neiman², Lorne M. Wolfe³ and Douglas R. Taylor¹

¹Department of Biology, University of Virginia, Charlottesville, VA 22904-4328, USA; ²Department of Biology, University of Iowa, Iowa City, IA 52242; USA; ³Department of Biology, Georgia Southern University, Statesboro, GA 30460, USA

Summary

Author for correspondence: Stephen R. Keller Tel: +1 434 982 2518 Email: srk3d@virginia.edu

Received: *17 February 2009* Accepted: *9 April 2009*

New Phytologist (2009) **doi**: 10.1111/j.1469-8137.2009.02892.x

Key words: adaptation, founder effect, invasion, migration, neutrality, range expansion, selection, *Silene*.

• Phenotypic and genetic clines have long been synonymous with adaptive evolution. However, other processes (for example, migration, range expansion, invasion) may generate clines in traits or loci across geographical and environmental gradients. It is therefore important to distinguish between clines that represent adaptive evolution and those that result from selectively neutral demographic or genetic processes.

• We tested for the differentiation of phenotypic traits along environmental gradients using two species in the genus *Silene*, whilst statistically controlling for colonization history and founder effects. We sampled seed families from across the native and introduced ranges, genotyped individuals and estimated phenotypic differentiation in replicated common gardens.

• The results suggest that post-glacial expansion of *S. vulgaris* and *S. latifolia* involved both neutral and adaptive genetic differentiation (clines) of life history traits along major axes of environmental variation in Europe and North America. Phenotypic clines generally persisted when tested against the neutral expectation, although some clines disappeared (and one cline emerged) when the effects of genetic ancestry were statistically removed.

• Colonization history, estimated using genetic markers, is a useful null model for tests of adaptive trait divergence, especially during range expansion and invasion when selection and gene flow may not have reached equilibrium.

Introduction

It has long been recognized that evolutionary change occurs by both adaptive and nonadaptive mechanisms (Gould & Lewontin, 1979). Accordingly, tests for adaptive divergence have become more rigorous, often involving comparisons of evolutionary rates or divergences relative to some neutral expectation (Lewontin & Krakauer, 1973; Kimura, 1980; Spitze, 1993; Vasemagi, 2006; Keller & Taylor, 2008).

Clines in allelic frequencies and quantitative traits along environmental gradients are often interpreted as evidence for adaptive evolution (Endler, 1977; Eanes, 1999). The near-ubiquity of clines in traits along gradients, such as latitude, has been used to identify major biogeographical trends, such as Bergmann's rule (Blackburn *et al.*, 1999) and Allen's rule (Scholander, 1955). Classic studies of clinal variation in plants have played a particularly noteworthy role in our understanding of local adaptation (Clausen *et al.*, 1948; Antonovics & Bradshaw, 1970), with plant populations frequently showing covariance between environmental gradients and phenology, morphology, physiology and life history (for example, Drezner, 2003; Stinchcombe *et al.*, 2004; Ingvarsson *et al.*, 2006; Savolainen *et al.*, 2007).

In the study of adaptation along a gradient, it is important to recognize and estimate other evolutionary processes that generate correlations between quantitative traits and geography or the environment. For example, the balance between genetic drift and migration can generate patterns of isolation by distance across geographical or environmental gradients that closely mimic adaptive clines (Vasemagi, 2006). Range expansion and colonization history can also leave a geographical signature of clinal variation through a series of successive founder events (Amsellem *et al.*, 2000; Clegg *et al.*, 2002), or when genetically distinct demes expand into a new range (Hewitt, 2000). A classic example of such 'demic diffusion' comes from clinal gradients in allelic frequencies among European humans, a consequence of the rapid expansion of Neolithic farmers into Europe (Cavalli-Sforza *et al.*, 1993).

Similarly, clines may also arise during the colonization phase of a species invasion. The colonization history of many invasive species is accompanied by multiple introductions from different native range sources (Wares et al., 2005; Dlugosch & Parker, 2008). Because founding events can generate genetic structure and deviations from the equilibrium balance between gene flow, drift and selection (Whitlock & McCauley, 1990, 1999; McCauley et al., 1995), multiple introductions can quickly generate patterns that resemble isolation by distance or clinal variation when sources establish at different points of introduction (for example, Roman, 2006; Keller & Taylor, 2008). Numerous studies of species invasions have reported latitudinal clines for traits such as body size and mass, height, number of stems, flowering time, fecundity and physiology (Johnston & Selander, 1964; Weber & Schmid, 1998; Huey et al., 2000; Kollmann & Banuelos, 2004; Maron et al., 2004, 2007; Leger & Rice, 2007; Montague et al., 2008). Although colonization history is expected to have a major influence on the population and quantitative genetics of introduced species, few studies have attempted to control for colonization history when analysing clines (but see Maron et al., 2004). The implications are that the evolutionary causes of clinal patterns and, in particular, the importance of selection in generating clines are often unclear (Keller & Taylor, 2008).

Silene vulgaris and Silene latifolia (Caryophyllaceae) are weedy herbaceous plants that have a history of post-glacial expansion throughout Europe from divergent refugia and a recent invasion in North America (Baker, 1948; Taylor & Keller, 2007). The inferred routes of expansion in Europe are congruent with the post-glacial history of many European plants and animals (Hewitt, 2000; Petit et al., 2003). In both S. latifolia and S. vulgaris, historical isolation and expansion coincided with phenotypic divergence, generating covariances between geography, climate, genotypes and phenotypes (Mastenbroek et al., 1983; Keller, 2008). During the last 200 yr, both species have colonized North America and rapidly expanded their distributions. The S. vulgaris and S. latifolia systems are thus ideally suited for an empirical evaluation of the extent to which phenotypic clines during range expansion represent adaptive evolution vs demographic and neutral genetic processes.

In this study, we tested for genetic differentiation in phenotypic traits across environmental gradients whilst statistically controlling for selectively neutral processes that can also generate clines (for example, colonization history, gene flow and genetic drift). The approach builds on the tradition of estimating the genetic divergence of traits relative to neutral loci ($F_{\rm ST}$ vs $Q_{\rm ST}$; Spitze, 1993), but is, in practice, more similar to methods that statistically account for back-

ground variation in genetic ancestry that frequently complicates genome-wide association studies (Price *et al.*, 2006). Specifically, we describe the multidimensional genotypic space of neutral loci using principal coordinates analysis (PCO), and remove the covariance between marker loci and phenotypes using multiple regression. Covariance between the residuals and geographical or environmental variables then provides an estimate of adaptive clinal variation in phenotype after the major axes of neutral genetic variation have been removed. We apply this approach to the post-glacial expansion and recent invasion history of *S. vulgaris* and *S. latifolia*, and show that adaptation and neutral/historical processes combine to generate clines in many aspects of the phenotype. In other traits, however, phenotypic trait variation is fully explainable by a null model of neutral genetic ancestry.

Materials and Methods

Silene vulgaris (Moenche) Garcke (bladder campion) and Silene latifolia Poiret (white campion) are short-lived, perennial plants in the family Caryophyllaceae. Silene vulgaris is gynodioecious, with self-compatible hermaphrodites and a mixed-mating system (Glaettli et al., 2006), whereas S. latifolia is dioecious. Both species have evolutionary histories of dynamic range expansion, becoming widespread following post-glacial expansion throughout their native range in Eurasia (Taylor & Keller, 2007). The species were first reported in North America in the early 1800s, probably introduced from Europe as contaminant in clover seed and waste from ocean-going ships that deposited their ballast (soil, sand, gravel, silage) in North American port cities (Martindale, 1876; Baker, 1948; McNeill, 1977). Both species are widespread weeds of disturbed areas in North America (McNeill, 1977; Randall, 2002), where their ranges extend over much of the northern USA and southern Canada, and southwards in the eastern USA at higher elevations.

Seed capsules (maternal families) were collected from both species throughout Europe and North America as part of an ongoing series of experiments focused on elucidating the nature and evolution of invasiveness. We then combined a population genetic analysis of neutral variation with common garden experiments to measure life history traits that impact on fitness and are likely to experience variation in the direction and intensity of selection in response to regionally varying climate. These included size and growth-related traits that reflect the capacity for light capture and carbon fixation (height, number of stems, number of leaves), phenological traits that reflect the schedule of life history events (germination time, time to flowering) and reproductive traits that reflect effort expended towards producing sexual progeny (number of flowers, weeks spent flowering, number of fruits). Because the functional values of these traits often vary with environmental conditions, especially temperature, precipitation and photoperiod, they have been used extensively in



Fig. 1 The geographical distribution of genotyped samples of *Silene latifolia* and *Silene vulgaris* in Europe and North America. Colours represent contours in genetic space using marker loci (interpolated using the inverse distance weighting method), with panels showing the first two principal coordinates (PCO1 and PCO2) obtained from amplified fragment length polymorphism (AFLP) genotyping. In all plots, cooler colours represent higher PCO scores and warmer colours represent lower PCO scores. The scaling of colour is identical across continents (within species) to facilitate interpretation of the introduction history.

previous studies of clines in plants (for example, Kollmann & Banuelos, 2004; Maron *et al.*, 2004; Stinchcombe *et al.*, 2004; Etterson *et al.*, 2008; Samis *et al.*, 2008).

Replicated experiments at different garden sites are essential when estimating evolved differences among ranges (for example, Williams *et al.*, 2008). Accordingly, the experimental design for each species involved one garden planted in Ontario, Canada and one garden established in Virginia, USA. These sites span most of the latitudinal breadth of each species in eastern North America. However, the two sites within Ontario and within Virginia were not identical, and the design and traits measured were different for each species, and so the two sets of gardens are treated as different experiments.

Common garden experiments

For the *S. vulgaris* experiment, maternal families were selected from 36 European and 23 North American localities (Fig. 1). Open-pollinated seed capsules were collected from up to seven maternal plants in each location. In all, 100 families from North America and 100 families from Europe were grown in two gardens: a fallow field in Ontario, Canada (45.8642°N, -79.4362°W) and a hay pasture in Virginia, USA (37.8577°N, -78.8208°W). There were *S. vulgaris* populations in the vicinity of each site (~1 km away), but not in the fields themselves. Six weeks before planting, seeds were surface sown in a randomized design into plug trays filled with a standard potting mix. Plants were germinated in growth chambers under misted conditions. The chambers were set to a diurnal photoperiod of 12 h : 12 h (light : dark) and a temperature cycle of 21 and 12° C, and trays were rotated daily within the chambers to reduce position effects. Germination was scored daily and four seedlings per family were randomly chosen for planting into the field (total N = 2 sites × 2 continents of origin × 100 families/continent × 4 plants/family = 1600 plants).

Seedlings were transplanted into the field during spring of 2005. The gardens consisted of four rectangular subplots, each containing 200 plants (one from each family). Seedlings were watered, and those that were lost as a result of transplant mortality < 2 wk post-planting were replaced with a seedling from the same family at the same stage of growth. The existing vegetation was cut back periodically at each site to avoid over-competition.

Weekly censuses were conducted throughout the 2005 growing season and again in 2006 to assay performance and measure traits important to plant fitness. During the first census in which a plant was observed flowering, we measured the height of the tallest stem (cm), the number of primary stems, the number of leaves > 1 cm long and the number of flowers. During each subsequent census, plants were checked for reproductive status (vegetative or flowering) and the number of new flowers and mature fruits. The *S. latifolia* field data are a subset of the data collected from a larger experiment involving common gardens established in Europe and North America. These garden sites included plots that excluded natural enemies with pesticides and control plots. For the current study, we only used data from the control plots in the North American sites. Sixty full sib families were grown in old fields near Queen's University Biological Station in Ontario, Canada (44.5788°N, –76.3804°W) and Mountain Lake Biological Station (37.3603°N, –80.5527°W).

The 60 families were generated using seed collections from 30 European and 30 North American populations. A sample of seeds was grown from two maternal families (capsules) from each population, and a male and a female each from separate families were crossed to yield one full sib family per population. The seeds were sown into a standard potting mix in plastic pots (diameter, 2.5 cm) (trade name 'conetainers'). Seeds were germinated on a mist bench, thinned to single plants and grown to rosettes under ambient conditions in the glasshouse. Germination was poor in some families, leaving us with 28 full sib families from Europe and 30 full sib families from North America.

The sites were planted in May 2006. Plants were cut back to 1 cm of growth above the root crown. The subset of the plots used in this study involved six plants per family (360 plants) in each of the two sites. The sites were divided into 12 subplots with an equal number of North American and European families in each. For planting, the soil plug was extracted from the conetainer and inserted into a small disturbance created with a 15-cm auger at 50-cm intervals. Plants were censused in the spring (late April to early May), summer (late July to early August) and autumn (mid to late October) for a total of five censuses over two seasons. At each census, we recorded the number of basal and stem leaves, heights of all stems to the nearest centimetre, number of flowers open or in bud and the number of fruits. We also calculated the reproductive speed of each plant as the inverse of the census at which we first observed a plant to be reproductive (five for the first census, one for the most recent, zero if never found to be reproductive).

Amplified fragment length polymorphism (AFLP) genotyping

We attempted to genotype one representative from each family in each species (~260 individuals overall) for genetic analysis. However, after losses caused by incomplete germination and mortality, we were able to obtain samples for molecular work from 175 *S. vulgaris* families (92 European, 83 North American) and 44 *S. latifolia* families (22 European, 22 North American). Genomic DNA was isolated from leaf tissue using Qiagen DNeasy plant miniprep kits. Plants were genotyped for AFLP loci following the methods described elsewhere (Keller, 2008). Briefly, we used an Applied Biosystems (Foster City, CA, USA) Plant Mapping Kit in which 50–100 ng of genomic DNA was digested at room temperature overnight with a pair of restriction enzymes (EcoRI and MseI; New England Biolabs, Ipswich, MA, USA) and ligated to doublestranded adapters in a single 10-µl reaction. Preselective and selective amplifications were performed in 10-µl reaction volumes using a touchdown polymerase chain reaction (PCR) protocol suggested by the manufacturer (Applied Biosystems Plant Mapping Kit). We used four primer pair combinations in S. vulgaris [two 5-FAM-labelled pairs (EcoRI: ACA/MseI: CTC, EcoRI: ACA/MseI: CAC) and two TAMRAlabelled pairs (EcoRI: ACC/MseI: CAG, EcoRI: AGC/MseI: CTG)] and three primer pairs in S. latifolia [one 5-FAMlabelled pair (EcoRI: ACA/MseI: CAC), one TAMRA-labelled pair (EcoRI: ACC/MseI: CAG) and one JOE-labelled pair (EcoRI: AGG/MseI: CAA)]. Selective amplification products were sized relative to the ROX-500 standard using GENEMAPPER v4.0 software (Applied Biosystems) on an ABI 3130xl sequencer. All samples were normalized to the sum of the sample signal within GENEMAPPER, calculated across the entire project. Polymorphic peaks > 50 relative fluorescence units (RFUs) were scored within a size range of 100-400 bp (S. vulgaris) or 50-400 bp (S. latifolia). Automated scoring was manually verified and peak presence/absence was converted to binary (0/1) coding for analysis. In total, 267 polymorphic AFLP loci were scored for S. vulgaris and 133 for S. latifolia.

Data analysis

We performed an initial analysis to remove variance attributable to site and plot effects using repeated measures analysis of variance (ANOVA) (PROC MIXED: SAS, 2004). For the S. vulgaris data, garden site and year were treated as fixed effects, whereas plot within each site was a random effect; individual plants were the repeated subject effects. Traits were either natural logarithmically transformed (time to germination, time to flowering, number of leaves at flowering, number of flowers, flowering period, number of fruits) or square root transformed (height, number of stems) before analysis to improve normality and homoscedasticity. For S. latifolia, traits were either logarithmically transformed (basal leaves, stem leaves, number of flowers, number of fruits) or square root transformed (average stem height, number of stems). Residuals from these ANOVAs were exported for further analysis.

We estimated the genetic similarity among AFLP genotypes using the first two eigenvectors from PCO to ordinate one individual from each family in continuous multilocus genotype space. Principal coordinates were computed on the binary AFLP data matrix using R PACKAGE v4.0 (http://www. bio.umontreal.ca/Casgrain/en/labo/R/v4/index.html). Genetic distances among individuals were defined as 1 - Jaccard'ssimilarity index J_{ip} which calculates the similarity based on shared bands among individuals and ignores shared absences $[J_{ij} = a/(a + b + c))$, where *a* is the number of bands shared by individuals *i* and *j*, *b* is the number of bands in *i* but not *j*, and *c* is the number of bands in *j* but not *i*]. We then combined each family's scores from the first two PCO axes with the phenotypic data (residuals from the above ANOVAs). To avoid pseudoreplicating the PCO results among individuals within sibships, we used family means for the phenotypic data, and hereafter treat families as the units of observation in all subsequent analyses.

We associated each family with the latitude from which it was collected, as well as climate-based environmental variables. Environmental variables were obtained from the WorldClim dataset of interpolated global climate (Hijmans et al., 2005), and we focused specifically on eight bioclimatic ('Bioclim') variables derived from long-term observations of temperature and precipitation (1950-90) mapped to a resolution of 5 arc-min (0.083°). The eight variables were the mean annual temperature, minimum temperature of the coldest month, maximum temperature of the warmest month, temperature seasonality (SD across months \times 100), mean annual precipitation, mean precipitation during the driest month, mean precipitation during the warmest quarter year and precipitation seasonality (CV). We chose these eight variables because they represent more biologically meaningful descriptors of the conditions that plants experience during the growing season and overwintering relative to annual means. We extracted values for each climate variable for each of our sites using a geographical information system (GIS) and pooled observations across species and continents. We then summarized the major trends in climate using a principal components analysis (PCA) to reduce the dimensionality of the environmental data (PROC PRINCOMP: SAS, 2004). Spearman rank correlation was used to assess correlations within each continent between geography (latitude, longitude), climate (Prin1, Prin2) and neutral genetic variation (PCO1, PCO2). To visually portray how climate and ancestry varied geographically, we mapped the variation in PCA and PCO interpolated using the inverse distance weighting method (Chang, 2008).

We tested for phenotypic diversification along genetic and environmental clines within Europe and North America using general linear models estimated using restricted maximum likelihood (PROC MIXED: SAS, 2004). Although the models did not contain random effects, we chose to use PROC MIXED for its maximum likelihood estimation, which permits the calculation of measures of model fitting to the data. Accordingly, we adopted a model selection approach, using the Akaike information criterion (AIC_c), to assess the fit among three different candidate models (Johnson & Omland, 2004). In the first model (climate-only, or 'C'), we regressed each phenotypic trait on the first two PCA axes (Prin1 and Prin2) to test for clines with climate. In the second model (ancestry-only, or 'A'), we regressed traits on the first two PCO

axes (PCO1 and PCO2) from the AFLP data to detect phenotypic associations with neutral loci that might have accumulated as a result of drift, migration, founder effects and other selectively neutral aspects of colonization history. Finally, we tested for phenotypic clines against the null expectation of ancestry by including as predictor variables the climate variables (Prin1 and Prin2) and the neutral genetic variables (PCO1 and PCO2) in the same model (climateancestry, or 'CA'). This last model was designed to test for clines with environmental variables that persist after controlling for nonadaptive processes that may have generated clines at neutral loci. It should be noted that the use of PCO to control for history assumes selective neutrality of AFLP loci; thus, any linkage disequilibrium between AFLPs and nearby loci under selection may remove some non-neutral variance. For each trait, we retained the model with the lowest AIC_c score, and reported the difference in fit (ΔAIC_{c}) between the best and next best models (Johnson & Omland, 2004). We also replicated the AIC_c model selection analysis using latitude instead of the WorldClim variables for comparison with other studies that traditionally report latitudinal clines.

Summarizing climate variation with PCA is useful because it avoids statistical problems of collinearity and multiple testing. However, there is no guarantee that the PCA axes represent the combination of environmental variables to which the plants are adapting. Therefore, as a complementary approach to model selection using PCA, we also used stepwise multiple regression to build models that contained the set of individual climate variables and PCO axes that best explained the phenotypic variance.

Results

Associations among climate, geography and genetic ancestry

The eight WorldClim climate variables were summarized by the first two principal components (Prin1 and Prin2), which collectively explained 69% of the total variance (Table 1). These axes describe major climate trends on each continent: large values of Prin1 are cool and wet, with low precipitation

 $\label{eq:tables} \begin{array}{l} \textbf{Table 1} & \mbox{Principal components analysis of eight climate variables from the WorldClim dataset} \end{array}$

	Prin1	Prin2
Mean annual temperature	-0.26057	0.43095
Minimum temperature of coldest month	-0.27154	0.55096
Maximum temperature of warmest month	-0.10822	-0.08102
Temperature seasonality	0.19115	-0.52623
Mean annual precipitation	0.37404	0.30430
Precipitation driest month	0.49674	0.28180
Precipitation warmest quarter	0.51655	0.01367
Precipitation seasonality	-0.39578	-0.23463

	Europe		North America	
	Latitude	Longitude	Latitude	Longitude
WorldClim variables:				
Prin1	0.32195*	-0.11862	-0.12117	0.81901****
Prin2	-0.21453	-0.62254****	-0.77199****	0.40126**
Silene latifolia AFLP:				
PCO1	-0.47134*	-0.79447****	0.02428	0.56748**
PCO2	-0.18874	-0.14229	0.17335	-0.64201**
Silene vulgaris AFLP:				
PCO1	-0.27129**	0.54292****	0.06259	-0.28222**
PCO2	-0.30230**	0.21518*	0.20545*	-0.22477*

Table 2 Spearman rank correlation coefficients between latitude/longitude, climate and genetic ancestry

Prin1 and Prin2 are the first two eigenvectors from a principal components analysis of eight climate variables obtained from the WorldClim dataset. PCO1 and PCO2 are the first and second eigenvectors from a principal coordinates analysis of multilocus amplified fragment length polymorphism (AFLP). *, P < 0.05; **, P < 0.01; ****, P < 0.001; ****, P < 0.001.

seasonality; small values are warm and dry, with more seasonal precipitation (Table 1). In Europe, this climate axis correlated positively with latitude, whereas, in North America, it correlated positively with longitude (Table 2; Fig. S1, see Supporting Information). For Prin2, large values are warm and wet, with low temperature and precipitation seasonality; small values are cold, dry and more seasonal (Table 1). In Europe, Prin2 correlated negatively with longitude, whereas, in North America, Prin2 correlated negatively with latitude and positively with longitude (Table 2; Fig. S1).

Neutral genetic diversity also showed strong geographical gradients on both continents (Fig. 1). In the native range of Europe, demic diffusion during the process of post-glacial expansion was evident as a strong east-west transition zone in central Europe for PCO1 in both species. This pattern generated a highly significant correlation of PCO1 with longitude (both species P < 0.0001, Table 2). Variation in PCO2 was more species specific: S. latifolia showed a genetically distinct region in central Europe, whereas, for S. vulgaris, there was evidence of a cohesive group between the southern Balkans and the area around the Black Sea. The invasion history into North America generated a more patchy spatial distribution of genetic diversity in both species. Nevertheless, the establishment points of the introduced genotypes and subsequent range expansions left significant correlations among PCO axes and geography, particularly for longitude (Table 2). Relating the distribution of PCO variation for both species between the native and introduced ranges suggests multiple introductions from eastern and western Europe (Fig. 1). In S. latifolia, most genotypes in eastern North America share PCO space with genotypes from central Europe (especially areas of eastern France and Germany), whereas genotypes around the Great Lakes and further west show greater affinity with genotypes from eastern Europe. In S. vulgaris, genotypes from eastern Europe are

found throughout most of the introduced range, whereas western European genotypes are established locally in Pennsylvania, Ontario and Quebec.

Clinal phenotypic evolution

Phenotypic clines with climate and ancestry were apparent in both species and on both continents (Tables 3, 4). In *S. vulgaris*, most traits were best fitted by models that combined climate and ancestry (12 of 16 models; Table 3). The importance of both selective and neutral processes governing genetically based phenotypic variation was indicated by the large number of significant regression coefficients for both climate of origin (P < 0.05 for 17 of 24 estimates for Prin1 and Prin2, collectively) and genetic ancestry (P < 0.05for 13 of 32 estimates for PCO1 and PCO2, collectively).

In Europe, plants from cool, wet climates with low seasonality (large Prin1) were larger (produced more leaves and stems), spent more weeks flowering and produced more flowers and fruits than plants from warmer, drier climates (Table 3). Plants originating from warm, wet environments with low seasonality (large Prin2) were smaller (shorter height and fewer leaves), spent fewer weeks flowering and produced fewer fruits than plants from colder, drier, more seasonal climates (Table 3). Altogether, plants originating from cooler climates (large Prin1, small Prin2) exhibited greater allocation to vegetative growth and reproductive effort than did plants from warmer climates. Clines predicted by latitude and the stepwise selection of climate variables were generally congruent with model selection using PCA. Plants were larger and had greater reproductive effort at higher latitudes and cooler temperatures (Tables S1, S3, see Supporting Information). Effects of precipitation were more variable and depended on the timing of precipitation during the year. Plants from sites with abundant precipitation during the warmest quarter

	Best model	$\Delta \mathrm{AIC}_\mathrm{c}$	Prin1 β	Simple model	Prin2 β	Simple model	ΡΟΟ1 β	ΡϹΟ2 β
Europe								
Germination time	А	7.6					-0.5399	-0.1737
Number of leaves	CA	8.8	0.1027****	\uparrow	-0.1512*		-1.7189**	-1.2979
Number of stems	CA	5.9	0.0819**	\uparrow	-0.0703		-0.7699	-0.5664
Stem height	CA	4.9	0.0254		-0.2347***	\downarrow	-0.0077	-1.3003†
Time to flowering	А	26.2					0.0097	-0.1797*
Weeks flowering	CA	8.8	0.0759****	\uparrow	-0.1057*	\downarrow	-0.2144	-0.9750†
Number of flowers	CA	10.7	0.1478***	\uparrow	-0.1835	\downarrow	0.3481	-2.4726*
Number of fruits	CA	11.4	0.1275**	\uparrow	-0.4667**	\downarrow	-2.3501*	-1.8814
North America								
Germination time	CA	4.8	0.03738		-0.06503**	\downarrow	-1.079****	1.0858**
Number of leaves	CA	1.7	0.0500		0.0868*	↑	-0.2534	-1.1983*
Number of stems	А	3.8		\uparrow			-0.7003†	-0.8398
Stem height	CA	6.0	-0.0076		2.005****	↑	0.5665	-1.0013†
Time to flowering	А	22.7				\downarrow	-0.0016	-0.0376
Weeks flowering	CA	11.4	0.1401**	\uparrow	0.0953***	\uparrow	0.8637**	-1.7084***
Number of flowers	CA	16.3	0.2877**	\uparrow	0.1633*	\uparrow	1.9851**	-3.4684***
Number of fruits	CA	4.2	0.4104****	\uparrow	0.0808		0.7688	0.0797

 Table 3 Evolution of clines in life history traits with climate in Silene vulgaris

Best models were chosen among three candidate models [C (climate), A (ancestry) and CA (climate and ancestry)] based on the lowest AIC_c score. ΔAIC_c reports the difference in fit between the best and next best models. Regression coefficients (β) from best models are given for climate (Prin1 and 2) and ancestry (PCO1 and 2) effects. Large values of Prin1 represent cool, wet climates, whereas large values of Prin2 represent warm, wet climates with low seasonality. Arrows show direction of clines significant under a simple (C only) model. Bold values are significant after sequential Bonferroni correction ($\alpha = 0.05$). t, P < 0.10; *, P < 0.05; **, P < 0.01; ***, P < 0.001; ****, P < 0.0001.

 Table 4
 Evolution of clines in life history traits with climate in Silene latifolia

	Best model	ΔAIC_{c}	Prin1 β	Simple model	Prin2 β	Simple model	ΡΟΟ1 β	ΡϹΟ2 β
Europe								
Basal rosette leaves	А	7.5					-0.0539	0.5664
Stem leaves	А	5.2				\uparrow	-0.8591†	-0.2703
Number of stems	А	3.0				\uparrow	-0.2487	-0.0388
Stem height	CA	0.5	-0.2743*	\downarrow	0.1804*	\uparrow	0.5875	0.7545
Reproductive speed	А	1.5				↑	-2.5177*	-3.2692*
Number of flowers	А	6.2					-0.7720	-0.1272
Number of fruits	А	8.7		\uparrow			0.5962*	-0.0230
North America								
Basal rosette leaves	А	8.4		\uparrow			0.5814†	0.3507
Stem leaves	А	9.1		\uparrow			0.3218	0.2664
Number of stems	А	8.5					0.0701	0.1222
Stem height	А	1.4					-0.1066	-0.2221
Reproductive speed	А	4.3					-1.7919	-0.3949
Number of flowers	А	7.0					-0.2777	0.7940†
Number of fruits	А	7.9					0.2020	0.6180

Best models were chosen among three candidate models [C (climate), A (ancestry) and CA (climate and ancestry)] based on the lowest AIC_c score. ΔAIC_c reports the difference in fit between the best and next best models. Regression coefficients (β) from best models are given for climate (Prin1 and 2) and ancestry (PCO1 and 2) effects. Large values of Prin1 represent cool, wet climates, whereas large values of Prin2 represent warm, wet climates with low seasonality. Arrows show direction of clines significant under a simple (C only) model. Bold values are significant after sequential Bonferroni correction ($\alpha = 0.05$). \dagger , P < 0.01; *, P < 0.05; **, P < 0.01; ***, P < 0.001; ****, P < 0.0001.

produced more leaves, flowered earlier and made more flowers, whereas plants from sites with high precipitation during the rest of the year produced fewer leaves and flowers (Table S3). Strong clines linked to climate have also evolved very recently since the invasion of North America. Plants originating from sites with wet climates (large Prin1, small Prin2) germinated more rapidly, grew taller with more leaves and produced



Fig. 2 Interpretation of phenotypic clines along environmental gradients which change on inclusion of genetic ancestry. Panels show the relationship between leaf number and environment in *Silene vulgaris* (a, b), and the relationship between fruit number and environment in *Silene latifolia* (c, d). (a, c) Phenotypic clines using uncorrected data; (b, d) partial correlations after controlling for the effects of genetic ancestry (PCO1 and PCO2) in the model. Regression lines are shown for the significant relationships (see also Tables 3 and 4).

more flowers and fruits over a longer reproductive period than plants from drier climates (Table 3). Interestingly, reversals between continents in the direction of clines were observed, especially for the warm/wet vs cold/dry climate axis described by Prin2 (Table 3). Reversals in clines were also evident for latitude (four of five clines significant in both ranges reversed; Table S1). North American plants grew smaller and produced fewer flowers when they were from high latitudes, in contrast with plants from Europe. Similarly, stepwise models often included different sets of significant climate predictors in North America relative to Europe (Table S3). When the same climate variable was predictive of a trait in both ranges, the direction of the cline was always reversed (three of three clines reversed).

The inference of clines in S. vulgaris under a simple climate-only model sometimes changed dramatically with the inclusion of ancestry (Table 3). Neutral genetic ancestry among S. vulgaris genotypes was associated with several phenotypic traits in Europe and North America (Table 3). Accounting for ancestry revealed an environmental cline in leaf number that was not previously evident (Fig. 2). European plants originating from cold, dry sites (small Prin2) had significantly greater leaf production (P = 0.0434), but this effect would not have been observed without controlling for ancestry (C model: P = 0.4016). Likewise, multiple traits showing clinal variation under a climate-only model became nonsignificant or were better fitted by a model including ancestry (Table 3). A striking example was flower production among European plants. Under a climate-only model, flower number showed a highly significant relationship with Prin2 (P = 0.0071), which disappeared once ancestry was taken into account (P = 0.1173). Similar results were observed among North American plants for the number of stems and time to flowering, where ancestry-only models provided a better fit to the data than did models that included clinal variation with climate (Table 3).

In contrast with S. vulgaris, most traits in S. latifolia were best explained by a simple ancestry model (13 of 14 models; Table 4). Stem height was an exception, showing significant phenotypic clines in Europe after controlling for ancestry. Taller plants originated from environments that experienced greater precipitation than environments that produced smaller plants (small Prin1, large Prin2). Similar trends were observed with latitude, with most traits explained by ancestryonly models (Table S2, see Supporting Information). Stepwise regression revealed some influence of climate on phenotypic clines, particularly temperature seasonality (Table S4, see Supporting Information). Plants from more seasonal regions were larger (taller with more stem leaves) and faster to reproduce, but made fewer fruits overall. However, none of these relationships remained significant after Bonferroni correction. Clines with climate were generally absent from North America, except for basal leaves, which showed a positive relationship with temperature during the coldest month of the year (Table S4).

In multiple instances, phenotypic clines that were evident in *S. latifolia* under climate-only models were no longer supported after including ancestry (Table 4). For example, among European plants, there was a significant positive relationship between fruit production and Prin1 (P = 0.0350) under a climate-only model. However, an ancestry-only model provided a substantially better fit to the data ($\Delta AIC_c = 8.7$; Fig. 2). Specifically, higher fruit production occurred among genotypes with large PCO1 scores that partially overlapped with the distribution of cool, wet climates in western and central Europe (Figs 1, S1). An ancestry-only model also provided a better fit to clines for stems, stem leaves and reproductive speed (Δ AIC_c from 1.5 to 9.1 for each trait; Table 4).

Discussion

Climate adaptation of *Silene* in Europe and North America

The history of post-glacial expansion and invasion in *S. latifolia* and *S. vulgaris* involved the differentiation of life history traits across geographically varying environments. Because family means for traits were observed across multiple common garden sites, these correlations are probably genetically based. In *S. latifolia*, variation in most traits was best explained by the genetic ancestry of families. In *S. vulgaris*, much of the phenotypic divergence occurred along the major axes of temperature and precipitation variation across Europe and North America. As most of these clines were significant after controlling for genetic ancestry, divergence in *S. vulgaris* probably reflects an evolutionary response to selection imposed by regional climate conditions.

Clines in size and growth traits often paralleled clines in reproductive traits. In Europe, S. vulgaris families from Mediterranean climates (hot, dry summers) were smaller and reproduced less than families from more temperate regions of northern Europe. In North America, families from the wetter eastern half of the continent were larger and had higher reproductive output than families from drier sites in western North America. Although tradeoffs between growth and reproduction are expected when resource acquisition is fixed, variance in resource acquisition leads to the evolution of positive correlations between growth and reproduction (Stearns, 1989). One hypothesis is that genotypes from Mediterranean Europe and western North America occupy marginal regions of the fundamental niche, and have evolved stress tolerance (for example, to heat and water availability) that limits the physiological function achievable under more permissive conditions. Ecological niche models for S. vulgaris suggest that this may be the case (S. Keller & D. Taylor, unpublished).

The evolution of phenotypic clines among North American *S. vulgaris* contributes to growing evidence that evolution can be pronounced and rapid following invasion (Blair & Wolfe, 2004; Lavergne & Molofsky, 2007; Facon *et al.*, 2008). The establishment of clines is a feature of many species invasions and has been interpreted as a hallmark example of adaptive evolution in action (Johnston & Selander, 1964; Lacey, 1988; Weber & Schmid, 1998; Huey *et al.*, 2000; Kollmann & Banuelos, 2004; Maron *et al.*, 2004, 2007; Leger & Rice, 2007; Etterson *et al.*, 2008; Montague *et al.*, 2008). In our study, divergence was often observed in excess of the neutral

expectation generated by ancestry, offering particularly strong evidence that the clines are adaptive. Complementary approaches, such as reciprocal transplant experiments and functional assays, would help to further the case for the adaptive nature of the clines, as well as help to reveal their mechanistic basis.

Clinal variation is often expressed as correlation between phenotypes and environmental variation or geographical variation that covaries with environment (for example, latitude or altitude). We observed clines with climate and, as expected, climate covaried with geography to generate geographical clines as well. Interestingly, environmental variation often varied in opposite geographical directions on the two continents (Table 2), resulting in a clear reversal of the phenotypic clines documented in our study. Reversals of cline direction were especially evident along the Prin2 climate axis (Table 3) and latitude (Table S1). This pattern might have been misinterpreted to reflect different colonization histories had we not corrected for genetic ancestry, as the PCO axes showed significant correlations with latitude and longitude that differed in sign between ranges (Table 2). However, most clines maintained their reversal in direction, even after correcting for ancestry (Fig. 3), pointing towards differences in climateimposed selection occurring within each range. This result is not wholly surprising, given that most of North America is characterized by a continental climate, whereas much of western and central Europe is moderated by the influence of the Gulf Stream (Menzel et al., 2005). Silene in southern Europe (vicinity of the Mediterranean) often seems to restrict its flowering with the onset of summer heat, whereas flowering in North America starts late and can be truncated early whilst plants are still producing flowers by the onset of cold temperatures (S. R. Keller et al., pers. obs.). These results suggest that unique patterns of selection may promote divergence between ranges as introduced populations adapt to novel climate conditions.

As seeds of *S. vulgaris* were field collected from open pollinated fruits, it is possible that some of the phenotypic variation observed could be attributable to maternal effects. Maternal effects in plants are principally manifested in early life history traits, especially seeds, and have diminishing influence beyond the seedling stage (Roach & Wulff, 1987). Only one of the traits measured in *S. vulgaris*, germination time, is expressed early in the life history, whereas all other traits are properties of adult plants. However, the cline in germination time with Prin2, observed for North American *S. vulgaris*, could be influenced by the environment in which the seeds developed (Table 3), and should therefore be interpreted cautiously.

Parsing adaptive and nonadaptive origins of clinal variation

Although it has long been appreciated that plant populations are capable of rapid adaptive evolution along strong



Fig. 3 Reversal of latitudinal clines for fruit production of *Silene vulgaris* in Europe (a) vs North America (b). Fruit production is expressed as residuals from a model that removes the effects of genetic ancestry (PCO1 and PCO2).

environmental gradients (for example, Antonovics & Bradshaw, 1970), the novelty of our study comes from a broad consideration of the evolutionary processes that may act during nonequilibrium conditions, such as range expansion and invasion. Divergence during expansion will often reflect a history of spatially distinct dispersal and establishment, founder effects, genetic drift and gene flow. For this reason, recent studies on the evolution of clines are now beginning to incorporate null models based on genetic ancestry into tests for adaptation (Samis *et al.*, 2008). The effect of selectively neutral demographic processes on divergence is highly acute during species invasions. Invasions frequently involve multiple introductions of genetically distinct lineages sampled from different portions of the native range and established at different points within the introduced range (see reviews in Bossdorf *et al.*, 2005; Novak & Mack, 2005; Wares *et al.*, 2005; Roman & Darling, 2007; Dlugosch & Parker, 2008). The complexity inherent in many introduction histories means that controlling for ancestry effects is of utmost importance when testing for evolution in invasive species (Keller & Taylor, 2008).

We found strong correlations between neutral genetic variance and gradients in geography and climate in both the native and introduced ranges of Silene. Because neutral variance covaries with phenotype, and is nonrandomly distributed in space, ancestry affects how we interpret the causal processes affecting phenotypic clines. Our interpretation of adaptively evolved clines relied on the pairing of phenotypic and molecular data on the same family lines, which allowed us to test for phenotypic divergence relative to a neutral expectation (Keller & Taylor, 2008). A few previous studies of phenotypic clines during invasions recognized the need for the consideration of alternative mechanisms of divergence, such as colonization history (Lacey, 1988; Maron et al., 2004; Montague et al., 2008), but ours is the first to explicitly control for the influence of founding events, drift and migration in the generation of clines. Controlling for demographic processes significantly altered our interpretation of the processes of evolution in several cases. For some traits, significant clines could be fully accounted for by shared history at neutral loci (for example, number of flowers in European S. vulgaris and number of fruits in European S. latifolia; Tables 3, 4; Fig. 2). In other traits, accounting for demographic and historical effects revealed support for adaptive differentiation that was not otherwise evident (for example, number of leaves in European S. vulgaris; Fig. 2).

How conservative are these tests of clinal adaptation? It is possible for putatively neutral loci to diverge via hitchhiking when they are closely linked to loci under selection. Therefore, recent and/or strong selection could generate linkage disequilibrium that sets up clines even at neutral loci. Any effect of hitchhiking would tend to make our test for adaptation more conservative, as some of the putatively neutral variance controlled for may be linked to regions under selection. Our approach should be complementary to other methods of inferring robust evidence for clinal evolution, such as the observation of coincident parallel clines reported for several species invasions (Huey *et al.*, 2000; Leger & Rice, 2007; Maron *et al.*, 2007). Our method could be especially useful in this context, by controlling for colonization history that might otherwise obscure a parallel cline.

Species differences in clinal differentiation

Phenotypic clines were observed more frequently in *S. vulgaris* than in *S. latifolia*. As the *S. vulgaris* experiment involved more families, part of this difference probably reflects differences in statistical power between the two field experiments. To explore this possibility, we used a power analysis to

estimate the likelihood of detecting significant phenotypic clines in S. latifolia relative to S. vulgaris. For each trait, we computed the partial correlation coefficient $(r_{\rm D})$ with one climate Prin axis, whilst controlling for the other Prin axis and both PCO axes, and computed the power to reject a false null hypothesis based on the sample size in each species on each continent (PROC POWER: SAS, 2004). The mean effect size did not differ between species (mean absolute value of $r_{\rm p}$: *S. latifolia*, 0.205; *S. vulgaris*, 0.239; *t* = -0.994; *P* = 0.3242). As expected, the difference in sample size between S. latifolia and S. vulgaris leads to differences in statistical power. Given the effect sizes observed, we had 4.4 times as much power to reject the null hypothesis in S. vulgaris relative to S. latifolia. However, we also note that our process of model selection relied only on how well different predictors explained the data. In this regard, C and A models should have been selected at about the same frequency, all else being equal, as they possessed the same sample sizes and number of parameters. Thus, the large number of traits favouring a neutral ancestry model in S. latifolia vs the predominance of models including environmental clines in S. vulgaris might reflect a genuine difference in biology between the species. It is also possible that the climate variation summarized by the PCA axes did not capture the selective gradients important to S. latifolia. When stepwise regression was used, there was more support for clines with climate, although none of these was significant after Bonferroni correction (Table S4). Interestingly, most of the evidence for phenotypic clines with climate was in Europe, with very few variables approaching significance in North America. Previous studies in S. latifolia have reported divergence in life history traits among European populations (Delph et al., 2002; Wolfe et al., 2004), and one trait (age at first flowering) has previously been shown to exhibit clinal variation with latitude (Jolivet & Bernasconi, 2007). There is also phenotypic divergence between Europe and North America consistent with adaptive evolution, although not expressly clinal in nature (Blair & Wolfe, 2004; Wolfe et al., 2004). Thus, although there is some evidence that life history traits vary clinally in the native range, adaptive evolution along regional environmental gradients in the introduced range appears to be less prominent relative to neutral divergence or other selective processes. Of course, it is also possible that S. latifolia is adapting along environmental gradients in North America not included in these analyses.

Conclusion

Phenotypic covariance between traits and the environment can arise from nonadaptive processes, and a complete understanding of adaptive differentiation – or lack thereof – requires that these processes are taken into account. We observed clines in the native and introduced ranges of two weedy plants that have a global history of post-glacial expansion and invasion. Many of these clines were robust to the inclusion of information on genetic ancestry, strongly supporting the inference of adaptive evolution within each range in response to regionally varying climate, whereas other clines were consistent with neutral genetic processes. Studies of phenotypic evolution during range expansions, species invasions and other situations characterized by nonequilibrium demographic and genetic conditions will benefit from the incorporation of a neutral expectation into tests of evolutionary change.

Acknowledgements

We thank R. Neville, N. Rothwell and C. Smith (Mountain Lake Biological Station and Queen's University Biological Station) for permission and assistance in establishing the common gardens. We also appreciate the contributions of numerous individuals who donated seeds or helped with data collection at the garden sites. We thank Ruth Shaw and three reviewers for providing constructive criticism that improved the manuscript. This work was supported by National Science Foundation (NSF) awards DEB 0349558 and DEB 0608358.

References

- Amsellem L, Noyer JL, Le Bourgeois T, Hossaert-McKey M. 2000. Comparison of genetic diversity of the invasive weed *Rubus alceifolius* Poir. (Rosaceae) in its native range and in areas of introduction, using amplified fragment length polymorphism (AFLP) markers. *Molecular Ecology* 9: 443–455.
- Antonovics J, Bradshaw AD. 1970. Evolution in closely adjacent plant populations VIII. Clinal patterns at a mine boundary. *Heredity* 25: 349–362.
- Baker HG. 1948. Stages in invasion and replacement demonstrated by species of *Melandrium. Journal of Ecology* 36: 96–119.
- Blackburn T, Gaston KJ, Loder N. 1999. Geographic gradients in body size: a clarification of Bergmann's Rule. *Diversity and Distributions* 5: 165–174.
- Blair AC, Wolfe LM. 2004. The evolution of an invasive plant: an experimental study with *Silene latifolia*. *Ecology* **85**: 3035–3042.
- Bossdorf O, Auge H, Lafuma L, Rogers WE, Siemann E, Prati D. 2005. Phenotypic and genetic differentiation between native and introduced plant populations. *Oecologia* 144: 1–11.
- Cavalli-Sforza LL, Menozzi P, Piazza A. 1993. Demic expansions and human evolution. *Science* 259: 639–646.
- Chang K-T. 2008. Introduction to geographic information systems. Singapore: McGraw-Hill.
- Clausen J, Keck DD, Hiesey WM. 1948. Experimental studies on the nature of species III. In: *Carnegie Institute of Washington Publication 581*. Washington DC, USA: Carnegie Institute of Washington.
- Clegg SM, Degnan SM, Kikkawa J, Moritz C, Estoup A, Owens IPF. 2002. Genetic consequences of sequential founder events by an island-colonizing bird. *Proceedings of the National Academy of Sciences, USA* 99: 8127–8132.
- Delph LF, Knapczyk FN, Taylor DR. 2002. Among-population variation and correlations in sexually dimorphic traits of *Silene latifolia*. *Journal of Evolutionary Biology* 15: 1011–1020.
- Dlugosch KM, Parker IM. 2008. Founding events in species invasions: genetic variation, adaptive evolution, and the role of multiple introductions. *Molecular Ecology* 17: 431–449.
- Drezner TD. 2003. Revisiting Bergmann's rule for saguaros (*Carnegiea gigantea* (Engelm.) Britt. and Rose): stem diameter patterns over space. *Journal of Biogeography* 30: 353–359.

Eanes WF. 1999. Analysis of selection on enzyme polymorphisms. Annual Review of Ecology and Systematics 30: 301–326.

Endler J. 1977. *Geographic variation, speciation, and clines*. Princeton, NJ, USA: Princeton University Press.

Etterson JR, Delf DE, Craig TP, Ando Y, Ohgushi T. 2008. Parallel patterns of clinal variation in *Solidago altissima* in its native range in central USA and its invasive range in Japan. *Botany-Botanique* 86: 91–97.

Facon B, Pointier J-P, Jarne P, Sarda V, David P. 2008. High genetic variance in life-history strategies within invasive populations by way of multiple introductions. *Current Biology* 18: 363–367.

Glaettli M, Pescatore L, Goudet J. 2006. Proximity-dependent pollen performance in *Silene vulgaris. Annals of Botany* 98: 431–437.

Gould SJ, Lewontin RC. 1979. The spandrels of San Marco and the Panglossian paradigm: a critique of the adaptationist programme. *Proceedings of the Royal Society of London Series B* **205**: 581–598.

Hewitt G. 2000. The genetic legacy of the Quaternary ice ages. *Nature* 405: 907–913.

Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A. 2005. Very high resolution interpolated climate surfaces for global land areas. *Journal of Climatology* 25: 1965–1978.

Huey RB, Gilchrist GW, Carlson ML, Berrigan D, Serra L. 2000. Rapid evolution of a geographic cline in size in an introduced fly. *Science* 287: 308–309.

Ingvarsson P, Garcia MV, Hall D, Luquez V, Jansson S. 2006. Clinal variation in phyB2, a candidate gene for daylength-induced growth cessation and bud set, across a latitudinal gradient in European Aspen (*Populus tremula*). *Genetics* 172: 1845–1853.

Johnson JB, Omland KS. 2004. Model selection in ecology and evolution. Trends in Ecology & Evolution 19: 101–108.

Johnston RF, Selander RK. 1964. House sparrows: rapid evolution of races in North America. *Science* 144: 548–550.

Jolivet C, Bernasconi G. 2007. Molecular and quantitative genetic differentiation in European populations of *Silene latifolia* (Caryophyllaceae). *Annals of Botany* 100: 119–127.

Keller SR. 2008. *History, chance, and adaptation: the evolution of* Silene vulgaris *in its native and introduced ranges.* PhD Thesis. University of Virginia, Charlottesville, USA.

Keller SR, Taylor DR. 2008. History, chance and adaptation during biological invasion: separating stochastic phenotypic evolution from response to selection. *Ecology Letters* 11: 852–866.

Kimura M. 1980. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution* 16: 111–120.

Kollmann J, Banuelos MJ. 2004. Latitudinal trends in growth and phenology of the invasive alien plant *Impatiens glandulifera* (Balsaminaceae). *Diversity and Distributions* 10: 377–385.

Lacey EP. 1988. Latitudinal variation in reproductive timing of a short-lived monocarp, *Daucus carota* (Apiaceae). *Ecology* 69: 220–232.

Lavergne S, Molofsky J. 2007. Increased genetic variation and evolutionary potential drive the success of an invasive grass. *Proceedings of the National Academy of Sciences, USA* 104: 3883–3888.

Leger EA, Rice KJ. 2007. Assessing the speed and predictability of local adaptation in invasive California poppies (*Eschscholzia californica*). *Journal of Evolutionary Biology* 20: 1090–1103.

Lewontin RC, Krakauer J. 1973. Distribution of gene frequency as a test of theory of selective neutrality of polymorphisms. *Genetics* 74: 175–195.

Maron JL, Elmendorf SC, Vila M. 2007. Contrasting plant physiological adaptation to climate in the native and introduced range of *Hypericum perforatum*. *Evolution* **61**: 1912–1924.

Maron JL, Vila M, Bommarco R, Elmendorf S, Beardsley P. 2004. Rapid evolution of an invasive plant. *Ecological Monographs* 74: 261–280.

Martindale IC. 1876. The introduction of foreign plants. *Botanical Gazette* 2: 55–58.

Mastenbroek O, Prentice HC, Heringa J, Hogeweg P. 1983. Corresponding patterns of geographic variation among populations of *Silene latifolia*

(= S. alba = S. pratensis) (Caryophyllaceae). Plant Systematics and Evolution 145: 227–242.

McCauley DE, Raveill J, Antonovics J. 1995. Local founding events as determinants of genetic structure in a plan metapopulation. *Heredity* 75: 630–636.

McNeill J. 1977. The biology of Canadian weeds. 25. Silene alba (Miller) E.H.L. Krause. Canadian Journal of Plant Science 57: 1103–1114.

Menzel A, Sparks TH, Estrella N, Eckhardt S. 2005. 'SSW to NNE' – North Atlantic Oscillation affects the progress of seasons across Europe. *Global Change Biology* 11: 909–918.

Montague JL, Barrett SCH, Eckert CG. 2009. Re-establishment of clinal variation in flowering time among introduced populations of purple loosestrife (*Lythrum salicaria*, Lythraceae). *Journal of Evolutionary Biology* 21: 234–245.

Novak SJ, Mack RN. 2005. Genetic bottlenecks in alien plant species: influence of mating systems and introduction dynamics. In: Sax DF, Stachowicz JJ, Gaines SD, eds. *Species invasions: insights into ecology, evolution, and biogeography.* Sunderland, MA, USA: Sinauer Associates, Inc., 201–228.

Petit RJ, Aguinagalde I, de Beaulieu J-L, Bittkau C, Brewer S, Cheddadi R, Ennos R, Fineschi S, Grivet D, Lascoux M et al. 2003. Glacial refugia: hotspots but not melting pots of genetic diversity. Science 300: 1563–1565.

Price AL, Patterson NJ, Plenge RM, Weinblatt ME, Shadick NA, Reich D. 2006. Principal components analysis corrects for stratification in genome-wide association studies. *Nature Genetics* 38: 904–909.

Randall RP. 2002. A global compendium of weeds. Melbourne, Australia: R.G. and F.J. Richardson.

Roach DA, Wulff RD. 1987. Maternal effects in plants. Annual Review of Ecology and Systematics 18: 209–235.

Roman J. 2006. Diluting the founder effect: cryptic invasions expand a marine invader's range. *Proceedings of the Royal Society B: Biological Sciences* 273: 2453–2459.

Roman J, Darling JA. 2007. Paradox lost: genetic diversity and the success of aquatic invasions. *Trends in Ecology & Evolution* 22: 454–464.

Samis KE, Heath KD, Stinchcombe JR. 2008. Discordant longitudinal clines in flowering time and *phytochrome C* in *Arabidopsis thaliana*. *Evolution* 62: 2971–2983.

SAS. 2004. SAS 9.1.3. Cary, NC, USA: SAS Institute.

Savolainen O, Pyhajarvi T, Knurr T. 2007. Gene flow and local adaptation in trees. Annual Review of Ecology, Evolution and Systematics 38: 595–619.

Scholander PF. 1955. Evolution of climatic adaptation in homeotherms. *Evolution* 9: 15–26.

Spitze K. 1993. Population-structure in *Daphnia obtusa* – quantitative genetic and allozyme variation. *Genetics* 135: 367–374.

Stearns SC. 1989. Trade-offs in life-history evolution. *Functional Ecology* 3: 259–268.

Stinchcombe JR, Weinig C, Ungerer M, Olsen KM, Mays C, Halldorsdottir SS, Purugganan MD, Schmitt J. 2004. A latitudinal cline in flowering time in *Arabidopsis thaliana* modulated by the flowering time gene FRIGIDA. *Proceedings of the National Academy of Sciences, USA* 101: 4712–4717.

Taylor DR, Keller SR. 2007. Historical range expansion determines the phylogenetic diversity introduced during contemporary species invasion. *Evolution* **61**: 334–345.

Vasemagi A. 2006. The adaptive hypothesis of clinal variation revisited: single-locus clines as a result of spatially restricted gene flow. *Genetics* 173: 2411–2414.

Wares JP, Hughes AR, Grosberg RK. 2005. Mechanisms that drive evolutionary change: insights from species introductions and invasions. In: Sax DF, Stachowicz JJ, Gaines SD, eds. *Species invasions: insights into ecology, evolution, and biogeography.* Sunderland, MA, USA: Sinauer Associates, Inc., 229–257.

- Weber E, Schmid B. 1998. Latitudinal population differentiation in two species of *Solidago* (Asteraceae) introduced to Europe. *American Journal of Botany* 85: 1110–1121.
- Whitlock MC, McCauley DE. 1990. Some population genetic consequences of colony formation and extinction: genetic correlations within founding groups. *Evolution* 44: 1717–1724.
- Whitlock MC, McCauley DE. 1999. Indirect measures of gene flow and migration: $F_{ST} \neq 1/(4Nm + 1)$. *Heredity* 82: 117–125.
- Williams JL, Auge H, Maron JL. 2008. Different gardens, different results: native and introduced populations exhibit contrasting phenotypes across common gardens. *Oecologia* 157: 239–248.
- Wolfe LM, Elzinga JA, Biere A. 2004. Increased susceptibility to enemies following introduction in the invasive plant *Silene latifolia*. *Ecology Letters* 7: 813–820.

Supporting Information

Additional supporting information may be found in the online version of this article.

Fig. S1 Distribution of WorldClim climate variation for *Silene latifolia* and *Silene vulgaris* sites in Europe and North America.

Table S1 Evolution of clines in life history traits with latitudein Silene vulgaris

Table S2 Evolution of clines in life history traits with latitudein Silene latifolia

Table S3 Stepwise regression of phenotypic clines predicted by ancestry (principal coordinates analyses, PCOs) and climate for *Silene vulgaris*

Table S4 Stepwise regression of phenotypic clines predicted by ancestry (principal coordinates analyses, PCOs) and climate for *Silene latifolia*

Please note: Wiley-Blackwell are not responsible for the content or functionality of any supporting information supplied by the authors. Any queries (other than missing material) should be directed to the *New Phytologist* Central Office.

	Best Model	ΔAIC_{c}	Latitude ^β	Simple model	PCO1 β	PCO2 β
Europe						
Germination time	А	8.0			-0.5399	-0.1737
Number of leaves	LA	3.4	0.0508^{****}	↑	-0.5086	-0.3553
Number of stems	А	1.0		↑	-0.7339	-0.5563
Stem height	А	4.1			-1.2984**	- 1.9686 ^{†*}
Time to flowering	L	3.1	0.0047^{****}	↑		
Weeks flowering	LA	9.0	0.0374****	↑	0.6231 [†]	-0.1218
Number of flowers	LA	12.9	0.0723****	↑	1.7930*	-0.7148
Number of fruits	LA	2.4	0.0540**	↑	0.8177	-1.4829
North America						
Germination time	А	6.5			-1.079****	1.0175**
Number of leaves	А	1.5		\downarrow	-0.3377	-1.3517*
Number of stems	А	7.3			-0.7003 [†]	-0.8398
Stem height	LA	6.3	-0.0625***	\downarrow	0.7267^{\dagger}	-0.7424
Time to flowering	LA	7.3	0.0022^{*}	↑	-0.0061	-0.0576
Weeks flowering	LA	7.1	-0.0336****	\downarrow	0.7051*	-1.8042****
Number of flowers	LA	3.8	-0.0607**	\downarrow	1.6228*	-3.6797***
Number of fruits	А	2.9		[↓]	0.1678	-0.4768

Table S1. Evolution of clines in life history traits with latitude in *Silene vulgaris*.

Best models were chosen among three candidate models: L (latitude), A (ancestry), and LA (latitude and ancestry) based on the lowest AIC_c score. Δ AIC_c reports the difference in fit between the best and next best models. Regression coefficients (β) from best models are given for climate (Prin1 and 2) and ancestry (PCO1 and 2) effects. Arrows show direction of clines significant under a simple (C only) model. [†]*P*<0.10, ^{*}*P*<0.05, ^{**}*P*<0.01, ^{****}*P*<0.001

	Best Model	ΔAIC_{c}	Latitude β	Simple model	PCO1 β	PCO2 β
Europe						
Basal rosette leaves	А	5.2			-0.0539	0.5663
Stem leaves	А	3.3			-0.8591 [†]	-0.2611
Number of stems	А	5.0			-0.2487	-0.0388
Stem height	А	4.9			-0.6172 [†]	0.1380
Reproductive speed	А	1.5			- 2.5177 [*]	-3.2692*
Number of flowers	А	5.1			-0.7722	-0.1272
Number of fruits	А	5.4			0.5962^{*}	-0.0230
North America						
Basal rosette leaves	А	6.2			0.5814^{\dagger}	0.3507
Stem leaves	А	3.4			0.3218	0.2664
Number of stems	L	1.3		[↓]		
Stem height	А	0.2			0.3766	0.2344
Reproductive speed	А	4.3			-1.7919	-0.3949
Number of flowers	А	7.0			-0.2777	0.7940^{\dagger}
Number of fruits	А	6.3			0.2020	0.6180

Table S2. Evolution of clines in life history traits with latitude in *Silene latifolia* (see footnote in Table S1 for details).

⁺*P*<0.10, **P*<0.05, ***P*<0.01, *****P*<0.001, *****P*<0.0001

Europe					North America				
Model	R ²	β	F	Р	Model	R ²	β	F	Р
Germination time	0.024					0.412			
PCO1		-0.53838	2.17	0.1438	PCO1		-1.07523	23.95	<0.0001
					PCO2		1.20229	13.22	0.0005
					AVGANNTMP		0.00446	3.55	0.0633
					MINTMPCLDMNTH		-0.00513	8.48	0.0047
Number of leaves	0.343					0.182			
PCO1		-1.21141	3.98	0.0493	PCO2		-1.03467	3.62	0.0609
AVGANNTMP		-0.00785	8.53	0.0045	AVGANNPCP		0.00048	3.55	0.0631
PCPDRYMNTH		-0.01253	10.21	0.0020	MAXTMPWRMMNTH		0.00333	2.97	0.0891
PCPSEASON		-0.01142	10.31	0.0019					
PCPWRMQTR		0.00165	2.65	0.1071					
Number of stems	0.171					0.102			
MAXTMPWRMMNTH		-0.00557	18.10	<0.0001	AVGANNPCP		0.00055	5.27	0.0244
					TMPSEASON		0.00009	4.67	0.0339
Stem height	0.330					0.283			
AVGANNPCP		-0.00076	29.04	<0.0001	PCO2		-1.16050	3.85	0.0534
MINTMPCLDMNTH		-0.00651	24.39	<0.0001	MINTMPCLDMNTH		0.00575	26.05	<0.0001
Time to flowering	0.302					0.074			
AVGANNTMP		0.00227	12.93	0.0005	MINTMPCLDMNTH		-0.00024	6.27	0.0143
PCPSEASON		-0.00125	15.73	0.0002					
PCPWRMQTR		-0.00012	2.45	0.1212					
TMPSEASON		0.00005	27.97	<0.0001					

Table S3. Stepwise multiple regression estimates of phenotypic clines predicted by ancestry (PCOs) and climate for *Silene vulgaris*.

	-0.00208	18.59	<0.0001					
0.344					0.472			
	-0.00060	20.42	<0.0001	PCO1		0.99331	12.37	0.0007
	0.00250	45.58	<0.0001	PCO2		-1.38248	10.21	0.0020
				AVGANNPCP		0.00166	9.61	0.0027
				PCPDRYMNTH		-0.01016	2.54	0.1151
				MAXTMPWRMMNTH		0.00355	4.82	0.0312
0.323					0.375			
	-2.26626	4.24	0.0427	PCO1		2.07806	10.33	0.0019
	-0.01785	13.05	0.0005	PCO2		-3.23411	11.16	0.0013
	0.00670	28.62	<0.0001	AVGANNPCP		0.00191	19.96	<0.0001
0.275					0.203			
	-0.01891	27.11	<0.0001	AVGANNPCP		0.00193	18.77	<0.0001
	-0.00139	20.29	<0.0001	TMPSEASON		0.00012	2.66	0.1070
	0.344 0.323 0.275	-0.00208 0.344 -0.00060 0.00250 0.00250 0.00250 -2.26626 -0.01785 0.00670 0.275 -0.01891 -0.00139	$\begin{array}{c} -0.00208 & 18.59 \\ 0.344 \\ -0.00060 & 20.42 \\ 0.00250 & 45.58 \end{array}$ $\begin{array}{c} 0.323 \\ -2.26626 & 4.24 \\ -0.01785 & 13.05 \\ 0.00670 & 28.62 \end{array}$ $\begin{array}{c} 0.275 \\ -0.01891 & 27.11 \\ -0.00139 & 20.29 \end{array}$	$\begin{array}{c} -0.00208 & 18.59 & <0.0001 \\ 0.344 \\ -0.00060 & 20.42 & <0.0001 \\ 0.00250 & 45.58 & <0.0001 \end{array}$ $\begin{array}{c} 0.323 \\ -2.26626 & 4.24 & 0.0427 \\ -0.01785 & 13.05 & 0.0005 \\ 0.00670 & 28.62 & <0.0001 \end{array}$ $\begin{array}{c} 0.275 \\ -0.01891 & 27.11 & <0.0001 \\ -0.00139 & 20.29 & <0.0001 \end{array}$	-0.00208 18.59 <0.0001 0.344 -0.00060 20.42 <0.0001 PCO1 0.00250 45.58 <0.0001 PCO2 AVGANNPCP PCPDRYMNTH MAXTMPWRMMNTH 0.323 -2.26626 4.24 0.0427 PCO1 -0.01785 13.05 0.0005 PCO2 0.00670 28.62 <0.0001 PCO2 AVGANNPCP PCO1 PCO2 AVGANNPCP PCO1 PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP PCO2 AVGANNPCP	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

The *P*-value to enter or exit the model was set to 0.15. Bold values are significant after sequential Bonferroni correction. Worldclim climate variables: AVGANNTMP (average annual temperature), MAXTMPWRMMNTH (maximum temperature during the warmest month of the year), MINTMPCLDMNTH (minimum temperature during the coldest month of the year), TMPSEASON (temperature seasonality), AVGANNPCP (average annual precipitation), PCPDRYMNTH (average precipitation during the driest month of the year), PCPWRMQTR (average precipitation during the warmest quarter of the year), PCPSEASON (precipitation seasonality).

	Eur	ope			North America					
Model	R ²	β	F	Р	Model	R ²	В	F	Р	
Basal rosette leaves	0.251					0.385				
PCO2		0.67632	4.75	0.0421	PCPWRMQTR		-0.00095	2.49	0.1323	
PCPSEASON		-0.00745	3.01	0.099	TMPSEASON		0.00014	4.92	0.0396	
					MINTMPCLDMNTH		0.00575	8.87	0.0081	
Stem leaves	0.427					0.146				
PCPWRMQTR		0.00199	3.64	0.0727	MINTMPCLDMNTH		0.00108	3.43	0.0789	
TMPSEASON		0.00031	10.08	0.0052						
MINTMPCLDMNTH		0.00851	5.83	0.0266						
Number of stems	0.414					0.107				
PCO1		-0.46767	6.82	0.0177	AVGANNTMP		0.00106	2.4	0.1368	
AVGANNTMP		0.00264	3.91	0.0636						
PCPDRYMNTH		0.00415	9.19	0.0072						
Stem height	0.394					0.146				
PCO1		1.18025	2.32	0.1447	AVGANNPCP		0.00076	3.42	0.0793	
PCPSEASON		0.01731	4.53	0.0474						
TMPSEASON		0.00034	8.81	0.0082						
Reproductive speed	0.629					0.124				
PCO2		-3.39764	11.14	0.0037	PCO1		-1.65399	2.84	0.1075	
AVGANNTMP		0.01409	3.24	0.0888						
TMPSEASON		0.00061	14.27	0.0014						
Number of flowers	0.259					0.343				
PCPSEASON		0.01523	5.43	0.0309	PCO2		1.38544	9.93	0.0053	

Table S4. Stepwise multiple regression estimates of phenotypic clines predicted by ancestry (PCOs) and climate for *Silene latifolia*.

PCPWRMQTR		0.00169	2.89	0.1054	AVGANNPCP	0.00032	2.85	0.1079
Number of fruits TMPSEASON	0.274	-0.00011	7.18	0.0148	(none)			

See legend for Table S3 for details.



Fig. S1. Distribution of WorldClim climate variation for *S. latifolia* and *S. vulgaris* sites in Europe and North America. Colors represent interpolated contours in climate (inverse distance weighting method) as summarized by the first two principal components (Prin1 and Prin2) obtained from eight WorldClim climate variables. Black circles are locations of *S. latifolia* and *S. vulgaris* used this study. Cooler colors correspond to larger values of Prin1 and Prin2; warmer colors correspond to smaller values.